ERSAF
ENTE REGIONALE PER I SERVIZI ALL'AGRICOLTURA E ALLE FORESTE

Final Report of the phytosanitary monitoring plan of the exhibition site EXPO 2015

Regione Lombardia

Final Report of the phytosanitary monitoring plan of the exhibition site EXPO 2015

Co-ordinated by:

Mariangela Ciampitti – ERSAF

Curated by:

Francesca Siena – ERSAF
Chiara Patti – ERSAF
Emma Zanotti – ERSAF

In cooperation with:

Beniamino Cavagna – Regione Lombardia
Andrea Tantardini – Fondazione Minoprio
Andrea Taddei – Fondazione Minoprio
Francesca Gaffuri – Fondazione Minoprio
Marica Calvi – Fondazione Minoprio
Stefano Sacchi – Fondazione Minoprio

Map cover and graphic editing: Chiara Patti

December 2016

Regione Lombardia
Plant Protection Service

EXPO MILANO 2015

Introductory Remarks

Movements of people and goods between countries and continents is constantly increasing at an exponential pace, and this also raises the risk of the introduction of invasive alien species. In recent years there has been an increase of attention to this topic at the international level, thanks to the recognition of the progressively greater impact that these species are having on biodiversity and food security.

The economic damage inflicted by the introduction of invasive alien species noxious to plants has been quantified in billions of dollars per year for America alone, and additional billions of euros In Europe. In poor countries, the repercussions include an insufficient food supply.

The Lombardy region has always been attentive to these issues: it is evident from the fact that over the past 10 years, Lombardy has invested over 20 million euros in the fight against the Asiatic longhorn beetle, the chestnut gall wasp and the Japanese beetle (*Popillia japonica*).

Once these organisms have been introduced and have colonized a new habitat, it takes an enormous effort to prevent their spread. The successful outcome of such efforts depends to no small degree on the early identification of specimens introduced accidentally, and this can only be achieved by implementing specific monitoring plans.

In 2015, Milan hosted EXPO2015, the 'world's fair'', with 140 participating countries and more than 20 million visitors, many of whom were from outside Italy. The "feed the planet" theme and the special attention devoted to topics addressing food and biodiversity made the event even more vulnerable to the accidental introduction of new harmful organisms, because the pavilions and booths were chock-full of a vast variety of plants, seeds and fruit from all over the world.

The regional Phytosanitary Service, in collaboration with EXPO2015, therefore designed and implemented a detailed monitoring plan intended to identify invasive alien species that posed a threat to plants. The planning stage involved a risk analysis, with crosschecking of data concerning the types of plants and vegetable matter introduced into the exhibition grounds and international reports of findings. The validity of the identification of targets to search for was subsequently confirmed by the European Commission, which added many of the organisms monitored during EXPO2015 to the monitoring programs that will be carried out by all the member countries in 2017.

The monitoring performed by ERSAF on the occasion of EXPO2015 was the first example of managing phytosanitary risk at a large-scale event. The publication of this report is intended to stimulate increased attention to prevention projects, so as to avoid having to later manage emergencies with a heavy economic and environmental cost.

Elisabetta Parravicini

President of ERSAF

FINAL REPORT ON THE PHYTOSANITARY MONITORING PLAN FOR THE EXHIBITION GROUNDS OF EXPO 2015

Foreword

Like humans and animals, plants are also subject to illness. They can be infected by fungi, viruses, bacteria, acari, insects and nematodes. The law protects plant health and the wellbeing of forests and the environment, and prohibits the transport of numerous plants and plant products without official authorization. But despite the controls conducted by the competent agencies, the number of new parasites and illnesses introduced into areas where they had previously been absent has increased considerably in recent years. The causes certainly include an increase in commerce and international travel, in addition to the altered environmental conditions resulting from climate change, which favor the spread and establishment of populations of many harmful species. Parasites and diseases can be concealed inside plants, seeds, fruits and vegetables and flowers. These new harmful organisms, which pose a threat to the economy and to biodiversity, are defined as 'alien', because they originate in environments other than the ones in which man has introduced them. Some do not survive, but others manage to adapt themselves all too well to their new habitat; the latter, extraneous to the ecosystem, are termed 'invasive'. Alien species often come into conflict with local species, and if the competition becomes fierce, it is often the latter who have the worst of it.

In recent years, Lombardy has seen the arrival and spread, via the introduction of infested plants, of some particularly noxious organisms, such as the Asiatic longhorn beetle, the chestnut gall wasp and the bacteria that cause fireblight. Damage to the Lombard environment has been heavy, with over 25,000 ornamental trees felled to block the spread of the borer beetle, the Asianc longhorn beetles, all the chestnut woods suffering because of the wasp and severe restrictions imposed on plant nurseries due to their proximity to hot spots of bacterial infestation causing fireblight. To combat the spread of these new and harmful alien organisms, which have become invasive, the Lombardy Region, in compliance with international plant health norms, had to invest over 20 million euro over the past 9 years.

Objectives of the monitoring plan

The introduction of plants and plant products from 144 countries on occasion of EXPO 2015 presented an elevated phytosanitary risk for the exhibition grounds and for the entire Lombardy region. In fact, it was impossible to exclude the involuntary and accidental transport of alien plant and animal species, with consequences that would result in an imbalance of local ecosystems and severe economic repercussions. International norms require that, in the presence of phytosanitary risk, plant health services must take all possible precautions to minimize the same, by applying the appropriate phytosanitsary measures.

Therefore, a plan of reinforced surveillance was designed and implemented, in accordance with the FAO ISPM 6 standard, based on *early detection*, because prompt identification of the presence of an alien organism is a key factor in achieving the objectives of fast eradication and preventing its spread.

In cases of suspected identification of an alien organism classified as harmful according to European norms or on the international warning lists, the plant health service intervened rapidly and immediately implemented the appropriate phytosanitary control measures.

Monitored area

The specifically designed monitoring plan covered the entire exhibition grounds.
In addition, the regional plant health service conducted surveillance activities in the immediate vicinity of the exhibition grounds and in the entire region, applying the modalities set out in the 2015 annual monitoring plan, which was continued also in 2016.
The exhibitin grounds covered an area of 988,337.75 m^2 (ca. 100 ha, 1 km^2), with a perimeter of 5,423.47 m (ca. 5 km). the entire area was subjected to surveillance, subdivided according to specific characteristics and using the same terminology utilized at EXPO, as follows:

1. bordering green areas;
2. green areas in the central part of the exhibition grounds, particularly the *hortus*;
3. pavilions;
4. clusters.

PRELIMINARY STUDY OF THE ORGANISMS TO BE MONITORED

The objective of the monitoring was the identification of alien organisms noxious to plants along with plant products presenting the highest risk of introduction during the event. Attention was focused above all on new species of arthropods, mostly insects, which have caused damage that has been calculated (Kenis & Branco 2010) at 10 billion euros per year to the EU economy. Other organisms monitored included fungi, nematodes, some bacteria, viruses and phytoplasms.
The following is a report of the preliminary analysis carried out during the preparatory phase.

INSECTS

Coleoptera Scolytidae (Bark and Ambrosia beetles)

The coleoptera known internationally by the common name of *Bark and Ambrosia beetles* comprise about 6,000 species, native mostly to tropical and subtropical regions. In Europe, there are about 350 native species. Scolytidae live in wood (bark and sapwood), in which they bore tunnels. 50 species are considered to present a phytosanitary risk, and these are specified in international warning and quarantine lists.
Invasive *scolytidae* are considered the greatest insect threat to the world's forests.
In addition, these insects are often associated with pathogenic fungi; a recent example of this is the finding, in Lombardy, of *Pityophthorus juglandis,* a vector of the fungus *Geosmithia morbida*, which transmits the Thousand Cankers Disease, a serious ailment that threatens walnut trees.

Monitoring instruments: traps for the capture of adult specimens and sampling of material found to be infested (wood used for packing materials and wooden objects).
The traps were baited with specific pheromones, available mostly for European and American species, and generic attractants used for the Asiatic, African and tropical species for which there are no specific pheromones available.
Multi-funnel type traps were used, as these had already been used successfully in Lombardy to capture *Pityophthorus juglandis.*
Possible entry pathways: raw and worked wood. Also packing crates and pallets made with infested wood. Introduction via ornamental plants is less likely, but still possible in trunks of *Dracaena* species, seeds, nuts, orchids and exotic plants of the *Ficus* species.

Coleoptera Buprestidae (Flatheaded borers or jewel beetles)

The *buprestidae* are all plant eaters. Many species are xylophagous and are found mostly under the bark, while others live inside bushes and larvae of the *Trachys* genus bore tunnels in the thickest parts of leaves. They are especially common in tropical habitats, and generally where it is very warm. The smallest species infest flowers (*Anthaxia* spp., *Acmaeodera* spp.) or tree leaves (*Agrilus* spp., *Coraebus* spp.), while the larger ones live in trees and are difficult to spot.

The species of *buprestidiae* that pose the greatest threat and whose introduction is most feared are those belonging to the genus *Agrilus*. The most important of these is indisputably *Agrilus planipennis*, known as the Emerald ash borer because of its bright emerald color and its pedilection for ash trees. Introduced by accident in the USA and Russia, it has caused heavy damage. *Agrilus anxius* has also recently been added to the EPPO alert list: known as the Bronze birch borer*,* it preys on birch trees. The alert list also includes *Agrilus auroguttatus*, the Goldspotted oak borer, a menace to oaks.

Monitoring instruments: reproductive pheromones and attractants have been developed for *A. planipennis*, but they do not seem very useful for early detection; purple prism-shaped sticky traps are instead recommended, baited with mixtures of oils (manuka oil/phoebe oil). These traps are not available on the Italian market, but we shall search for a supplier on the international level, or we will substitute red chromotropic traps. The traps will be placed on ash trees, which will also serve as indicator plants. For the other species of the *Agrilus* genus, we will also use yellow and blue chromotropic traps.

For the genus *Buprestis* it is also possible to use multi-funnel traps baited with pheromones.

Possible entry pathways: wood for industrial use. Bonsai plants.

Coleoptera Chrysomelidae (*Leaf beetles)*

This family includes many insects that cause damage to agricultural crops, such as *Diabrotica virgifera virgifera*, accidentally introduced to Lombardy in the first decade of this century, which has now spread to all the areas in which corn is cultivated. Some *chrysomelidae*, like *Cerotoma trifurcata* and *Epitrix* species, for example, can transmit dangerous viruses to plants.

Monitoring instruments: chromotropic traps, with or without pheromones. Specific pheromones are available for *Oulema melanopus* and species of *Galerucella*. Aspirators can also be utilized.

Possible entry pathways: seeds and vegetables such as potatoes. *Brontispa longissima* is considered one of the most dangerous organisms for palms and can spread through ornamental palms.

Coleoptera Cerambycidae (*Longhorn beetles)*

The *cerambycidae* are a family of beetles that has attracted increasing international attention in recent years, especially due to the high risk of diffusion of some species through wood packing crates and pallets. The treatment system for wooden packing materials made obligatory by the FAO ISPM 15 standard has proven to be not entirely effective in preventing the spread of dangerous insects such as *Anoplophora glabripennis* and *Aromia bungii,* both present in Lombardy, though in circumscribed areas.

Insects of the genus *Monochamus* can transmit the noxious pine wood nematode, *Bursaphelenchus xylophilus*.

Anoplophora chinensis, also present in some areas of Lombardy, is another dangerous insect that preys on a number of broad-leaved ornamental plants.

Xylotrechus chinensis is another species that has been found in wooden crates from China. At present, there is a hot spot in Spain, where it has infested mulberry trees.
Monitoring instruments: for species of *Anoplophora* and *Monochamus,* multi-funnel and cross-vane traps baited with pheromones and kairomones are available and already utilized by the Lombardy plant health service. For *Aromia bungii,* artisanal traps made with plastic bottles baited with wine vinegar are used, but they are not sufficiently effective. The particular vulnerability of plants of the genus *Acer* to species of *Anoplophora* spp. and plants of the genus *Prunus* (pitted fruit trees) to species of *Aromia bungii* make it possible to utilize these plants as indicators.
Possible entry pathways: wooden crates and pallets, wood products, ornamental plants, bonsai.

Coleoptera Curculionidae (*Weevils)*

The best-known member of this family is the red palm weevil, *Rhynchophorus ferrugineus*, present in many parts of Italy but not yet found in Lombardy. There is also a risk of introduction of another species from this genus, *Rhynchophorus palmatum,* which in addition to the direct damage it inflicts on plants can also transmit the nematode *Rhadinaphelenchus cocophilus* (which in turn causes red-ring disease).
Anthonomus eugenii has recently been found in the Lazio region. The species poses a threat to the bell peppers it feeds on.
Sternochetus mangiferae is a *curculionidae* often found at Malpensa airport in shipments of mangoes, *Mangifera indica*, from Sri Lanka.
Megaplatypus mutatus is a weevil native to South America. When adult specimens of *M. mutatus* colonize a new tree, they bore tunnels in the wood. This usually happens to adult trees, with trunks of at least 15 cm in diameter, and the damage can be noted thanks to numerous holes in the trunk and main branches. These insects weaken the plants considerably, due to the extensive tunneling activity of both adults and larvae, diminishing their health and stability and the commercial value of their wood. The insect can be introduced to new areas by the transport of planting stock, sawn trunks or infested wooden crating.
Monitoring instruments: these insects are mostly cryptic and nocturnal, making them difficult to spot.
For some genera, such as *Rhynchophorus,* or for *Megaplatypus mutates,* pheromones are available, with which bucket traps can be baited. Yellow chromotropic traps baited with specific pheromones can also be used to monitor *Anthonomus eugenii*. For other genera, pheromones are still in the developmental stage, so samples of symptomatic and asymptomatic plants will have to be collected.
Possible entry pathways: palms, pines and eucalyptus species; fruit, vegetables and seeds contaminated with larvae or pupae, soil that may contain larvae or pupae.
Curculionidae inside mangoes and traps for red palm weevil.
Megaplatypus mutatus can penetrate new areas through planting stock, sawn trunks or infested wooden crating.

Diptera Tephritidae (*Fruit flies)*

Tephritidae are an important family of plant-eating flies, many of whose species are of considerable economic interest, since they damage agricultural crops.
Many non-European fruit flies are subject to quarantine, and they are frequently found at airports in shipments of fruit and vegetables from foreign countries. The most noxious belong to the genera *Anastrepha, Bactrocera*, *Ceratitis, Dacus* and *Rhagoletis*.
Species of the genus *Anastrepha,* mostly native to tropical forests, do not pose a great threat to plant health, because our habitat is unsuitable to them. But insofar as Asiatic and Australian species of *Bactrocera* and

African *Dacus*, the risk that they may adapt to our climatic conditions is instead high. A species like *B. dorsalis,* for example, which is present in several Asiatic regions whose average temperatures resemble those of the Mediterranean area, is frequently found during phytosanitary inspections on mangoes arriving in Europe from Thailand. The same can be said for *B. tsuneonis*, the Japanese orange fly, and *B. tryoni*, the infamous "Queensland Fruit Fly", whose habitat, in terms of climatic conditions, resembles ours and which may be considered to pose the greatet potential threat, given its extremely varied diet. The threat to plant health is instead quite high for all species of the genus *Rhagoletis*, native to the temperate and northern areas of the United States:

Rhagoletis completa: threatens walnut trees
Rhagoletis pomella: threatens mostly for apple trees (Apple maggot fly)
Rhagoletis fausta: Black cherry fruit fly
Rhagoletis cingulata: Eastern cherry fruit fly
There is a moderate risk of introduction of *Strauzia longipennis* in sunflowers and Jerusalem artichokes, and cut flowers are also a possible pathway.
Monitoring instruments: the procedures for monitoring these dangerous insects are set at the international level by the FAO ISPM No. 26 (2006) *Establishment of pest free areas for fruit flies (Tephritidae)* standard, including instructions on the traps to use and the sampling of fruit.
Both pheromones and parapheromones are used. The traps can be chromotropic, with panels, pagoda, delta or container shaped.
Possible entry pathways: fruit and vegetables.

Diptera Agromyzidae *(Leaf-miners)*

Agromyzidae, or leaf miners are a family of plant-eating insects of the order *Dipterae* (*Brachycera*: *Cyclorrhapha*: *Acalyptratae*). Some species belonging to this family are considered of great economic importance because of the heavy damage they cause to various crops. The species belonging to this family are also commonly referred to as “fruit flies”. The *Agromyzidae* are one of the few families of the order *Dipterae* that include species exclusively adapted to phytophagy in the larval stage. About three quarters of the *Agromyzidae* species tunnel in plants in the larval stage, which explains their common name, leaf miners, while the remaining quarter consists of species that prey on other parts of plants (seeds, roots, stems of grassy plants and sprouts or seedlings of woody plants). Therefore, only a limited number of *Agromyzidae* are of economic importance, and only in specific contexts.
In general, these species feed on a wide range of plants and display a multivoltine reproductive cycle: their ideal habitats are greenhouses and degraded agrosystems. Some species are indicated as harmful, all of which are polyphagous and belong to the genera *Liriomyza* and *Chromatomyia*:
Liriomyza bryoniae: highly polyphagous species. Among the host plants of commercial interest are cabbage (*Brassica oleracea* var. capitata), cucumber (*Cucumis sativus*), lettuce (*Lactuca sativa*), zucchini (*Cucurbita pepo*), melon (*Cucumis melo*), tomato (*Lycopersicon esculentum*) and watermelon (*Citrullus lanatus*). They are a serious threat to tomatoes cultivated in greenhouses.
Liriomyza huidobrensis: 14 families of plants are listed as hosts, without a clear preference. Among the host plants are *Amaranthus* spp, *Aster* spp., eggplant (*Solanum melongena*), beets (*Beta vulgaris*), *Capsicum annuum*, celery (*Apium graveolens*), chrysanthemums (*Dendranthema morifolium*), cucumber (*Cucumis sativus*), *Dahlia* spp., *Dianthus spp*., fava beans (*Vicia faba*), garlic (*Allium sativum*), *Gypsophila* spp., cannabis (*Cannabis sativa*), *Lathyrus* spp., lettuce (*Lactuca sativa*), alfalfa (*Medicago sativa*), melon (*Cucumis melo*), onions (*Allium cepa*), peas (*Pisum sativum*), *Phaseolus vulgaris*, potatoes (*Solanum tuberosum*), *Primula* spp., horseradish (*Raphanus sativus*), spinach (*Spinacia oleracea*), tomatoes (*Lycopersicon esculentum*), *Tropaeolum*

spp., *Verbena* spp. and *Zinnia* spp. *L. huidobrensis*; it has the potential to become the primary parasite for a wide variety of plants. Can damage potato crops severely.
Liriomyza sativae: this species favors host plants from the *Solanaceae* and *Fabaceae* families, but has also been found on other plants, such as alfalfa*, Amaranthus* spp*., Aster* spp*.,* eggplant*, Capsicum annuum*, celery*,* cucumber, *Cucurbita pepo, Dahlia* spp*.*, fava beans, *Lathyrus* spp*.,* melon*,* beans*, Phaseolus lunatus*, *P. vulgaris*, potatoes, tomatoes, *Tropaeolum* spp. and *Igna spp.* Causes severe damage to potatoes, tomatoes and zucchini.
Liriomyza trifolii is native to North America. *L. trifolii* is listed as preying on 25 families of plants, with a preference for *Asteraceae,* also including the following plants: Aster spp., beets, *Bidens* spp., *Brassica chinensis*, *Capsicum annuum*, celery, *Chinese cabbages*, chrysanthemums, cotton, cucumber, *Dahlia* spp., *Dianthus* spp., garlic, *Gerbera* spp., *Gypsophila* spp., *Lathyrus* spp., leeks, lettuce, zucchini, melon, onions, peas and green beans.
Chromatomyia horticola has been observed on the common *Brassicaceae, Fabaceae* and *Asteraceae*. This polyphagous insect is native to various regions of Africa, Asia and Europe.
Chromatomyia singenesiae has several host plants, such as *Asteraceae*, *Cynara scolymus*, artichoke, lettuce, *Cichorium endivia* and some ornamental plants, like chrysanthemums and gerbera.
Phytomyza gymnostoma: host plants include several species of the genus *Allium,* particularly leeks *(A. porrum*) but also chives *(A. schoenoprasum*), onions *(A. cepa*), garlic *(A. sativum*) and shallots *(A. ascallonicum*).
Monitoring instruments: the procedures for monitoring the presence of these dangerous insects have been esablsihed at the international level by the FAO ISPM No. 26 (2006) *Establishment of pest free areas for fruit flies (Tephritidae)* standard, which provides indications for utilization of traps and sampling of fruit.
Both pheromones and parapheromones are used. The traps can be chromotropic, with panels, pagoda or delta-shaped or shaped like containers.
Possible entry pathways: fruit and vegetables.

Diptera Drosophilidae *(Fruit flies)*

There are many species in the *Drosophilidae* family: this is a widely distributed family of *Diptera*, commonly known as “fruit flies”.

Drosophila suzukii

This polyphagous insect is important in economic terms because it infests many kinds of summer-ripening fruit. Its host plants are in fact numerous, both cultivated and spontaneous. Fruit flies prefer cultivated fruit with thin skin of the genus *Prunus* (cherries, apricots, peaches, plums), *Rubus* spp. (raspberries, blackberries) and *Vaccinium corymbosum* (blueberries), as well as *Fragaria vesca* (strawberries), *Actinidia* spp. (*A. arguta*), *Diospyros kaki* (persimmons), *Ficus carica* (figs), *Vitis vinifera* (grapes of all kinids) and *Malus domestica* (apples), and spontaneous fruit such as *Sambucus nigra* (elderberries), *Frangula alnus* (huckleberries), *Rubus* spp. (thornberries and wild raspberries), *Vaccinium myrtillus* (wild blueberries), *Prunus laurocerasus* (laurel berries), *Morus nigra* (mulberries), *Viburnum dilatatum* (Viburnum), *Cornus controversa* (dogwood berries) and *Lonicera* spp. (honeysuckle).
Monitoring instruments: efforts to control these insects are primarily directed against adults, with the aim of reducing their numbers and minimizing egg deposition. The use of traps baited with food monitors the presence of adults and verifies their numbers, but does not pinpoint the best time to intervene.
Possible entry pathways: fruit, ornamental and cut flowers.

Homoptera Cicadellidea and Psyllidea (Leafhoppers, Planthoppers and Psyllids)

Cicadellidae and *Psyllidae* are dangerous carriers of viruses, bacteria and phytoplasms.
Megacopta cribraria is a new species that preys on soy and other legumes. It is introduced concealed in shipments of vegetables and plants, and not only these.
Homalodisca coagulata (*Homoptera*: *Cicadellidae*) vector of *Xylella fastidiosa* bacteria*. Scaphoideus titanus* (*Homoptera*: *Cicadellidae*) vector of Grapevine flavescence dorée phytoplasm, native to North America.
Trioza erytreae: vector of *Candidatus liberibacter* spp bacteria, a serious threat to citrus crops, is causing widespread damage to Florida citrus plantations and is also found in other pars of the world. Its introduction to Europe is feared.
Monitoring instruments: yellow chromotropic traps, aspirators, insect netting, threshing, Malaise traps.
Possible entry pathways: plants, fruit and vegetables.

Hemiptera Aphidoidea (*Aphids)*

More than a thousand species of aphids are present in Europe, 7% of which are of alien origin.
International attention is currently focused on *Toxoptera citricida*, because this species is considered the most efficient vector of the *citrus tristeza* virus, which afflicts citrus fruit.
Monitoring instruments: adhesive chromotropic traps can be used, as well as traps consisting of containers filled with liquid. Suction/aspiration traps are also used, generally by researchers, since they facilitate identification of the aphids captured.
Possible entry pathways: ornamental plants, fruit and vegetables.

Lepidoptera (Moths and butterflies)

Lepidoptera and micro *lepidoptera* can be introduced to our region accidentally. The most recent reports concern *Cameraria ohridella*, *Cacyreus marshalli*, *Paysandisia archon*, *Tuta absoluta* and *Cydalima perspectalis*, known as the box tree moth.
Neoleucinodes elegantalis is the key insect threat to *Solanacae* (nightshades) in Central and South America.
It is not present in Europe, but has been repeatedly found at EU borders in inspections of shipments of eggplant from Suriname and at passenger controls at Schiphol (Netherlands) airport.
Thaumatotibia leucotreta is dangerous to fruit, particularly citrus fruit, and plants of the nightshade family. It has been found in shipments of cut flowers – especially roses – and peppers, peaches and pomegranates from Ethiopia, Tanzania, Uganda and Ghana, as well as in containers and in passengers' luggage coming from those same countries.
Helicoverpa armigera: in the Mediterranean region, it poses a threat to nightshades and corn. It has been found repeatedly in shipments of ornamental plants and cut flowers from Israel.
Spodoptera litura and *S. littoralis:* present in some areas of Europe, but never found in Lombardy, where they could pose a serious threat to corn crops. The eggs and larvae can be introduced accidentally in flowers, especially roses, and vegetables*. S. litura* has also been found in shipments of acquatic plants from Singapore.
Monitoring instruments: the presence of these insects is monitored by visual inspection, luminous traps and pheromone-baited traps.
Possible entry pathways: the primary risk of introduction is via plants, but they can also be found in container shipments of fruit and packing materials. Insects belonging to the *Spodoptera* genus are often found on roses impoted from African countries. Special attention must be devoted to leaf miners.

Homoptera Aleyrodoidea (*Whiteflies)*

These insects pose a direct threat to flowers and leaves and also carry dangerous viruses. The species whose introduction from foreign contries is most feared is *Bemisia tabaci*, especially the insect's non-European populations, which could transmit viruses requiring quarantine measures.
Monitoring instruments: the presence of these insects can be detected by visual inspection of plant samples or by the use of yellow chromotropic traps.
Possible entry pathways: *Aleyrodoidea* can be introduced on a broad range of ornamental plants and vegetables. In particular, they have repeatedly been found on some flowering plants, such as *Corchorus* spp., *Ipomea* sp. and *Colocasia* sp.

Thrips (*Thrips palmi*)

In Europe, *Thrips palmi* is considered to pose the greatest risk. The threat is due to its capacity to transmit the tospoviruses, especially TSWV.
Monitoring instruments: blue and yellow chromotropic traps, visual inspection of symptomatic vegetables and *battage* of asymptomatic flowers and plants.
Possible entry pathways: Thrips have primarily been found at European borders on fruit, vegetables and flowers originating in Asian countries.

FUNGI

The risk of introducing materials infected by fungal maladies is considered high, in part because of the timing of the event, which will be held during seasons favorable to the possible establishment of new fungal pathogens. In addition, given the complexity of the flora surrounding the grounds where the event will be held, including crop cultivation, gardens and nurseries, the spores could migrate to the surrounding areas, spread and adapt to the local conditions.
Monitoring instruments: monitoring is carried out by visual inspection, aimed at identifying alterations on materials and plant tissue (fruit, leaves, trunks, flowers, roots and wood) such as necrosis, rot, mold, blight pustules and chromatic alterations of wood.
In order to identify spores of airborne fungal diseases such as plant and grain blights (e.g. *Tilletia indica* potentially carried by and linked to infected caryopsids and pallets, *Puccinia horiana* potentially carried by chrysanthemum plants and/or cut chrysanthemum flowers, *Plasmopara halstededii* potentially carried by sunflower plants and/or cut sunflowers, *Gymnosporangium* sp. potentially carried by pome fruits) or of other fungi (*Alternaria* spp. potentially carried by infected leaves and insects, *Penicilium* sp. and/or *Monilinia* spp. potentially present on rotten fruit) a spore trap will be installed and direct sampling will be carried out on plants, parts of plants, fruit, vegetables and seeds that are known to be potential carriers of these diseases.
To monitor the presence of zoospores and spores in water, we will utilize bait made with pieces of fruit (ex. pieces of apples or pears) inserted in plastic netting and immersed in the canal that runs along the border of the exhibition grounds. After the portions of apple or pear have been in contact with fluids or potentially contaminated surfaces for a period of seven days, they will be collected and brought to the laboratory in order to isolate oomycetes fungi (longer exposure times for the pieces of fruit could imply rot and contamination by saprophytes and/or parasites).

Atropellis pinicola

Potentially carried by conifers/bonsai /evergreen branches.

Monitoring instruments: inspection of potted conifers or planted trees, coniferous bonsai, collection of necrotic branches of conifers.

Possible entry pathways: plant material

Colletotrichum acutatum

Potentially carried by strawberries and small fruit.

Monitoring instruments: inspection and sampling of rotten fruit

Possible entry pathways: rotten or overripe fruit

Didymella ligulicola

Potentially carried by plants and flowers of chrysanthemum and *asteracea*.

Monitoring instruments: inspection and sampling of flowers with necrotic/rotten petals

Elsinoë australis

Potentially carried by citrus fruit.

Monitoring instruments: inspection and sampling of citrus fruit with lesions/necrotic spots/reticulation on the pericarp.

Possible entry pathways: citrus plants and fruit.

Deuterophoma tracheiphila

Potentially associated with Citrus fruit

Monitoring instruments: inspection and sampling of Citrus plants showing partial wilting/shriveling of the leafy tops.

Possible entry pathways: Citrus plants.

Fusarium oxysporum f.sp. albedinis

Potentially associated with palms and cycadacae

Monitoring instruments: inspection and sampling of palms and cycads showing wilting/shriveling of the fronds.

Possible entry pathways: plants and vegetable matter.

Guignardia citricarpa

Potentially associated with citrus fruit/plants/bonsai.

Monitoring instruments: inspection and sampling of citrus fruit showing lesions /necrosis /reticulation on the pericarp (special attention to fruit originating from South Africa and Australia)

Possible entry pathways: Citrus fruit.

Gymnosporangium clavipes

Potentially associated with pomaceous fruit.

Monitoring instruments: inspection and sampling of fruit of the Malus family with lesions/necrosis on the pericarp.

Possible entry pathways: Fruit.

Monilia fructicola

Potentially associated with pomaceous and drupaceous fruit.

Monitoring instruments: inspection and sampling of drupes and pitted fruit with circular rotten spots and necrosis on the pericarp.
Possible entry pathways: Fruit.

Phoma andina

Potentially associated with solanaceous fruit and potatoes.
Monitoring instruments: inspection and sampling of necrotic leaves of potato plants.
Possible entry pathways: fruit and vegetable material.

Phyllosticta solitaria

Potentially associated with drupaceous and pomaceous fruit.
Monitoring instruments: inspection and sampling of drupaceous and pomaceous fruit with necrosis on the pericarp.
Possible entry pathways: fruit.

Phytophthora fragariae var. fragariae

Potentially associated with strawberries and strawberry plants and small fruit.
Monitoring instruments: inspection and sampling of strawberry plants and small fruit showing wilting, vascular browning and rotten spots on the fruit.
Possible entry pathways: vegetable matter and fruit.

Phytophthora ramorum

Potentially associated with fruit of *Rhodeondron* spp. and *Camellia Viburnum*
Monitoring instruments: inspection and sampling of plants of *Rhododendron Viburnum tinus* Camelias with wilting, leaf or branch necrosis, basal rot and wilting of parts of the leafy tops.
Possible entry pathways: vegetable matter, plants and fruit.

Septoria lycopersici var. malagutii

Associated with solanaceous fruit/potatoes
Monitoring instruments: inspection and sampling of necrotic potato plant leaves
Possible entry pathways: vegetable matter and tubers

Synchytrium endobioticum

Potentially associated with potato tubers
Monitoring instruments: inspection and sampling of potato tubers with deformities or tubercles.
Possible entry pathways: tubers and vegetable matter.

Thecaphora solani

Potentially associated with potato tubers
Monitoring instruments: inspection and sampling of potato tubers with deformities or tubercles.
Possible entry pathways: vegetable matter and tubers.

Tilletia indica

Potentially associated with grain cereals.
Monitoring instruments: inspection and sampling of kernels of grain with cavities and powdery decay.
Possible entry pathways: vegetable material

Verticillium albo-atrum and Verticillium dahlia

Potentially associated with maple, olive and hop plants

Monitoring instruments: inspection and sampling of plants of maple, bonsais of maple and olive and solanacee (eggplant) showing shriveling.

Possible entry pathways: vegetable material.

Diaporthe vaccinii

The main host plant in America and Europe are blueberry plants *(Vaccinium macrocarpon, V. oxycoccos, V. oxycoccos var. intermedium).*

Monitoring instruments: visual inspection of the plants, which can manifest small picnids on leaves, stems and berries.

Possible entry pathways: vegetable material and fruit.

Ramularia collo cygni

Found on barley, wheat and oats

Monitoring instruments: visual inspection and sampling on plants with leaves and stems presenting discolored cankers and/or shriveling.

Possible entry pathways: vegetable material.

Specific plant health risk for some origins and product categories

Particular attention will have to be paid to stands from Latin America containing varieties of potatoes and corn and their germoplasms, as well as other solanaceous plants, due to the risk of introduction of:
Synchytrium endobioticum, Thecaphora solani, Phoma andina and *Stenocarpella maydis.*

NEMATODES

The *Phylum Nematoda* includes worm-shaped animals normally of small size, with bodies composed of circular sections. They have evolved in many different forms, so that nowadays there are saprophytic, mycetophagic, bacterivore and predator species in addition to plant and animal parasites.

The spread of Nematode plant parasites (*Plant Parasitic Nematodes*, PPN) occurs actively, but also and above all through the passive transport of infested vegetable material or soil.

Non-migratory underground endoparasites (Family *Heteroderidae*)

The Family *Heteroderidae* includes species of endoparasitic Nematodes found in roots; during the the event, particular attention will be focused on tubers, taproots and edible roots (potatoes, sweet potatoes, garlic and onions), as well as any whole plants, potted or planted directly in the ground.

Some of the species for which there is a risk of introduction are *Meloidogyne minor* (potatoes and cereals, *Poaceae*) and *Meloidogyne enterolobii* (sweet potatoes, *Hypomea batatas, Coffea arabica, Capsicum annuum*).

On tubers, *M. minor* causes numerous small bumps (galls) that are easily identifiable on the surface. The females lodge immediately under the skin and can cause small points of brown necrotic tissue on the peels of tubers.

Meloidogyne exigua (not included in the EPPO lists) attacks coffee plants, garlic, rice and citrus fruit.

Meloidogyne graminicola is a parasite of rice plants: in cases of serious infestation, they become discolored and their leaves wilt. *Meloidogyne ethiopica* attacks plants of actinidia (kiwi), tomatoes and vines, leading to the general decline of the entire plant, with mold on the stems and galls on the roots.

Meloydogine caffeicola (not included in the EPPO lists) attacks only coffee plants, causing their decline and decay.
Heterodera zeae (corn and cereals, sugar cane) stops the growth of corn plants and causes them to shrivel, with wizened, pale and narrow leaves. The root system is poorly developed and dense, showing cysts on the roots surfaces.
Monitoring instruments: sampling of tubers that manifest pear-shaped galls and roots with basal thickening, as well as symptomatic and asymptomatic cereals, fruit and vegetables.

Migratory underground endoparasites (Burrowing Nematodes)

Radopholus similis is the species name of the so-called "burrowing nematodes"; parasites that are particularly dangerous to banana and citrus trees, but can also be found on coconuts, avocados, coffee, sugar cane, other graminacea and ornamental plants. They cause lesions by digging holes in the roots: infected plants appear malnourished and in need of water. In banana trees they weaken the roots so badly that the plants cannot anchor themselves in the soil and fall over.

Migratory above ground endoparasites (Family *Pratylenchidae*)

Among this group of nematodes, particular attention is devoted to monitoring the presence of *Pratylenchus coffeae,* which was found in Italy in 1992. It attacks coffee and cucurbitacea (as well as banana and citrus plants), parasitizing the canopy and roots.
Infected plants show small necrotic and hydropic spots in the leafy canopy that expand progressively, as well as lacking vigor, due to the simultaneous attack on their hypogeal organs.

Non-migratory above ground endoparasites (Genus *Bursaphelenchus* spp)

Aong the epigeous endoparasitic nematodes, attention is paid to the species *Bursaphelenchus xylophilus,* a nematode of interest to the forestry service that colonizes the resin-producing canals of many conifers, causing their death. It is transmitted by coleoptera of the genus *Monochamus*, which feed on trees infected by *B. xylophilus,* attracted by the particularly pleasing odor of terpenoids.
At present, *B. xylophilus* is feared by forestry managers worldwide and has already caused serious damages in America and Japan. In Europe, its presence has recently been noted in Portugal and Spain. It is introduced via infested pine mulch and wooden crates.
Monitoring instruments: sampling of symptomatic wood, fruit, vegetables and tubers.
Sampling of bark and wood shavings for nematological analysis.
Placing of pheromone-baited traps to monitor the presence of insect carriers (e.g. Gen. *Monochamus spp)*.
Possible entry pathways: wooden packing crates, mulch, fruit and vegetables, tubers.

VIRUSES, VIROIDS AND PHYTOPLASMS

Virus, viroids and phytoplasms are pathogens confined to the interior of a host plant's tissues, and apart from a few exceptions, can only be transmitted to other plants by way of a vector organism that acquires and transfers them. From the EPPO A1 List, we believe we can identify (see Table) some viruses and phytoplasms not yet present in the EU but responsible for serious damages to crops in the areas where they are present, that pose a potential risk of introduction through the movement of vegetable matter connected with the operations of Expo 2015. For some of these, the natural vector is already present in Lombardy (*B. tabaci, T. vaporariorum, M. persica*) and these will be subjected to monitoring using chromotropic traps.

Viruses and phytoplasms associated with potatoes

Andean potato mottle virus, or APMoV, is a virus of the genus *Comovirus* that attacks potatoes and other solanaceae. It is common in Central and South America (Honduras, Chile, Ecuador, Peru, Brazil, Colombia) and is transmitted via contact, by tubers or through an insect vector (*Diabrotica* spp.). It causes necrosis, slow growth and deformed leaves. Potato yellow vein virus (PYVV) attacks only potatoes and is present in Colombia, Ecuador, Peru and Venezuela. It can be transmitted by tubers or by vectors (*Trialeurodes vaporariorum*-semipersistent) and causes yellowing, especially of the veins, and deformed tubers. Potato yellowing virus (PYV or AMV) attacks potatoes and other solanaceae and can be inoculated by aphids, *Myzus persicae* and via tubers. It is present in Chile, Peru and Ecuador and causes yellowing of the plants.

Viruses and phytoplasms associated with tomatoes

Tomato mottle virus (ToMoV) is a virus of the genus Begomovirus that attacks tomatoes and other solanaceae. It is found in Mexico, the USA, Puerto Rico, Guatemala, Honduras, Nicaragua and Cuba. It spreads by contact or through a vector (Aleurodideae like *Bemisia tabaci*-persistent) and causes symptoms like chlorotic patches, slow growth and deformed leaves. Tomato spotted wilt virus (TSWV) is a Tospovirus and is also present in Italy. It attacks numerous vegetable species and ornamental plants and is transmitted by vectors (e.g. *F. occidentalis, T. tabaci*). It causes chlorotic spots on the leaves, which become necrotic and lead to the death of the plant. The observation of this virus is carried out to prevent the introduction of new sources of inoculation from foreign countries.

Viruses and phytoplasms associated with drupes

Prunus mosaic virus is found in the United States and Mexico and is transmitted by the aphid species *E. insidiosus.*

Peach X-disease phytoplasm attacks peach trees, and other drupes and herbaceous plants as well. It is found in Canada and the United States, where it is transmitted through plant propagation material and insect vectors (grasshoppers/leafhoppers). It causes chlorotic spots and rolling up of the leaves.

In addition, there is Lettuce infectious yellow virus, which has several hosts, including lettuce, beets, zucchini, melon, carrots, *Helianthus* spp., *Ipomoea* spp., *Malva parviflora* and *Physalis heterophylla.* It is found in Mexico and the United States, where it is transmitted by the vector *Bemisia tabaci*.

Monitoring instruments: for all types of viruses, viroids and phytoplasms, symptomatic plants are inspected and their vectors are monitored.

Possible entry pathways: seeds, vegetables, plants.

BACTERIA

There are many bacteria whose introduction into Lombardy is feared, and this is why we believe it advisable to constantly control the exposition grounds, in order to prevent the introduction of bacterioses such as, for example, *Erwinia amylovora,* which, although it is already present in limited areas in Lombardy, could be further spread through the importing of ornamental plants, even from other European countries.

Bacteria already present in Italy

Bacteria of the genus *Erwinia*

Erwinia amylovora is responsible for the disease known as fire blight. It mostly afflicts plants of the subfamily Pomoideae, including apple and pear trees, but also ornamental and spontaneous species of the Family Rosaceae, such as *Cotoneaster* spp., *Crataegus* spp., *Cydonia* spp. and *Pyracantha* spp.. The stricken plants

show brown shoots and crook-shaped folding, blackened flowers and brown, shriveled leaves, which look burned: hence the name of the disease. It is carried by numerous insects, including honeybees, or introduced in imported vegetables, fruit and ornamental plants.
Monitoring instruments: sampling of asymptomatic material, inspection of symptomatic plants, monitoring of vectors.
Possible entry pathways: bacteria can be found in seeds, fruit, vegetables, fruit trees and ornamental plants.

Bacteria of the genus *Xylella*

The *Xylella fastidiosa* bacteria calls for particular attention, as it is known to be one of the agents responsible for the rapid wasting of olive trees (CodiRo) in the Puglia region, starting in 2013. It has been identified in pathopgenic variants categorizable in 4 sub-species: subsp. *fastidiosa*, agent of the Pierce disease that affects grapevines, and found in alfalfa, almond trees and maples; subsp. *sandyi* afflicts oleanders, subsp. *multiplex*, agent of the peach leaf scald, is pathogenic to elms, plum trees, almond trees and plane trees; subsp. *pauca* is an agent of variegated chlorosis of citrus trees and includes variants pathogenic to coffee.
At present, it can infect more than 300 plant species and can be carried by insects of the families Cicadellidae and Cercopidae.
Monitoring instruments: collection of vegetable matter from asymptomatic host plants, inspection and collection of symptomatic plants and monitoring of vectors.
Possible entry pathways: host plants of the bacteria and/or plants originating from areas in which the disease is present (for example, coffee plants from Costa Rica and Honduras are at risk).

Other noxious bacteria to monitor, because of the threat they pose to the production of solanaceae, are: ***Ralstonia solanacearum,*** of which several strains are known, some of which could be introduced from tropical countries, and ***Clavibacter michiganensis*** **subsp.** ***Michiganensis,*** potentially transmissible also through imported seeds.

Bacteria not yet present in Italy

Monitoring may also be carried out for other bacteria whose introduction is feared and which pose threats of particular importance not only for Lombardy but for the rest of Italy and Europe. Some of these are listed below.

Bacteria of the genus *Xanthomonas*

Xanthomonas oryzae includes two pathogens, identified by the names of ***Xanthomonas oryzae*** **pv** ***oryzae*** **(**bacterial withering of rice) and ***Xanthomonas oryzae*** **pv.*oryzicola*** (bacterial striation of rice), originating mostly from tropical and equatorial areas of Asia, America and Africa. Both bacteria cause necrotic lesions delimited by veins and sometimes also hydropic areas. They can be spread by irrigation water, through crop residue and seeds. There are various hosts, including the genus *Leersia* spp., *Leptochloa* spp., *Oryza* spp., *Paspalum scrobiculatum*, *Zizani*a, *Zoysia* spp. and other herbaceous plants of the family *Poacea*s.
Xanthomonas campestris pv. Translucens causes bacterial withering in Graminaceae, but can also attack wheat and oats. It is present in Asia, America, Africa and Australia. The pathogen is transmissible via seeds and can infect other plants in the seedling stage.
Xanthomonas axonopodis **pv.** ***Allii*** is found in America, Africa and Asia and is considered a new emergency in several parts of the world. It can infect numerous plants belonging to the species *Allium* (*A. cepa A. sativum, A. porrum*, *A. schoenoprasum*, *A. ascallonicum*, *A. fistulosum*). The disease can be transmitted by seeds, bulbs, irrigation water and atmospheric agents.

Monitoring instruments: collection of seeds, seedlings and asymptomatic material, inspection of symptomatic plants.
Possible entry pathways: these bacteria can be transmitted by seeds, vegetables, fruit trees and ornamental and herbaceous plants.

Bacteria of the genus *Liberibacter*

Candidatus Liberibacter is a bacteria that cannot be cultured, including several species that attack mostly *Citrus* (HLB-huanglongbing) and solanaceae, including carrots (ZC-zebra chip)**.**
HLB includes three species, *Ca*. *Liberibacter africanus*, *Ca. Liberibacter americanus* and *Ca. Liberibacter asiaticus,* while *Ca.solanacearum* (CLso) is associated with zebra chips.
The genus Liberibacter is found in China, Asia, Africa, Oceania and America; Clso has recently been found in Europe as well, in carrots. The genus Liberibacter can be transmitted through plant propagation material and has been found on seeds, but transmission has not been confirmed, and two vectors have been identified, both psylloidea: *Trioza erytreae* and *Diaphorina citri*.
Monitoring instruments: collection of seeds, seedlings and asymptomatic material, inspection of symptomatic plants, monitoring of vectors.
Possible entry pathways: seeds, host plants and/or plants originating from infested areas and insects.

Bacteria of the genus *Ralstonia*

Ralstonia solanacearum *(syn Pseudomonas solanacearum)* is responsible for the disease known as bacterial wilt of potatoes. It attacks several solanaceae, capsaceae, *Musa spp, Pelargonium spp., Zingiber officinale, Nicotiana tabacum* and *Olea europaea.* There are a number of races, some of which can be introduced from tropical countries in seeds and ornamental plants.
Monitoring instruments: sampling of seeds, seedlings and asymptomatic material, observation of symptomatic vegetable matter.
Possible entry pathways: these bacteria can be carried by seeds, fruit, vegetables, fruit trees, ornamental plants and grasses.

Bacteria of the genus *Clavibacter*

Clavibacter michiganensis subsp. sepedonicus (syn Pseudomonas michiganensis), responsible for the plant disease known as Potato ring rot, attacks only potatoes. The disease symptoms vary and appear relatively late in the growing season. Important vectors include infected seeds and tubers, in addition to insects such as grasshoppers, aphids and *Leptinotarsa decemlineata*.
Monitoring instruments: sampling of tubers and asymptomatic material, observation of symptomatic vegetable matter, monitoring of vectors.
Possible entry pathways: infected tubers.

GASTROPODS

Pomacea

The *Pomacea* (Perry) genus consists of freshwater snails belonging to the Family *Ampullariidae*. Though native to South America, they are now common in the United States and Southeast Asia, and since 2009 the species *Pomacea insularum* has colonized the Ebro river basin, in Spain. Numerous plant species that grow in aquatic habitats or humid conditions are vulnerable to these snails.
In Southeast Asia snails of genus *Pomacea* are considered agricultural parasites, with the greatest threat posed by species of the "canaliculata" group, the most common being *Pomacea canaliculata*, *Pomacea insularum* and *Pomacea lineata*.

Pomacea canaliculata, (the golden ampullaria), is regarded as posing a major threat to rice cultivation, and is capable of altering the ecosystem of the humid areas it colonizes, resulting, if its population density is high enough, in the near total absence of aquatic plants. This accounts for its being listed among the one hundred worst invasive alien species in the world. The principal vector of the possible introduction of these parasites in Europe is the importing of infested aquatic plants or of specimens of the snails intended for aquarium hobbyists. Once they have been introduced into a new area, thanks to their prolific reproductive rate, snails of the genus *Pomacea* raidly invade the bodies of water and wetlands they find themselves in.

Monitoring instruments: visual inspection of rice paddies and wetlands.

Possible entry pathways: their eggs can be introduced attached to cuttings of aquatic plants.

PHYTOSANITARY RISK BY PATHWAY

	WPM	Woodwork	Ornamental trees	Seeds, floweres, fruits and vegetables
Bark and Ambrosia beetles	HR	R	R	NO
Flatheaded borers or jewel beetles	HR	R	R	NO
Leaf beetles	NO	NO	R	R
Longhorn beetles	VHR	R	R	NO
Weevils	SI	NO	R	HR
Fruit flies	NO	NO	NO	VHR
Aphids	NO	NO	LR	R *Citrus*
Leafhoppers, planthoppers and Psyllids	NO	NO	R	R
Whiteflies	NO	NO	VHR	NO
Moths and butterflies	R	NO	R	R
Thrips	NO	NO	R	R
Fungi	NO	NO	R	HR
Nematodes	VHR	NO	R	HR
Virus and Phytoplasms	NO	NO	R	R
Bacteria	NO	NO	R	R

PHYTOSANITARY RISK BY COUNTRY OF ORIGIN

	AFRICA	NORTH AMERICA	CENTRAL AND SOUTH AMERICA	ASIA
Bark and Ambrosia beetles	Camerun HR Congo HR	USA R		Vietnam HR
Flatheaded borers or jewel beetles				
Leaf beetles				
Longhorn beetles and other wood borers		USA R		Cina VHR+ India VHR
Weevils	Rep. Dominicana HR			Cina HR Sri Lanka HR
Fruit flies	Bangladesh HR Costa D'avorio VHR Gambia R Ghana HR Kenia HR Mali HR Rep. Dominicana HR Senegal HR		Jamaica HR Messico R Perù HR Suriname R	Cambogia HR Cina R India HR Pakistan HR Sri Lanka VHR Thailandia HR Vietnam R
Aphids				
Leafhopeers, planthoppers and Psyllids	Uganda HR			
Whiteflies	Bangladesh HR Ghana HR Nigeria HR Sierra Leone R			Cambogia VHR Giordania HR India HR Indonesia R Israele R Malesia R Sri Lanka HR
Moths and Butterflies	minatori fogliari Etiopia R Ghana VHR Kenia HR Sud Africa HR Uganda spodoptera VHR		Minatori fogliari Colombia R Ecuador HR	Minatori fogliari Cambogia VHR Israele R Sri Lanka HR
Thrips	Bangladesh HR Ghana VHR+ Rep. Dominicana VHR		Suriname R	Cambogia HR India HR Pakistan HR Thailandia R
Fungi	Bangladesh R Sud Africa HR Swaziland HR Zimbabwe R		Argentina R	
Nematoded			Costa Rica R	Cina R
Viruses and Phyitoplasms				
Bacteria	Bangladesh HR		Uruguai R	Cina R Pakistan HR

MONITORING INSTRUMENTS

	Pheromon traps	Sticky traps	Other traps	Asymptomatic samples	Symptomatic samples	Sentinel trees
Bark and Ambrosia beetles	YES	NO	NO	NO	YES	NO
Flatheaded borers or jewel beetles	YES	YES	NO	NO		Fagus Betula
Leaf beetles	YES	YES	NO	NO	YES	NO
Longhorn beetles	YES	NO	NO	NO	YES	Acer
Weevils	YES	YES	NO	YES	YES	NO
Fruit flies	YES	YES	NO	YES	YES	NO
Aphids	NO	YES	Extractor fan	NO	NO	NO
Leafhopeers, planthoppers and Psyllids	NO	YES	Extractor fan Malaise	NO	YES	NO
Whiteflies	NO	YES		YES	YES	NO
Moths and Butterflies	YES	NO		NO	YES	NO
Thrips	No	YES		YES	YES	NO
Fungi	NO	NO	Spore caps/apple bites	YES	YES	NO
Nematodes	NO	NO	NO	YES	YES	YES
Viruses and phytoplasms	NO	YES for vectors	NO	NO	YES	NO
Bacteria	NO	YES for vectors	NO	NO	YES	NO

TIME FRAME

2015 MONTH	1	2	3	4	5	6	7	8	9	10	11	12
Setting up checkpoints												
Placing pheromon lures												
Placing sticky traps												
Visual inspections												
Symptomatic sampling												
Asymptomatic sampling												
2016 MONTH	1	2	3	4								
Check apple bites in waterways												
Check sentinel trees												

RESULTS OF THE MONITORING ACTIVITIES

Monitoring for the possible presence of alien organisms on the exhibition grounds was carried out using various instruments and surveillance methods, depending on the type of organism in question.
The work consisted of a phase that took place directly on the premises of the exhibition, involving the placing of the traps and instruments, their periodic control and the visual inspection of indicator plants particularly vulnerable to certain types of organisms. These activities required the nearly constant presence of specialized technicians on the exhibition ground (Table 1).

Table (1): types of controls.

Monitoring tool	Target
Sticky traps and pheromon traps	Insects
Spore caps	Spores
Apple Bites in waterways	Fungal spores
Asymptomatic sampling	Fungi, viruses and bacteria
Symptomatic sampling	Fungi, viruses and bacteria
Sentinel trees	Insects, fungi, viruses and bacteria a

Subsequently, all the material collected was analyzed and identified in the laboratory, where it was subjected to various diagnostic, morphological and molecular tests and procedures.

Monitoring instruments

Short Multifunnels: 14 funnel traps were placed at various points, covering the entire exhibition grounds. These instruments were used to capture insects such as *Anoplophora sp.* (Asiatic longhorn beetle), *Monochamus sp*. (pine cerambicid beetle) and *Pityophthorus juglandis* (walnut scolitidae). They were placed on specific host plants at a height of 3-5m and baited with pheromone attractants during the season in which the adult insects take flight.

Purple prisms: Along the entire perimeter of the Expo where vegetation was present, **10** prismatic traps for *Agrilus sp*. were placed. These traps consist of large purple prisms with a sticky side and are baited with specific pheromones for the genus *Agrilus*. The traps were set at a height of about 4-5m, on oaks, ashes and birches, which are hosts, respectively, for *Agrilus auroguttatus*, *Agrilus planipennis* and *Agrilus anxius*, species of miners not yet present in Italy.

Flanged green traps: 10 cross traps were distributed throughout the Expo grounds to capture the noxious insect *Popillia japonica*, recently introduced into Lombardy. These traps consist of an upper part shaped like a cross, on which a specific pheromone bait is placed, and a bottom bucket in which the insect remains trapped.

Bucket traps: 4 bucket traps to capture *Rhyncophorus spp.,* red palm weevil, were placed on the exhibition grounds. The buckets were placed under plams of the genus *Phoenix,* baited with specific pheromones and filled with a mixture of soap and water that prevents the insect from crawling out of the bucket.

Planotraps: 3 planotraps were set to capture *Planococcus sp*., in particolar for *Planococcus citri* and *Planococcus ficus*, two mealy bugs already present in Italy. The traps consists of a red cover with a sticky side and were baited and placed on lemon and fig trees inside the grounds.

Pagodas/Delta traps: 14 pagoda traps were placed inside the grounds and used to capture various insects: *Spodoptera littoralis* (cotton moth), *Thaumatotibia leucotreta* (false codling moth), *Keiferia lycopersicella* (tomato "Oxyuris"), *Tecia solanivora* (Guatemalan potato moth), not yet present in Italy. They consist of small pagoda-shaped structures with a sticky bottom, inside which a specific pheromone bait is placed.

Apple bait: In the canals running through the exhibition grounds and in the waterway that flanks it **6** mesh cages were placed containing apples cut in half, used to identify Oomycetic fungi. The spores of this fungus are carried by water and proliferate on apples.

Rebell white/yellow: **8** white chromotropic cross traps were placed on the exhibition grounds, as well as **8** yellow chromotropic cross traps, used to capture sawflies and fruit flies (*Tephritidae*), respectively. These traps consist of two crosspieces covered with glue, whose color attracts the target insects. One of the yellow Rebell traps was baited with a specific pheromone for *Rhagoletis sp.*

Spore trap: Inside the external perimeter vegetation around the Expo grounds, an instrument was installed to capture airborne fungal spores present on the site. This instrument is equipped with solar panels that power a small roating drum on which a silicone strip is attached. The airborne spores are trapped by a slit in the drum's surface.

Chromotropic yellow/blue traps: 55 chromotropic yellow and blue traps were installed throughout the exhibition grounds to capture various kinds of fruit flies. The strips are glued on both sides and attract the insects because of their color.

Chromotropic yellow/blue traps for Thrips: Inside the exhibition grounds and along the perimeter with vegetation, **20** chromotropic yellow and blue traps were installed and baited with a specific pheromone for the genus *Thrips.*

Monitoring by indicator plants

For the control of viruses, viroids, bacteria and fungi, indicator plants were identified inside the exhibition grounds, and at the same time, a visual inspection of possible host plants was also conducted. Periodically, these were checked for disease symptoms, targeting the various organs of the identified plants. In cases in which visual inspection targeting specific symptomatologies did not allow for a definite diagnosis in the field, samples were collected and sent to the laboratory for analysis.
Inside the site **17** plants were identified and marked as controls for specific pathologies and/or to check for the presence of noxious organisms, such as *Anoplophora sp*., *Agrilus sp*., *Erwinia amylovora*, *Phytophthora ramorum*, *Bursaphelencus sp*. and *Gibberella sp*., *Flavescenza dorata*, *Xylella fastidiosa*, *Sharka sp*. and *Citrus tristeza virus*.

INSECTS

The presence of insects requiring quarantine was monitored using various kinds of traps, depending on the type of insect targeted. Table 2 shows the types, numbers and target organisms of the traps used.

A total of 149 traps were installed, covering the entire surface area of the Expo, and various controls and replacements were carried out; 87 traps were baited with pheromones and/or kairomones.
Most of the traps used were chromotropic of different colors and shapes according to the target parasite. Exactly 75 chromotropic traps were installed, and all the traps with a sticky surface were inspected or replaced, based on the type of trap and the insects captured.
As for the multifunnel traps, the green traps with flanges and the bucket traps for the red palm weevil, they were inspected during controls and their contents were removed for identification purposes, if considered important.

Table (2): summary of the species of insects for which Early Detection measures were appplied, with the respective number of traps installed on the exhibition grounds

TRAPS				
Trade name	**Kind of traps**	**Pests**	**N.° traps**	**Lures**
WitaTrap Multi Funnel Trap	Trappole multi-imbuto con attrattivi	*Anoplophora spp.*	10	Pheromone α, β- Kairomone 1,2,3 - ChemTica Internacional
		Pityophthorus juglandis	1	Bubble cap formulation of WTB lure from Contech Enterprises Inc.
		Monochamus spp.	3	Galloprotect Pack (1 Pheromone, 3 kairomone) - SEDQ
Purple Prism Trap (EAB Trap)	Trappola a prisma adesiva sui tre lati	*Agrilus spp.*	10	Pheromone Manuka oil and Leaf alcool
Biogard Rhynchotrap	Secchielli	*Rhyncophorus spp.*	4	EROGATORE RHYFER 220 – BIOGARD/ CBC
Trécé Catch Can Trap – (TBC Jap. Beetle Trap)	Trappole a doppio attrattivo	*Popilia japonica*	10	Double lure baitpack system from GREAT LAKES IPM - USA
Koppert Delta trap	Pagoda Delta Trap con fondo collante	*Thaumatotibia leucotreta*	5	Erogatore in gomma con feromone specie-specifico PHERODIS - KOPPERT
		Tecia solanivora	3	
Biogard BDT Delta Trap	Trappola a pagoda con aperture laterali e fondo collante	*Spodoptera littoralis*	3	Erogatore in gomma con feromone specifico BIOGARD/ CBC
		Keiferia lycopersicella	3	Erogatore in gomma con feromone specifico BIOGARD/ CBC
Traptest Isagro	Trappola adesiva con tettuccio e fondo collante	*Sesia* sp.	1	Isagro pheromone dispenser
Biogard Planotrap	Trappola rossa a capannina	*Planococcus ficus*	2	Erogatore in gomma con feromone specifico BIOGARD/ CBC
		Planococcus citri	1	
Biogard Rebel Amarillo	Cromotrappole collanti a croce gialla	*Tefritidi (Bactrocera spp.; Ceratitis spp.)*	7	Nessun attrattivo
		Rhagoletis spp.	1	Erogatore a cilindro con attrattivo ammoniacale BIOGARD/CBC
Biogard Rebel Bianco	Cromotrappole collanti a croce bianca	*Coleotteri (Byturus spp.); Imenotteri (Hoplocampa spp.)*	8	Nessun attrattivo

Trade name	Kind of traps	Pests	N.° traps	Lures
Biogard Glutor Blu	Blue sticky traps	*Thrips spp.*	10	Kairomone - LUREM-TR. - KOPPERT
		Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; Agrilus auroguttatus; Thrips palmi.	18	No lures
Pherocon Pew Monitoring Trap-two-sided yellow "sticky card"	Yellow sticky traps	*Anthonomus eugenii*	8	Pherocon PEW Controlled Release Systems (two component lure) from GREAT LAKES IPM – USA
Biogard Glutor Giallo	Yellow sticky traps	*Thrips spp.*	10	Kairomone - LUREM-TR. - KOPPERT
		Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; Agrilus auroguttatus; Thrips palmi; Diabrotica virgifera; Anthonomus eugenii; Anastrepha spp.; Megacopta cribraria; Trioza eritreae; Bemisia tabaci.	25	No lures

The various traps were positioned to cover the entire area of the exhibition grounds and, where possible, near host plants for the target parasite.
ATTACHMENT 1 contains a map of the Expo grounds, showing the positions of all the traps and control points.
ATTACHMENT 2 contains a Table with the identification codes of all the traps and the coordinates of their location with reference to the map of the Expo grounds.

Laboratory activities

Traps and/or samples of insect specimens removed from the traps were delivered to laboratory personnel once or twice monthly, at regular intervals. The traps and the individual samples were examined under a stereoscopic microscope to identify the species captured, with particular attention to identifying specimens of species requiring quarantine measures. Some specimens were photographed through the stereo microscope. The samples that could not easily be identified morphologically were removed from the traps (either the entire insect or a fragment thereof) and subjected to molecular analysis to amplify gene fragments for DNA barcoding, in order to identify the species, they belonged to.
The DNA was extracted according to a protocol that calls for the mechanical breakup of the sample in a CTAB 2% buffer (Doyle&Dolye, modified) and subsequent steps using chloroform/isoamylic alcohol 24:1, isopropanol and 70% ethanol.
The molecular analysis consisted of the isolation of a fragment of DNA using the PCR (Polymerase chain reaction) technique, with universal barcode primers (LCO1490/HCO2198, Folmer et al., 1994; Lepf1/Lepr1, Hebert et al., 2004) and its sequencing, which was outsourced to specialized foreign companies. The

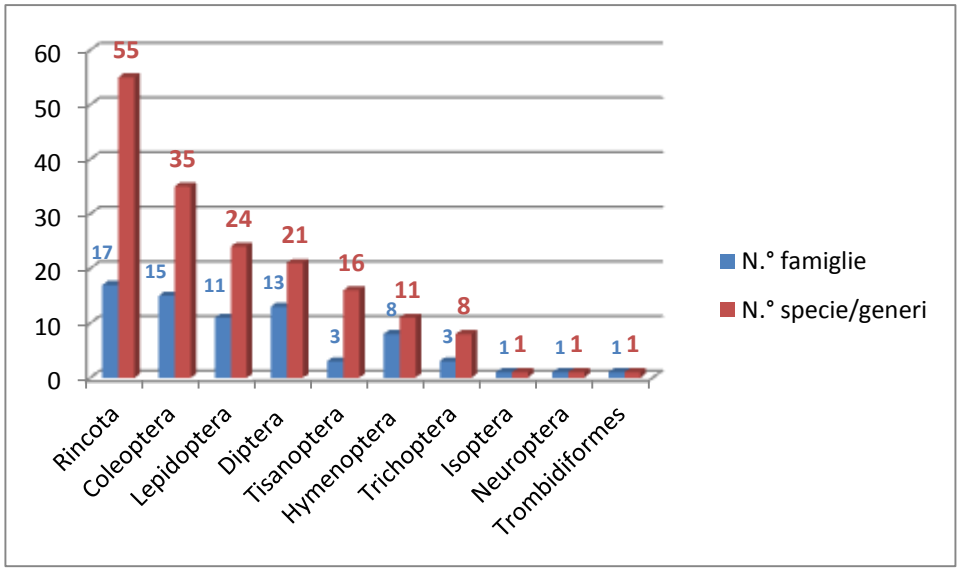
sequences obtained were then compared to sequences stored in international databanks (BOLDSYSTEMS – Barcode of Life Data Systems).
In the case of Thrips, species-specific PCR profiles were applied (Yeh et al., 2014a, Yeh et al., 2014b), to recognize the most common species in the territory, such as *Frankliniella occidentalis, F. intonsa* and *Thrips tabaci*.

Results and discussion

Monitoring of entomofauna

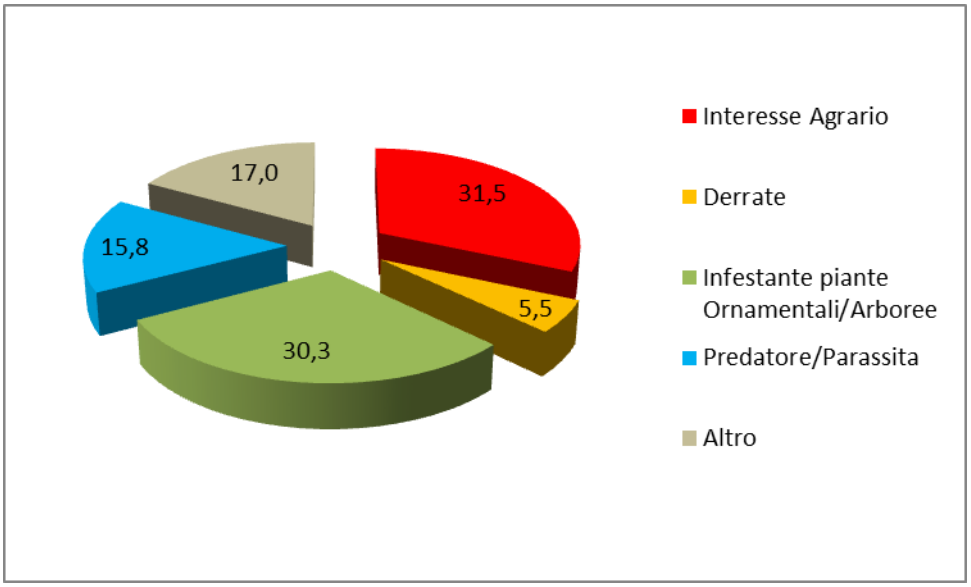
During the monitoring period for EXPO, 340 samples were sent to the laboratory, consisting of traps and entomological specimens collected from traps directly on site.
In some cases, the specimens had been exposed to adverse atmospheric conditions in the traps and had been smeared with glue, which did not allow a clear morphological identification of the insects. Specimens in good condition which could not be identified morphologically were subjected to molecular analyses to ascertain the species they belonged to. 453 single specimens were subjected to DNA extraction and gene amplification: in 331 cases, amplifiable DNA was obtained, while in 317 cases the genus or species were unequivocally determined through sequencing of the amplified product or species-specific matching trials.
The total number of species identified using either morphological or molecular methods was 141. In some cases, it was only possible to identify the genus, for a total of 32 genera. The total number of taxa identified was 173 (Table 4).

Graph (1): number of species/genera and families identified, subdivided by Order



The specimens identified can be subdivided into 5 basic categories: insects of agricultural interest, insects related to foodstuffs, insects that infest ornamental plants and trees, predator/parasitic insects and Others (category including pollinators and aquatic insects) (Graph 2).

Graph (2): total number of taxa identified subdivided in the 5 categories (percentage values).



Of the species identified, almost 62% are **phytophagous**, characterized by trophic relationships with herbaceous host plants or bushes and trees, some **of interest for agricultural crops** and others which **infest ornamental plants**. Among these the following are worthy of note:

- Parasites of vegetables: *Tuta absoluta*, the codling moth; *Helicoverpa armigera, Autographa gamma, Heliothis peltigera* and *Spodoptera exigua*, moths that prey on solanaceae; *Pieris rapae* and *Plutella xylostella,* lepidoptera that defoliate cruciferous plants; *Athalia rosae rosae*, sawflies that prey on cruciferous plants; *Acrolepiopsis assectella*, the leek moth;
- Parasites of cereals: *Diabrotica virgifera virgifera*, a key corn phytophage;
- Parasites of fruit plants: *Drosophila suzukii*, fruit fly preying on small fruit; *Cydia funebrana*, the drupe moth, and *Rhagoletis cerasi*, the cherry fly; the brown marmorated stink bug *Halyomorpha halys,* and *Raphigaster nebulosa*; *Scaphoideus titanus, Orientus ishidae, Philaenus spumarius*, grasshoppers and grapevine spittlebugs; *Pseudococcus citri, P. longispinus, P. viburni*, mealybugs that prey on citrus and other fruit trees;
- Xylophagous insects: *Ips sexdentatus, Xyleborinus saxeseni, Chlorophorus varius varius*;
- Parasites of ornamental plant species: *Galerucella luteola*, the elm beetle; *Gypsonoma aceriana*, a moth that develops inside young poplar seedlings; *Duponchelia fovealis, Cacoecimorpha pronubana*, *Clepsis peritana, Nomophila noctuella, Paracorsia repandalis, Aproaerema anthyllidella,* Lepidoptera that feed on herbaceous plants;
- Thrips: *Frankliniella occidentalis, F. intonsa, Thrips flavus*, *T. tabaci*, *T. major*, characterized by a broad range of host plants; *Microcephalothrips abdominalis. Tenothrips frici, Thrips physapus, T. trehernei,* which infest asteraceae; *Haplothrips aculeatus, Anaphothrips obscurus, Frankliniella tenuicornis*, parasites of graminaceae.

8 species, equal to 5.5%, infest stored **food supplies**, and are divided between Coleoptera and Lepidoptera. Among the coleoptera, *Stegobium paniceum*, the bread weevil, is widespread and infests vegetable matter rich in starch, as well as horsehair and spices; *Cryptophagus pilosus* infests wheat, flour and its derivatives and dried fruit; *Lyctus africanus* attacks roots of licorice and ginger and papaya seeds. Among the lepidoptera, *Sitotroga cerealella*, the grain moth, attacks cereals, as well as legumes and dried chestnuts; *Cadra cautella*, a cosmopolitan species whose larvae feed on dried fruit, cocoa, walnuts and grain seeds. Species of the genus *Corticarina*, *Typhaea stercorea* and *Nemapogon clematella* infest food supplies colonized by fungi, which they feed on.

A substantial percentage (15.8%) of the species fall into the categories of insects that **prey on or are parasites/parassitoids** of plant-eating insects. Among these are the carabids *Trechus quadristriatus* and *Calathus* sp., predators, and various species of coccinellidae: *Adalia bipunctata, Adalia decempunctata, Propylaea 14-punctata, Scymnus auritus* and *Stethorus punctillum. Harmonia axyridis*, a species native to Asia, was voluntarily introduced to Europe as part of a program to combat aphids using their natural enemies and is now a threat to local coccinellidae and infests domestic locales.

Orius majusculus is a rhyncote of the anthocoridae family, a voracious predator of thrips often utilized in programs of biological pest control. *Deraeocoris lutescens* and *D. nebulosus* (Miridae) are predator species, feeding mostly on aphids.

Among the diptera, *Coenosia attenuata* is a species that preys on other insects, common greenhouse parasites; the larvae feed on juvenile forms of fungus gnats found in the soil, while adults actively hunt the winged forms of aphids, white flies, fungus gnats and leaf miners.

The species of the genus *Aphelinus* are hymenoptera that are endoparasites of aphids, while the diptera *Tomosvaryella kuthyi* parasitizes species of Cercopoidea, Cicadellidae and Fulgoridae. The genus *Euplectrus* is an ectoparasite of Lepidoptera larvae, while *Alloxysta* parasitizes braconid and chalcid wasps, which in turn are parasitoids of aphids (hyperparasitism).

Finally, 17% of the species fall into the category of **other insects**, not easily included in other categories, which include pollinators, decomposers of organic substances and detritus feeders, as well as aquatic insects.

The traps captured several species of trichoptera (caddisflies), holometabolic insects whose immature stages develop in aquatic habitats: *Hydropsyche modesta, Agraylea sexmaculata, Hydroptila angulata, Hydroptila angulans, Hydroptila vectis* and *Mystacides azureus*. Other aquatic species include chironomid diptera *Cryptochironomus supplicans* and *Cricotopus* sp., and psychodidae dipteral (moth flies) *Clogmia albipunctata* and *Psychoda alternata*, found in water rich in organic substances.

An adult of *Kalotermes flavicollis*, one of the two species of termites present in Italy, was captured by a yellow chromotropic trap.

Below are brief descriptions of some species found during the monitoring but **never described in Italian fauna lists**.

Thrips hawaiiensis (Figure 1)

A member of the Thrips family, common and widely distributed in tropical Asia and areas of the Pacific, it is also present in some localities of Central and North America and in Africa. Extremely polyphagous, it feeds on a broad range of flowers and fruit of host species, among which tobacco, roses, gladiolus, coffee, cabbage, mango, citrus fruit, apple, pear and banana trees (Reynaud et al., 2008). In the past it was often confused with *T. florum*, so the list of host species attributed to the insect must be considered with caution. It does not carry viruses.

Despite some captures by phytosanitary services in the United Kingdom and France, this thrips was never indicated as a colonizer in the Palearctic area until 2008, when it was found in the south of France on *Abelia* sp. and *Nerium* sp. (Reynaud et al., 2008).

In 2011 it was identified in Spain, where the thrips was found on *Rosa* sp., *Nerium oleander* and *Gerbera* sp. (Goldarazena, 2011). Most recently, in 2015, the insect was found on lemon trees in the Turkish province of Mersin (Atakan et al., 2015).

A study reports that the Mediterranean basin is a potential habitat for this thrips in terms of availability of host plants and favorable climatic conditions (Reynaud et al., 2008).

Fig (1): image of *Thrips hawaiiensis*: female (left) and male (right)

Clepsis peritana (Figure 2)

Also known as the garden or strawberry leaf-roller, this butterfly species belongs to the Tortricidae family. This insect is common in North America: its presence has been noted in southern Canada and throughout the United States, and as far south as Cuba. In Europe it was identified for the first time in Denmark in the late '80s (Larsen and Vilhelmsen, 1991); subsequently, it was found in the province of Granada and on Gibraltar (Nupponen et al., 2003; Clifton, 2007). In these areas, the adults are present from March to September and there are several generations each year. The larvae feed on leaves of *Chrysanthemum* sp., *Cynara cardunculus*, *Senecio jacobaea*, *Stachys* sp., *Fragaria* sp., *Citrus* sp., *Scrophularia californica* and *Solanum torvum*.

Fig (2): image of *Clepsis peritana*: left, a specimen with wings extended, right, a specimen with wings folded

Deraeocoris nebulosus (Figure 3)

A member of the Mirid family, this insect preys on many plant-eating insects and acari. It is associated with many infesting insects, commonly found on over 50 ornamental arboreal species and bushes (Wheeler et al., 1975). *D. nebulosus* has been identified in southern Canada and is currently found throughout the United States (Henry and Wheeler, 1988) and is common in the eastern states (Knight, 1941). It is the first mirid to be described as a predator in North America (Wheeler et al., 1975).

D. nebulosus has been found on more than 75 different plant species (Wheeler et al., 1975, Snodgrass et al., 1984), including in apple orchards (Parrella et al. 1981), peach orchards and walnut stands (Gorsuch et al., 1989, Mizell and Schiffhauer 1987), cotton fields (Snodgrass, 1991) and on spontaneous herbaceous plants (Wheeler et al. 1975). It preys on white flies, aphids, psyllids and lady bugs.

Fig (3): image of ***Deraeocoris nebulosus***

Eutettix variabilis (Figure 4)

Of the family Cicaldellidae, subfamily Deltocephalinae, present in the Neoarctic regions of North America.

Fig (4): image of *Eutettix variabilis*

Other species deserve closer examination, because these represent **allochtonous insects recently found in other areas of Italy**.

Halyomorpha halys

The Brown Marmorated Stink Bug (*Heteroptera*: *Pentatomidae*) is native to the Far East (China, Korea, Japan, Taiwan) (Lee et al., 2013). It has proven to be highly invasive in the countries where it has been introduced accidentally through commercial channels. Present in the US from the mid-nineties on, it developed into a key phytophagous pest on fruit trees starting in 2010 (Leskey et al., 2012a). In Europe, it was first identified in Switzerland and is currently present in France, Hungary and Greece (Haye et al., 2015). In Italy, *H. halys* was found for the first time in 2012, in Modena, and in 2014 it was found in the regions of Emilia-Romagna, Lombardy and the Piedmont (Maistrello et al., 2014; Cesari et al., 2015).

H. halys passes the winter in the adult stage, emerging from its shelter in April-May and attacking the vegetation. In June-July the females deposit groups of eggs on the underside of leaves (Hoebeke and Carter, 2003; Lee et al., 2013). After passing through five developmental stages, the insect reaches its adult form, with growth rates depending on temperature and humidity: in the colder regions, the insect can only complete one reproductive cycle, while two generations per year are the general rule in warmer climates (Hoebeke and Carter, 2003). This stink bug consumes a vast range of plants, both spontaneous and cultivated, including all fruit plants, grapevines, almost all vegetables, legumes and grains (corn, wheat), along with numerous ornamental plants (Leskey et al., 2012b). Both immature and adult specimens prefer to eat seeds and fruit, but they can also feed on every other part of the host plant above ground. The suction holes left by the insect's proboscis can lead to lesions including: abscesses, deformities, discoloration, scarring, depressions, dark patches and the suberization of the pulpy parts of fruit, resulting in serious economic damage (Hoebeke and Carter, 2003). In Asia, it is recognized as a carrier of phytoplasms that strike Paulownia and is suspected to also be a vector for phytoplasms noxious to other plant species (Jones and Lambdin, 2009). *H. halys* has also drawn attention as an urban infestant, because during the winter months adults crowd together, seeking refuge in man-made structures (Lee et al., 2013; Hoebeke and Carter, 2003).

Hishimonus hamatus

A deltacephalous ladybug native to Asia, it was officially identified for the first time in Europe in 2012 in Slovenia (Seljak, 2013). There have been various unofficial sightings in Italy, posted in web forums (www.entomologiitaliani.net and www.naturamediterraneo.com), in Lombardy dating back to 2008, on the Island of Elba in July 2010, in Bovolone (VR) in November 2011 and in Castel Maggiore (BO) at the end of 2013.

In the province of Reggio Emilia it was captured for the first time in the summer of 2014, on yellow chromotropic traps hung in a vineyard.

We don't know much about its biological cycle. The adults measure 4-5 mm; greenish yellow head; upper thorax greenish yellow, lower thorax brownish-yellow; scutellum brownish-yellow with a pair of brown triangular markings and a transversal central black line; anterior wings grayish-white with brown veins and a large semi-circular brown spot covering all the central portion.

It has been found on *Ligustrum, Lagerstroemia indica, Euonymus japonicus, Chamaecyparis lawsoniana, Cupressus sempervirens* and *Thuja occidentalis*. Some species of the genus *Hishimonus* have been proven to be vectors for various phytoplasms, genetically quite different from each other.

Luperomorpha xanthodera

The insect (Coleoptera: *Chrysomelidae: Alticinae*) is native to Asia and is common in tropical to temperate habitats. It was identified in Europe for the first time in England in 2003, on a rose (Johnson and Booth 2004). Subsequently, it was found in Italy (Conti and Raspi 2007; Del Bene and Conti 2009), Francia (Doguet 2008), Holland (Beenen and Roques 2010), Germany (Dö-berl and Sprick 2009), Switzerland (Döberl 2010), Hungary (Bodor 2010), Austria (Geiser and Bernhard 2011) and Poland (Wojciech Kozłowski and Legutowska 2014).

In Italy, the insect was found on ornamental plants in several nurseries in Tuscany. Initially identified erroneously as *L. nigripennis* Duvivier. In the areas in Italy in which it was found, this insect completes two reproductive cycles in a year and can pass the winter underground in the larval, pupa or adult stage (Del Bene and Conti, 2009). Adults emerge from their winter slumber or from their chrysalids and begin to be active at the end of March or beginning of April and feed on the first flowers to bloom (generally citrus plants in greenhouses), and then shift to inflorescences of various plant species (polyphagous), including *Nerium oleander*, *Trachelospermum jasminoides, Buddleia* spp., *Viburnum* spp., *Weigelia florida, Euonymus japonicas, Hypericum* spp., *Genista* spp., *Yucca gloriosa, Lagerstroemia indica, Hibiscus syriacus, Eugenia myrtifolia, Bougainvillea glabra, Ligustrum* spp. *Pittosporum tobira, Pyracantha* spp., *Rosa* spp., *Gardenia grandiflora, Citrus* spp., *Clematis* spp., *Hydrangea* spp., *Philadelphus coronaries* and *Datura arborea* (Del Bene and Conti, 2009). The trophic activity decreases in the summer monthsin the hottest months of the summer, and the first specimens of the second generation make their appearance around the end of July.

The insect is anthophagous, damaging petals without touching the ovaries. Consequently, the fruit develops despite the insect's predation (Del Bene and Conti, 2009). The larvae eat rhizomes, but their trophic activity and consumption cause neglibible damage and do not affect the health and vitality of the host plants.

***Jikradia* spp.**

The genus *Jikradia* (Hemiptera: *Cicadellidae: Coelidiinae*: *Teruliini*) is comprised of 22 species known to man and is common in most Neotropical and Neoarctic regions, from the Galapagos to Canada. *Jikradia olitoria* (Say) was found and identified for the first time in Italy by the entomologist Francesco Poggi in 2013 on *Populus tremula* and *Salix alba* in Maresso (LC) (personal communication). The first findings appear to date back to 2010 and mark the first time a species of the genus *Jikradia* has been identified in the Old Continent (Nielson et al., 2014).

Taxon	
Coleoptera	
	Apionidae
	Protapion fulvipes
	Ischnopterapion virens
	Anobidae
	Stegobium paniceum
	Bostrichidae
	Lyctus africanus
	Carabidae
	Calathus sp.
	Trechus quadristriatus
	Harpalus rufipes
	Cerambycidae
	Chlorophorus varius varius
	Coccinellidae
	Adalia bipunctata
	Adalia decempunctata
	Harmonia axyridis
	Propylaea 14-punctata
	Scymnus auritus
	Scymnus sp.
	Stethorus sp.
	Stethorus punctillum
	Chrisomelidae
	Altica oleracea
	Bruchus brachialis
	Chaetocnema tibialis
	Diabrotica virgifera virgifera
	Galerucella luteola
	Luperomorpha xanthodera
	Ophraella communa
	Curculionidae
	Coccotrypes dactyliperda
	Ips sexdentatus
	Sitona hispidulus
	Xyleborinus saxeseni
	Elateridae
	Athous haemorrhoidalis
	Throscidae
	Trixagus meybohmi
	Cryptophagidae
	Cryptophagus pilosus
	Lathridiidae
	Corticarina sp.
	Mycetophagidae
	Typhaea stercorea
	Scarabaeidae
	Cetonia aurata
	Oryctes nasicornis
	Staphylinidae
	Omalium caesum
Lepidoptera	
	Acrolepiidae
	Acrolepiopsis assectella

Taxon	
Lepidoptera	
	Choreutidae
	Choreutis nemorana
	Crambidae
	Duponchelia fovealis
	Nomophila noctuella
	Paracorsia repandalis
	Gelechiidae
	Aproaerema anthyllidella
	Scrobipalpa ocellatella
	Sitotroga cerealella
	Tuta absoluta
	Lycaelidae
	n.i.
	Noctuidae
	Autographa gamma
	Helicoverpa armigera
	Heliothis peltigera
	Spodoptera exigua
	Plutellidae
	Plutella xylostella
	Pieridae
	Pieris rapae
	Pyralidi
	Cadra cautella
	Tineidae
	Monopis imella
	Nemapogon clematella
	Tortricidae
	Cacoecimorpha pronubana
	Clepsis peritana
	Cydia (Grapholita) funebrana
	Grapholita janthinana
	Gypsonoma aceriana
	Pammene albuginana
Rincota	
	Aphididae
	Anoecia sp.
	Aphis craccivora
	Eucallipterus tiliae
	Rhopalosiphum padi
	Rhopalosiphum rufiabdominalis
	Aphrophoridae
	Philaenus spumarius
	Aleyrodidae
	Dialeurodes citri
	Siphoninus phillyreae
	Anthocoridae
	Orius majusculus
	Cicadellidae
	Cicadella viridis
	Euscelidius variegatus
	Eutettix variabilis
	Fieberiella florii
	Hishimonus hamatus

Taxon
Rincota
Japananus hyalinus
Jikradia sp.
Macrosteles sp.
Orientus ishidae
Psammotettix sp.
Psammotettix confinis
Scaphoideus titanus
Empoasca pteridis
Typhlocyba sp.
Zygina lunaris
Zygina nivea
Zygina rhamni
Zyginidia pullula
Coccidae
Saissetia coffeae
Corixidae
Sigara striata
Delphacidae
Laodelphax striatellus
Dictyopharidae
Dictyophora europaea
Flatidae
Metcalfa pruinosa
Lygaeidae
Nysius senecionis
Nysius graminicola
Nysius sp.
Megalonotus sp.
Megalonotus sabulicola
Membracidae
Stictocephala bisonia
Miridae
Deraeocoris nebulosus
Deraeocoris lutescens
Deraeocoris sp.
Lygus sp.
Lygus pratensis
Taylorilygus apicalis
Pentatomidae
Aelia acuminata
Eurydema ventralis
Halyomorpha halys
Rhaphigaster sp.
Rhaphigaster nebulosa
Pseudococcidae
Planococcus citri
Pseudococcus viburni
Pseudococcus longispinus
Rhopalidae
Chorosoma schillingi
Tingidae
Corythucha ciliata
Stephanitis pyri

Taxon	
Diptera	
	Anthomyiidae
	Pegomya sp.
	Calliphoridae
	Lucilia sericata
	Chironomidae
	Cricotopus sp.
	Cryptochironomus supplicans
	n.i.
	Dolichopodidae
	Chrysotus n. sp. choricus grp.
	Drosophilidae
	Drosophila sp. (prob. *simulans*)
	Drosophila suzukii
	Muscidae
	Coenosia attenuata
	Psychodidae
	Clogmia albipunctata
	Psychoda alternata
	n.i.
	Pipunctulidae
	Tomosvaryella kuthyi
	Sciomyzidae
	Dictya sp.
	Sepedon sphegea
	Syrphidae
	Eristalinus taeniops
	Eristalinus sp.
	Stratiomyidae
	Oplodontha viridula
	Tephritidae
	Acanthiophilus helianthi
	Dioxyna sp. (prob. bidentis)
	Euleia sp.
	Rhagoletis cerasi
	Tephritis formosa
Hymenoptera	**Aphelinidae**
	Aphelinus sp.
	Apidae
	Apis mellifera
	Colletidae
	Hylaeus sp.
	Hylaeus leptocephalus
	Eulophidae
	Euplectrus sp.
	Figitidae
	Alloxysta sp.
	Formicidae
	Formica sp.
	Vespidae
	Polistes sp.
	Vespa crabro
	Vespula sp.

Taxon	
Hymenoptera	
	Tenthredinidae
	Athalia rosae rosae
Tisanotteri	
	Aeolothripidae
	Aeolothrips intermedius
	Phlaeothripidae
	Haplothrips aculeatus
	Haplothrips leucanthemi
	Haplothrips sp.
	Thripidae
	Anaphothrips obscurus
	Frankliniella intonsa
	Frankliniella occidentalis
	Frankliniella tenuicornis
	Microcephalothrips abdominalis
	Tenothrips frici
	Thrips flavus
	Thrips hawaiiensis
	Thrips major
	Thrips physapus
	Thrips tabaci
	Thrips trehernei
Isoptera	
	Kalotermitidae
	Kalotermes flavicollis
Trichoptera	
	Hydropsychidae
	Hydropsyche modesta
	Hydropsyche sp
	Hydroptilidae
	Agraylea sexmaculata
	Hydroptila angulata
	Hydroptila angulans
	Hydroptila sp.
	Hydroptila vectis
	Leptoceridae
	Mystacides azureus
	n.i.
Neuroptera	
	Coniopterygidae
	Semidalis sp. (prob. Aleyrodiformis)
Ephemeroptera	
	n.i.
Trombidiformes	
	Eriophyidae
	Aceria erineus

Bibliografia

Atakan E., Pehlivan S., ÖLÇÜLÜ M., Satar S.(2015) A new thrips species recorded in Turkey: Thrips hawaiiensis. Türk. entomol. bült., 2015, 5(2): 77-84

Clifton J. (2007) Clepsis coriacana (Rebel, 1894) new to Eurfurther records of Clepsis peritana (Clemens, 1860) from Gibraltar (Lepidoptera: Tortricidae). SHILAP Revta. lepid., 35 (137), 2007: 47-48.

Folmer O, M, Black WH, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome C oxidase subunit I from metazoan invertebrates. Molecular Marine Biology and Biotechnology 3: 294-299.

Hebert PDN, Penton EH, Burns JM, Janzen DH, Hallwachs W (2004) Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly Astraptes fulgerator. Proceedings of the National Academy of Sciences of the United States of America 101: 14812-14817.

Kenis M, Branco M (2010) Chapter 5: Impact of alien terrestrial arthropods in Europe. In: Roques A et al. (Eds) Alien terrestrial arthropods of Europe. BioRisk 4(1): 51–71. doi: 10.3897/biorisk.4.42

Roques A, Rabitsch W, Rasplus JY, Lopez-Vaamonde C, Nentwig W, Kenis M (2009) Alien terrestrial invertebrates of Europe. In: Hulme PE, Nentwig W, Pyšek P, Vilà M (Eds) DAISIE, Th e Handbook of Alien Species in Europe. Heidelberg, Germany: Springer, 63–79.

Goldarazena A. (2011) First record of Thrips hawaiiensis (Morgan, 1913) (Thysanoptera: Thripidae), an Asian pest thrips in Spain. Bulletin OEPP/EPPO, 41(2): 170-173.

Gorsuch, C. S., G. T. Lee, and D. R. Alverson. 1989. Arthropod species collected from peach trees in South Carolina utilizing a whole-tree sampling method. J. Agric. Entomol. 6: 233-255.

Henry, T. J. and A. G. Wheeler, Jr. 1988. Family Miridae Hahn, 1833 (=Capsidae Burmeister, 1835), pp. 251-507. In T. J. Henry and R. C. Froeschner [eds.], Catalog of the Heteroptera, or true bugs, of Canada and the continental United States. Brill, New York.

Knight, H. H. 1941. The plant bugs, or Miridae, of Illinois. Bull. Ill. Nat. Hist. Surv. 22: 1-234.

Larsen K. and Vilhelmsen F. (1991) Clepsis peritana (Clemens, 1860) fundet I Danmark (Lepidoptera, Tortricidaae).- Lepidoptera, 4(1): 22-24.

Mizell, R. F. III and D. E. Schiffhauer. 1987. Trunk traps and overwintering predators in pecan orchards: Survey of species and emergence times. Fla. Entomol. 70: 238-244.

NUPPONEN, K., NUPPONEN, T, SAARELA, E. & SIPPOLA, L., 2003.– New records on Microlepidoptera from the western Mediterranean region Lepidoptera: Nepticulidae, Scythrididae, Tortricidae).– SHILAP Revta. lepid., 31 (123): 229-233.

Snodgrass, G. L. 1991. *Deraecoris* (sic) *nebulosus* (Heteroptera: Miridae): Little known predator in cotton in the Mississippi Delta. Fla. Entomol. 74: 340-344.

Snodgrass, G. L., T. J. Henry, and W. P. Scott. 1984. An annotated list of the Miridae (Heteroptera) found in the Yazoo-Mississippi Delta and associated areas in Arkansas and Louisiana. Proc. Entomol. Soc. Wash. 86: 845-860.

W. B. YEH, M. J. TSENG, N. T. CHANG, S. Y. WU, Y. S. TSAI (2014a) Development of Species-Specific Primers for Agronomical Thrips and Multiplex Assay for Quarantine Identification of Western Flower Thrips. J. Econ. Entomol. 107(5): 1728Ð1735

Wen-Bin Yeh, Cindy Sin-Ying Wu, Mei-Jung Tseng, Niann-Tai Chang, Wen-Ping Chen (2014b) Multiplex PCR for the Molecular identification of Economically Important Thrips. Formosan Entomol. 34: 211-225

Wheeler, A.G., Jr., B. R. Stinner, and T. J. Henry. 1975. Biology and nymphal stages of Deraeocoris nebulosus (Hemiptera: Miridae), a predator of arthropod pests on ornamentals. Ann. Entomol. Soc. Am. 68: 1063-1068.

Bodor J. 2010. Az ázsiai földibolha (Luperomorpha xanthodera Fairmare) megjelenése Magyarországon. [The first occurrence a flea beetle (Luperomorpha xanthodera Fairmare) in Hungary]. Növényvédelem 47 (3): 115–116

Beenen R., Roques A. 2010. Leaf and seed beetles (Coleoptera, Chrysomelidae). Chapter 8.3. BioRisk 4 (1): 267–292.

Cesari, M., Maistrello, L., Ganzerli, F., Dioli, P., Rebecchi, L. & Guidetti, R. 2015. A pest alien invasion in progress: potential pathways of origin of the brown marmorated stink bug Halyomorpha halys populations in Italy. Journal of Pest Science 88, 1–7.

Conti B., Raspi A. 2007. Prima segnalazione in Italia di Luperomorpha nigripennis Duvivier (Coleoptera: Chrysomelidae). [First record for Italy of Luperomorpha nigripennis Duvivier (Coleoptera: Chrysomelidae)]. Informatore Fitopatologico 57 (8/7): 51–52.

Del Bene G., Conti B. 2009. Notes on the biologyand ethology of Luperomorpha xanthodera, a flea beetle recently introduced into Europe. Bull. Insectol. 62 (1): 61–68.

Döberl M. 2010. Subfamily Alticinae Newman. p. 491–563. In: „Catalogue of Palaearctic Coleoptera. Vol. 6. Chrysomeloidea" (I. Löbl, A. Smetana, eds.). Apollo Books, Stenstrup, 924 pp.

Döberl M., Sprick P. 2009. Luperomorpha Weise, 1887 in Western Europe (Coleoptera, Chrysomelidae, Alticinae). Entomologische Blätter 105: 51–56.

Doguet S. 2008. Présence en France de Luperomorpha nigripennis Duvivier, 1892 (Col. Chrysomelidae, Alticinae). [The presence in France of Luperomorpha nigripennis Duvivier, 1892 (Col., Chrysomelidae, Alticinae)]. Le Coléoptériste 11 (1): 62–63.

Folmer O, M, Black WH, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome C oxidase subunit I from metazoan invertebrates. Molecular Marine Biology and Biotechnology 3: 294-299.

Geiser E. Bernhard M. 2011. Der Flohkäfer Luperomorpha xanthodera (Fairmaire, 1888) (Alticinae, Chrysomelidae) Erstnachweis für Österreich in einem Salzburger Garten. [The flea beatle Luperomorpha xanthodera (Fairmaire, 1888) first time found Austria, in a garden in Salzburg]. Newsletter der Salzburger Entomologischen Arbeitsgemeinschaft 3–4: 1–3.

Haye, T., Gariepy, T., Hoelmer, K., Rossi, J.-P., Streito, J.-C., Tassus, X. & Desneux, N. 2015. Range expansion of the invasive brown marmorated stinkbug, Halyomorpha halys: an increasing threat to field, fruit and vegetable crops worldwide. Journal of Pest Science, 88, 665-673.

Hebert PDN, Penton EH, Burns JM, Janzen DH, Hallwachs W (2004) Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly Astraptes fulgerator. Proceedings of the National Academy of Sciences of the United States of America 101: 14812-14817.

Hoebeke, E.R. & Carter, M.E. 2003. Halyomorpha halys (Stål) (Heteroptera: Pentatomidae): a polyphagous plant pest from Asia newly detected in North America. Proceedings of the Entomological Society of Washington, 105, 225–237.

Hoebeke, E.R. & Carter, M.E. 2003. Halyomorpha halys (Stål) (Heteroptera: Pentatomidae): a polyphagous plant pest from Asia newly detected in North America. Proceedings of the Entomological Society of Washington, 105, 225–237.

Jones, J.R. & Lambdin, P.L. 2009. New county and state records for Tennessee of an exotic pest, Halyomorpha halys (Hemiptera: Pentatomidae), with potential economic and ecological implications. Florida Entomologist, 92, 177–178.

Johnson C., Booth R.G. 2004. Luperomorpha xanthodera (Fairmaire): a new British flea beetle (Chrysomelidae) on Garden Centre Roses. The Coleopterist 13 (4): 81–86.

Kozłowski W. M., Legutowska H. (2014) The invasive flea beetle Luperomorpha xanthodera (Coleoptera: Chrysomelidae: Alticinae), potentially noxious to ornamental plants – first record in Poland. JOURNAL OF PLANT PROTECTION RESEARCH Vol. 54, No. 1, 106-107.

Lee, D.-H., Short, B.D., Joseph, S.V., Bergh, J.C., Leskey, T.C., 2013. Review of the Biology, Ecology, and Management of Halyomorpha halys (Hemiptera: Pentatomidae) in China, Japan, and the Republic of Korea. Environmental Entomology, 42(4), 627–641.

Leskey, T.C., Short, B.D., Butler, B.R. & Wright, S.E. 2012a. Impact of the Invasive Brown Marmorated Stink Bug, Halyomorpha halys (Stål), in Mid-Atlantic Tree Fruit Orchards in the United States: Case Studies of Commercial Management. Psyche: A Journal of Entomology, Article ID 535062, pp. 1-14. doi:10.1155/2012/535062

Leskey, T.C., Hamilton, G.C., Nielsen, A.L., Polk, D.F., Rodriguez-Saona, C., Bergh, J.C., Herbert, D.A., Kuhar, T.P., Pfeiffer, D., Dively, G.P., Hooks, C.R.R., Raupp, M.J., Shrewsbury, P.M., Krawczyk, G., Shearer, P.W., Whalen, J., Koplinka-Loehr, C., Myers, E., Inkley, D., Hoelmer, K.A., Lee, D.-H., Wright, S.E., 2012b. Pest Status of the Brown Marmorated Stink Bug, Halyomorpha halys in the USA. Outlooks on Pest Management, 23(5), 218–226.

Maistrello, L., Dioli, P., Vaccari, G., Nannini, R., Bortolotti, P., Caruso, S., Costi, E., Montermini, A., Casoli, L. & Bariselli, M. 2014. Primi rinvenimenti in Italia della cimice esotica Halyomorpha halys, una nuova minaccia per la frutticoltura. Atti Giornate Fitopatologiche, 1, 283–288.

Nielson M. W., Zack R. S., Poggi F., Nickel H. (2014) New leafhopper species of Jikradia from Mesoamerica with new records, revised key to species, distribution, origin, and checklist (Hemiptera: Cicadellidae: Coelidiinae: Teruliini). Rev. Biol. Trop. (Int. J. Trop. Biol. ISSN-0034-7744) Vol. 62 (4): 1375-1383

Reynaud P, Balme´s V & Pizzol K (2008) Thrips hawaiiensis (Morgan, 1913) (Thysanoptera: Thripidae), an Asian pest thrips now established in Europe. Bulletin OEPP / EPPO Bulletin 38 (1), 155–160.

FUNGI

Insofar as the monitoring of the presence of some categories of fungi of phytosanitary interest, we proceeded as follows:

- monitoring of the air through spore traps;
- monitoring of the water, using apples as bait.

Monitoring by spore capture

Aerobiology studies the sources and dispersion of airborne particles, with numerous possible applications in various sectors. One in which it may be of fundamental utility is in the protection of specific systems by the identification and monitoring of fungal parasites that pose a threat to agricultural crops and the identification of the optimal times in which to take measures to constrast them.

The tool that allows us to amass all this information is called a spore trap. It can be used for monitoring airborne spores in a qualitative sense, identifying the genus and species of the desired fungus, and quantitatively, permitting the counting of the units forming colonies as a proportion of the volume of air aspirated.

The instrument was therefore chosen to control for the possible dispersion of spores responsible for numerous pathologies linked to cultivation (airborne diseases). One example are the spores of blights like *Tilletia indica,* potentially carried by caryopsidae and infected straw packing material, or *Puccinia horiana,* transported by numerous plants and/or by cut chrysanthemums. In addition, the spores of *Plasmopara halstededii* are also of phytosanitary interest, carried by whole sunflower plants or their cut flowers, and *Alternaria* spp., *Pennicilium* sp. and /or *Monilinia* spp. Potentially present on rotting fruit.

The spore trap consists of a metal structure with a rotating drum inside it, on whose surface there is a transparent plastic silicone strip, which the airborne spores stick to.

The silicone strip on the cylindrical drum must be replaced every seven days, after which the spores are observed with an optical microscope. The silicone strips, collected weekly, were sent to the laboratory in specific boxes for analysis.

Slides were prepared for analysis by cutting small pieces from the strips, using gel to fix them, and then viewed at a magnification of 400X.

Monitoring the air thus permitted the identification of saprophytic and ubiquitous fungi of scarce interest in phytosanitary terms, belonging to the genera *Cladosporium, Epicoccum, Drechslera, Curvularia, Alternaria, Pestalotiopsis.*

Monitoring of waters (*baiting*)

To monitor the presence of zoospores and spores in the waters on and around the site, it was decided to employ a baiting technique which has been indicated by numerous studies as a simple but efficient method to identify and measure the growth of some genera of fungi. The method takes advantage of the natural effectiveness of bait consisting of pieces of fruit (e.g. pieces of apple or pear) in attracting fungal spores, and in particolar allows the isolation of spores of Phytophthora spp. and Fusarium spp.

We thus used pieces of apple, which were placed in a mesh cage and submerged in the canal that runs along the outer perimeter of the Expo exhibition grounds.

After seven days (Protocol FI) the cages were collected and sent to the laboratory for tests to isolate the oomycetic fungi.

It was decided not to leave the traps in the water for more than seven days to prevent the fruit's rotting and/or contamination by saprophytes and/or parasites whose presence would complicate the laboratory analysis.

The procedure is known as 'baiting', with the bait consisting of pieces of apple, to which spores that could be carried by visitors' shoes will adhere. The technique allows the isolation of spores of *Phytophthora* spp. and *Fusarium* spp.

Laboratory activities

In the lab, fragments were removed from the surface of the apples, placed on *Parp Agar,* a selective substrate, and then purified on *Cornmeal Agar*.

These substrates contain antibiotics that inhibit bacterial growth and thus facilitate the culturing of pure fungus.

The fungi obtained were then subjected to molecular analysis: their DNA was extracted and put through gene amplification using specific primers for the ITS portion (White et al., 1990).

The subsequent sequencing and biostatistical analysis permitted the identification of several species of fungi.

Results

Microscopic observation of the fragments and the isolation and genetic amplification of the fragments of fungi collected on the bait permitted the identification of several different organisms, none of which are of interest in phytosanitary terms.

In any case, we can confirm the identification of:

Pythium dissotocum Drechsler 1930 (Oomycetes, Pythiales)

Phytium dissotocum is a widespread fungus and has been found in many countries, but not in Italy (source: Systematic Mycology and Microbiology Laboratory Fungus-Host Distributions Database. U.S. Department of Agriculture, Agricultural Research Service) It is found in seeds and seedlings and in the roots of numerous plants. It causes root necrosis and kills seedlings.

It generally infects (with root necrosis) *Beta vulgaris*, *Carica papaya*, *Citrus nobilis*, *Fragaria×ananassa*, *Glycine max*, *Lactuca sativa*, *Lupinus digitatus Australia* and *Medicago sativa* (and, isolated from roots), *Oryza sativa*, *Phaseolus vulgaris* (or with root rot), *Spinacia oleracea,Triticumaestivum and Zea mays.*

Geotrichum candidum Link 1809 (*Saccharomycetes, Saccharomycetales*)

This fungus is a pathogen for European plants.

It causes acid rot in citrus fruit, tomatoes, carrots and other vegetables.

***Phytophthora lacustris* Brasier**

Phytophthora lacustris Brasier, Cacciola, Nechwatal, Jung & Bakonyi, reported in the literature in numerous forest, fluvial and lacustral ecosystems.

Present in Italy, the United Kingdom, Denmark, Germany, Poland, Hungary, New Zealand and China.

It has been isolated and identified on roots of *Fraxinus excelsior, Prunus* spp. *Quecus petraea* and *Salix alba* and is considered an opportunistic pathogen.

Phytopythium litorale (Nechw.) Abad, de Cock, Bala, Robideau, Lodhi & Lévesque 2014

This fungus is closely associated with waterways and is found in Germany, North America and Australia.

It is a pathogen for *Xanthorrehoea preissii*, *Banksia* spp. and *Casuarina obesa*.

The table below contains all the results of the laboratory analysis for the sample analyzed.

Microscopic observations of the silicone strips and spore identification: AndreaTantardini
Isolation in selective substrate (PARP) from apple bait: Andrea Tantardini
Molecular sequencing of the colonies cultured in isolation: Dr. Francesca Gaffuri

NUMERO CAMPIONE	TIPOLOGIA E DATA PRELIEVO	ORGANISMI RISCONTRATI
15_0056.1	Captaspore 09.02.15	*Alternaria* sp. , *Diplodia*,
15_0056.2	Baiting	Assenza oomiceti
15_0060.1	Captaspore 16.2.15	Assenza spore
15_0060.2	Baiting	***Pythium littorale***
15_0060.3	Baiting	Assenza oomiceti
15_0076.1	Captaspore 23.2.15	Assenza spore
15_0076.2	Baiting	*Alternaria alternata*
15_0087.1	Captaspore 02.03.215	*Leptosphaeriasp. Cladosporium sp.*
15_0087.2	Baiting	*Mucor* sp., ***Galactomyces geotrichum***
15_0087.3	Baiting	***Pythium dissotocum***
15_0091.1	Captaspore 09.03.15	*Cladosporium* sp. *Epicoccum* sp. *Alternaria* sp.
15_0091.2	Baiting	Assenza oomiceti
15_0109.1	Captaspore 16.03.15	*Alternaria* sp.
15_0109.2	Baiting	***Pythium dissotocum***
15_0109.3	Baiting	***Pythium dissotocum***
15_0109.4	Baiting	***Pythium dissotocum***
15_0131.1	Captaspore 23.03.15	*Sphaeropsis* sp.
15_0131.2	Baiting	Assenza miceti
15_0143.1	Captaspore 30.03.15	*Alternaria* sp.
15_0143.2	Baiting	*Trichoderma sp.*
15_0143.3	Baiting	Assenza miceti
15_0161.1	Captaspore 07.04.15	*Alternaria* sp. *Cladosporium* sp.
15_0161.1	Baiting	*Clonostachys rosea*
15_0161.1	Baiting	*Candida sp.*
15_0161.1	Baiting	***Galactomyces geotrichum***
15_0178.1	Captaspore 07.04.15	*Diplodia sp. Alternaria sp. Cladosporium* sp.
15_0178.2	Baiting	*Mucor* sp.
15_0178.3	Baiting	*Mucor* sp.
15_0178.4	Baiting	Assenza miceti
15_0222.1	Captaspore 20.04.15	*Alternaria* sp. *Tilletia* sp., *Cladosporium* sp.
15_0222.2	Baiting	*Trichoderma asperellum*
15_0222.3	Baiting	Assenza miceti
15_0222.4	Baiting	Assenza miceti
15_0223.1	Captaspore 27.04.15	*Alternaria* sp. *Tilletia* sp.
15_0223.2	Baiting	*Mucor* sp.
15_0223.3	Baiting	*Mucor* sp.
15_0223.4	Baiting	*Mucor* sp.
15_0270.1	Captaspore 04.05.15	*Alternaria alternata*, *Cladosporium* sp.
15_0270.2	Baiting	Assenza miceti
15_0270.3	Baiting	*Fusarium* sp.
15_0270.4	Baiting	*Mucor* sp.
15_0278.1	Captaspore 11.05.15	*Alternaria alternata*, *Diplodia* sp., *Cladosporium* sp.
15_0278.2	Baiting	Assenza miceti
15_0278.3	Baiting	*Fusarium tricinctum*
15_0278.4	Baiting	***Phytopthtora lacustris***
15_0278.5	Baiting	***Pythium dissotocum***
15_0278.6	Baiting	*Trichoderma koningopsis*
15_0300.1	Captaspore 18.05.15	*Alternaria alternata*, *Epicoccum* sp.
15_0300.2	Baiting	Assenza di miceti
15_0300.3	Baiting	***Galactomyces* sp**.
15_0300.4	Baiting	Assenza di miceti
15_0300.5	Baiting	Assenza di miceti
15_0300.6	Baiting	Assenza di miceti
15_0301.1	Captaspore 25.05.15	*Epicoccum* sp., *Diplodia* sp., *Cladosporium* sp.
15_0301.2	Baiting	Assenza miceti
15_0301.3	Baiting	*Fusarium avenacearum*
15_0301.4	Baiting	*Fusarium sp.*
15_0301.5	Baiting	Assenza miceti

NUMERO CAMPIONE	TIPOLOGIA E DATA PRELIEVO	RISULTATO *organismi riscontrati*
15_0301.6	Baiting	*Fusarium* sp.
15_0301.7	Baiting	Assenza miceti
15_0398.1	Captaspore 01.6.15	Assenza di spore fungine
15_0398.2	Baiting	Pythium sp.
15_0398.3	Baiting	*Fusarium solani*
15_0398.4	Baiting	Assenza miceti
15_0398.5	Baiting	*Fusarium avenacearum*
15_0398.6	Baiting	Assenza miceti
15_0398.7	Baiting	Assenza miceti
15_0399.1	Captaspore 08.6.15	*Cladosporium* sp. *Tilletia* sp. *Epicoccum* sp.
15_0399.2	Baiting	*Trichoderma atroviride*
15_0399.3	Baiting	*Galactomyces candida*
15_0399.4	Baiting	Assenza miceti
15_0399.5	Baiting	*Fusarium oxysporum*
15_0399.6	Baiting	Assenza miceti
15_0399.7	Baiting	Assenza miceti
15_0400.1	Captaspore 15.6.15	*Alternaria alternata, Cladosporium sp., Drechslera sp.* *Curvularia* sp.
15_0400.2	Baiting	*Colletotrichum fiorinae*
15_0400.3	Baiting	*Colletotrichum fiorinae*
15_0400.4	Baiting	*Colletotrichum fiorinae*
15_0400.5	Baiting	-
15_0400.6	Baiting	-
15_0400.7	Baiting	-
15_0401.1	Captaspore 22.6.15	*Alternaria alternata, Epicoccum sp., Cladosporium sp., Sphaeropsis* sp.
15_0401.2	Baiting	-
15_0401.3	Baiting	-
15_0401.4	Baiting	*-Colletotrichum acutatum*
15_0401.5	Baiting	*-Colletotrichum acutatum*
15_0401.6	Baiting	*-Colletorichum fiorinae*
15_0401.7	Baiting	-
15_0402.1	Captaspore 29.6.15	*Alternaria alternata, Epicoccum sp., Cladosporium sp.* *Oospore di Plasmopara viticola*
15_0418	Captaspore 07.07.15	*Alternaria alternata, Epicoccum sp., Cladosporium sp.* *Oospore di Plasmopara viticola*
15_0434.1	Baiting	*Glomerella fiorinae*
15_0434.2	Baiting	*Glomerella fiorinae*
15_0434.3	Baiting	*Colletorichum acutatum*
15_0434.4	Baiting	-
15_0445	Captaspore 14.07.15	*Alternaria alternata,Epicoccum* sp.,*Cladosporium* sp.
15-0552	Captaspore 20.07.15	Trichotecium roseum, *Alternaria alternata*, *Cladosporium* sp.,*Epicoccum* sp.,Oospore di Plasmopara .
15_0553	Captaspore 27.07.15	*Cladosporium* sp.,Oospore di Plasmopara, *Alternaria alternata, Epiccoccum sp.*
15_0554.1 Mela 1_3007	Baiting 30.07.15	*Colletotrichum fiorinae*
15_0554.2 Mela 2_3007	Baiting 30.07.15	-
15_0554.3 Mela 3_3007	Baiting 30.07.15	-
15_0554.4 Mela 4_3007	Baiting 30.07.15	*C.acutatum*
15_0554.5 Mela 5_3007	Baiting 30.07.15	*C.fiorinae*
15_0555	Captaspore 03/08/2015	*Epicoccum Sp., Trichotecium roseum, Alternaria alternata*
15_0556	Captaspore 10/08/2015	*Alternaria alternata,Cercospora sp.,Oospore di Plasmopara, Cladosporium,Epicoccum*.
15_0557	Captaspore17/08/2015	*Alternaria alternata, Cladosporium,Epicoccum*
15_0558	Captaspore25/08/2015	*Alternaria sp.*
15_0559.1 -Mela 1_2808	Baiting 28.08.15	*C.acutatum*
15_0559.2 Mela 2_2808	Baiting 28.08.15	*C.acutatum*
15_0559.3 Mela 3_2808	Baiting 28.08.15	*C.fiorinae*

NUMERO CAMPIONE	TIPOLOGIA E DATA PRELIEVO	RISULTATO *organismi riscontrati*
15_0559.4	Baiting 28.08.15	-
15_0559.5 Mela 5_2808	Baiting 28.08.15	*C.fiorinae*
15_0559.6 Mela 6-2808	Baiting 28.08.15	-
15_0581	Nastro Captaspore 31/08/2015	*Alternaria sp.,Epicoccum sp.Ooospore di Plasmopara,Cladosporium*
15_0602	Nastro Captaspore 07/09/2015	*Alternaria sp., Epicoccum sp.,Drechslera sp.,Cladospoirum sp.,*
15_0627.1	Baiting 17.9	-
15_0627.2	Baiting 17.9	- ***Pythium dissotocum***
15_0627.3	Baiting 17.9	- *C.acutatum*
15_0627.4	Baiting 17.9	-
15_0627.5	Baiting 17.9	*Pythium dissotocum*
15_0627.6	Baiting 17.9	-
15_0630	Arbutus unedo per Phythopthora	*Presenza di Pestalotiopsis sp.*
15_0916	Nastro captaspore 15/09/2015	*Alternaria sp.,Epicoccum sp.,Puccinia sp.,*
15_0917	Nastro captaspore 21/09/2015	*Alternaria sp.,Cladosporium sp.,Drechslera sp.,Epicoccum sp.Pestalotiopsis sp.*
15_0918	Nastro captaspore 28/09/2015	*Alternaria sp., Epicoccum sp.,Drechslera sp., Diplodia sp.,Cladosporium sp.*
15_0919	Nastro captaspore 05/10/2015	*Epicoccum sp.*
15_0920.1	Baiting 05.10.2015 1	***Pythium dissotocum***
15_0920.2	Baiting 05.10.2015 2	-
15_0920.3	Baiting 05.10.2015 3	***Pythium dissotocum***
15_0920.4	Baiting 05.10.2015 4	**-**
15_0920.5	Baiting 05.10.2015 5	***Pythium dissotocum***
15_0920.6	Baiting 05.10.2015 6	-
15_0921	Nastro captaspore 13/10/2015	*Epicoccum sp., Alternaria sp., Cladosporium sp.*
15_0922	Nastro captaspore 19/10/2015	*Alternaria sp, Epicoccum sp.*
15_0923.1	Baiting 22.10.2015 1	*C.acutatum*
15_0923.2	Baiting 22.10.2015 2	-
15_0923.3	Baiting 22.10.2015 3	*C.acutatum*
15_0922.4	Baiting 22.10.2015 4	*C.acutatum*
15_0923.5	Baiting 22.10.2015 5	***Geotrichum***
15_0923.6	Baiting 22.10.2015 6	-
15_0924	Nastro captaspore 26/10/2015	*Epicoccum Sp.,Alternaria sp.*
15_0925	Nastro captaspore 02/11/2015	*Epicoccum Sp.,Alternaria sp.*

Bibliografia

Systematic Mycology and Microbiology Laboratory Fungus-Host Distributions Database. December 14, 2015 U.S. Department of Agriculture, Agricultural Research Service.

De Hoog, G.S., Smith, M.T., and Gueho, E. 1986. A revision of the genus *Geotrichum* and its teleomorphs. Stud. Mycol. 29: 1-131.

Brasier, C. M., Hamm, P. B., Hansen, E. M. 1993. Cultural characteristics, protein patterns and unusual mating behavior of *Phytophthora gonapodyides* isolates from Britain and North America. Mycol. Res. 97:1287-1298.

Brasier, C. M., Cooke, D. E. L., Duncan, J. M., and Hansen, E. M. 2003. Multiple new phenotypic taxa from trees and riparian ecosystems in *Phytophthora gonapodyides-P. megasperma* ITS Clade 6, which tend to be high-temperature tolerant and either inbreeding or sterile. Mycol. Res. 107:277-290.

Hansen, E. M., Reeser, P. W., Sutton, W. 2012. *Phytophthora borealis* and *Phytophthora riparia*, new species in *Phytophthora* ITS Clade 6. Mycologia: In Press. doi:10.3852/11-349

Nechwatal, J., Bakonyi, J., Cacciola, S.O., Cooke, D. E. L., Jung, T., Nagy, Z. A., Vannini, A., Vettraino, A. M., Brasier, C. M. 2012. The morphology, behaviour and molecular phylogeny of *Phytophthora* taxon Salixsoil and its redesignation as *Phytophthora lacustris* sp. nov. Plant Pathology Doi:10.1111/j.1365-3059.2012.02638.x

Reeser, P. W., Hansen, E. M., Sutton, W., Remigi, P., Adams, G. C. 2011. *Phytophthora* species in forest streams in Oregon and Alaska. Mycologia 103: 22-35

De Cock, A.W.A.M., Lodhi, A.M., Rintoul, T.L., Bala, K., Robideau, G.P., Abad, Z.G., Coffey, M.D., Shahzad, S., and Levesque, C.A. 2014. *Phytopythium*: molecular phylogeny and systematics. Persoonia 34: 25-39.

Parkunan, V., and Ji, P. 2013. Isolation of *Pythium litorale* from irrigation ponds used for vegetable production and its pathogenicity on squash. Canad. J. Pl. Pathol. 35: 415-423.

Uzuhashi, S., Tojo, M., and Kakishima, M. 2010. Phylogeny of the genus *Pythium* and description of new genera. Mycoscience 51: 337-365.

XIV CONGRESSO NAZIONALE A.I.A. "Trent'anni di Aerobiologia in Italia": Minoprio, 24-26 settembre 2015 D. Magyar (Budapest) *morfologia delle spore fungine - Metodi di campionamento delle spore fungine - Ascomiceti, Basidiomiceti e Funghi imperfetti*

ERWIN & RIBEIRO 1996 Phythopthora disease World wide APS PRESS - Baiting tecniques 14.01.2016 A

NEMATODES

The decoration of several pavilions called for the introduction of various plants (e.g. *Musa* spp., *Coffea* spp., etc.) or edible parts of plants (tubers like *Solanum* spp., *Hypomea* spp.) which could potentially be hosts for phytoparasitic nematodes of the sort described above, as well as plants shavings (conifer bark) used as ground cover for the organic composted soil of the plant and flower beds present on the premises.
Particular attention was placed on controlling for the presence of *B. xylophilus*, with the measures consisting of sampling and laboratory nematological analysis of probable host material (11 samples) and surveillance for its vector, *Monochamus galloprovincialis,* described above, as well as through the location of a control point composed of indicator plants (Gen. *Pinus* spp.), which was periodically checked to verify the possible presence of symptoms.
Periodic visual inspections were also carried out on host plants for the other PPN described, to check for symptoms of infestation; particular attention was paid to plants of the Musaceae (*Musa* spp.) and *Citrus* spp.
Laboratory analyses and inspection results indicated no possible infestations during the event.

VIRUSES AND PHYTOPLASMS

Monitoring of the various viruses that could potentially be found on the Expo grounds, distributed in line with the location of trees and other plants, focused mostly on Sharka (Plum Pox Virus), which is responsible for causing blisters on pitted fruit and CTV (Citrus tristeza virus), the pathogen of tristeza, which affects citrus trees. To check for the presence of these pathogens, we set up 2 control ponts at strategic locations, next to small groups of lemon trees, for CTV, and a single point, next to some plum trees, for Sharka. The “indicator plants” were marked as such with small signs and underwent periodic controls, during which samples were collected if symptoms were observed, and then sent to the laboratory for analysis. Every analysis carried out proved negative for the presence of the viruses in question.
Insofar as the phytoplasms, controls were performed for flavescence dorée, already common in Lombardy, which causes yellowing of grapevine leaves. The monitoring was carried out by locating an indicator point, which was a large wall of vines on the site. Visual inspections were always negative, making it unnecessary to collect samples and analyze them in the lab.

BACTERIA

The monitoring of possible introductions of noxious bacteria into the exhibition grounds was mostly focused on *Erwinia amylovora*, the bacteria that causes fire blight, which is already present in some areas of Lombardy, and *Xylella fastidiosa,* the fly responsible for the withering disease that is afflicting olive trees in the Apulia region and in France.
To monitor the presence of the bacteria responsible for fire blight, we located three strategic indicator spots, near small groups of Viburnum plants, which are hosts for *Erwinia*, and marked them with small cards. These were periodically checked and, when symptoms were found, samples were collected and sent to the lab for analysis.
Monitoring for *Xylella fastidiosa* was instead conducted by controlling and sampling all the olive and coffee plants found inside the exhibition grounds. In addition, we sampled plants of rosemary, myrtle, oleander, broom and cherry, which can also carry the bacteria, for a total of almost 400 plants. Details of the sampling are shown in the following table:

Data di prelievo	Pianta campionata	Codice campione	Tipologia campione	Luogo di prelievo
23/04/2015	Caffè	4374380/1/2	12 piante	Padiglione Brasile
			80 piante	Cluster Caffè
27/04/2015	Olivo	4373957	1 di 1 pianta	Padiglione Francia
27/04/2015	Olivo	4373958	1 di 1 pianta	Padiglione Francia
05/05/2015	Olivo	4373966	Pool di 2 piante su 5 totali	Bahrein
05/05/2015	Olivo	4373965	Pool di 3 piante su 5 totali	Bahrein
05/05/2015	Olivo	3021117	Pool di 15 piante	Marocco
05/05/2015	Oleandro	4095362	Pool di 20 piante	Marocco
05/05/2015	Olivo	3021118	1 di 1 pianta	Padiglione Save the Children
05/05/2015	Olivo	4373967	Pool di 5 piante su 5 totali	Iran
06/05/2015	Olivo	4373869	1 di 1 pianta	Padiglione Sicilia
06/05/2015	Olivo	4373872	Pool di 4 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373865	Pool di 6 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373858	Pool di 6 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373866	Pool di 6 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373859	Pool di 6 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373874	Pool di 6 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373864	Pool di 6 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373863	Pool di 7 piante su 46 totali	Collina Mediterranea
06/05/2015	Olivo	4373867	Pool di 7 piante su 14 totali	Collina Mediterranea retro slow food
06/05/2015	Olivo	4373873	Pool di 7 piante su 14 totali	Collina Mediterranea retro slow food
06/05/2015	Olivo	4373857	Pool di 3 piante su 13 totali	Hortus 3
06/05/2015	Olivo	4373856	Pool di 4 piante su 13 totali	Hortus 3
06/05/2015	Olivo	4373855	Pool di 3 piante su 13 totali	Hortus 3
06/05/2015	Olivo	4373854	Pool di 3 piante su 13 totali	Hortus 3
06/05/2015	Olivo	4373862	2 di 2 piante	Padiglione Birra Moretti
06/05/2015	Olivo	4373861	1 di 1 pianta	Padiglione U.E.
06/05/2015	Olivo	4373860	1 di 1 pianta	Padiglione ENEL
06/05/2015	Olivo	4373870	Pool di 5 piante su 5 totali	Padiglione Argentina
06/05/2015	Olivo	4373868	Pool di 5 piante su 5 totali	Parco Biodiversità
06/05/2015	Olivo	4094729	Pool di 4 piante su 4 totali	Padiglione Eco Pasta&Pizza
07/05/2015	Olivo	4373882	Pool di 3 su 3 totali	Parco Biodiversità (Leccino Provenienza Toscana PT/C9/4206)
07/05/2015	Olivo	4373871	1 di 1 pianta	Padiglione Emirati Arabi Uniti
07/05/2015	Olivo	4373876	Pool di 4 piante su 12 totali	Collina Mediterranea
07/05/2015	Olivo	4373875	Pool di 4 piante su 12 totali	Collina Mediterranea
Data di prelievo	**Pianta campionata**	**Codice campione**	**Tipologia campione**	**Luogo di prelievo**
07/05/2015	Olivo	4373877	Pool di 4 piante su 12 totali	Collina Mediterranea
07/05/2015	Olivo	4373878	Pool di 4 piante su 9 totali	Collina Mediterranea parte superiore

07/05/2015	Olivo	4373879	Pool di 5 piante su 9 totali	Collina Mediterranea parte superiore
07/05/2015	Caffè	4373881	Pianta di Caffè n. 12	Padiglione Brasile
04/06/2015	Olivo	4373959	Pool di 2 piante su 2 totali	Coppini Olea
04/06/2015	Olivo	4373960	Pool di 2 piante su 2 totali	San Pellegrino
04/06/2015	Olivo	4373961	Pool di 4 piante su 4 totali	Cluster riso
10/07/2015	Olivo	3020381	Pool di 1 pianta su 16 totali	Cluster Biomediterraneo 6 (Montenegro)
10/07/2015	Olivo	3020380	Pool di 2 piante su 16 totali	Cluster Biomediterraneo 5
10/07/2015	Olivo	3020379	Pool di 3 piante su 16 totali	Cluster Biomediterraneo 4 (Tunisia)
10/07/2015	Olivo	3020378	Pool di 3 piante su 16 totali	Cluster Biomediterraneo 1
10/07/2015	Olivo	3020377	Pool di 3 piante su 16 totali	Cluster Biomediterraneo 2
10/07/2015	Olivo	3020376	Pool di 4 piante su 16 totali	Cluster Biomediterraneo 3
14/07/2015	Olivo	4373921	Pool di 3 piante su 20 totali	Cluster Biomediterraneo 7 (Sicilia Mediterranean Market)
14/07/2015	Olivo	4373920	Pool di 2 piante su 20 totali	Cluster Biomediterraneo 8 (Palco)
14/07/2015	Olivo	4373919	Pool di 4 piante su 20 totali	Cluster Biomediterraneo 9 (San Marino)
14/07/2015	Olivo	4373918	Pool di 4 piante su 20 totali	Cluster Biomediterraneo 10 (Algeria)
14/07/2015	Olivo	4373917	Pool di 3 piante su 20 totali	Cluster Biomediterraneo 11 (Algeria)
14/07/2015	Olivo	4373916	Pool di 4 piante su 20 totali	Cluster Biomediterraneo 12 (Malta-Libano-Grecia)
14/07/2015	Olivo	4373915	1 di 1 pianta	Padiglione del biologico
29/07/2015	Caffè	4373902	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
29/07/2015	Caffè	4373903	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
29/07/2015	Caffè	4373904	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
29/07/2015	Caffè	4373905	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
29/07/2015	Caffè	4373906	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
29/07/2015	Caffè	4373907	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
29/07/2015	Caffè	4373963	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
29/07/2015	Caffè	4373964	Pool di 3 piante su 26 totali	Padiglione Colombia piano terra
Data di prelievo	**Pianta campionata**	**Codice campione**	**Tipologia campione**	**Luogo di prelievo**
29/07/2015	Caffè	4094716	Pool di 2 piante su 26 totali	Padiglione Colombia terrazza
06/08/2015	Olivo	4096420	1 di 1 pianta	Cluster Biomediterraneo

06/08/2015	Olivo	4096421	1 di 1 pianta	Cluster Biomediterraneo
06/08/2015	Olivo	4096422	1 di 1 pianta	Cluster Biomediterraneo
06/08/2015	Olivo	4096423	1 di 1 pianta	Cluster Biomediterraneo
06/08/2015	Olivo	4096424	1 di 1 pianta	Cluster Biomediterraneo
06/08/2015	Olivo	4096425	1 di 1 pianta	Cluster Biomediterraneo
06/08/2015	Olivo	4096426	1 di 1 pianta	Cluster Biomediterraneo
06/08/2015	Olivo	4096427	1 di 1 pianta	Cluster Biomediterraneo
06/08/2015	Olivo	4096428	1 di 1 pianta	Cluster Biomediterraneo
22/09/2015	Rosmarino	4094718	Pool di 2 piante su 5 totali	Parco biodiversità
22/09/2015	Rosmarino	4094719	Pool di 2 piante su 10 totali	Collina mediterranea
22/09/2015	Spartium	4094720	Pool di 2 piante su 10 totali	Collina mediterranea
22/09/2015	Ciliegio	4094721	Pool di 2 piante su 15 totali	Hortus fianco padiglione Oman
22/09/2015	Spartium	4094722	Pool di 2 piante su 10 totali	Rotonda vicino cluster caffè
22/09/2015	Mirto	4094723	Pool di 2 piante su 10 totali	Padiglione Emirati Arabi Uniti
22/09/2015	Mirto	4094724	Pool di 2 piante su 10 totali	Cluster Biomediterraneo
22/09/2015	Ciliegio	4094725	Pool di 2 piante su 10 totali	Hortus fianco padiglione Turchia
28/09/2015	Olivo	3020476	Pool di 3 piante su 6 totali	Area UN, retro padiglione Cina
28/09/2015	Olivo	3020477	Pool di 3 piante su 6 totali	Area UN, retro padiglione Cina
05/10/2015	Olivo	3020405	Pool di 2 piante su 5 totali	Parco biodiversità (Biomarket)
05/10/2015	Olivo	3020406	Pool di 3 piante su 5 totali	Parco biodiversità (Biomarket)
05/10/2015	Olivo	3020407	Pool di 3 piante su 7 totali	Parco biodiversità (Teatro)
05/10/2015	Olivo	3020408	Pool di 4 piante su 7 totali	Parco biodiversità (Teatro)
14/10/2015	Caffè	4096584	1 pianta di 10 totali	Padiglione Angola (Terrazzo)
14/10/2015	Caffè	4096586	1 pianta di 10 totali	Padiglione Angola (Terrazzo)
14/10/2015	Caffè	4096587	1 pianta di 10 totali	Padiglione Angola (Terrazzo, ultimo grupppo di 3 piante a sinistra)

None of the samples were found to be positive for bacteriosis.

GASTROPODS

We also monitored for the possible introduction of *Pomacea* sp., a genus of freshwater aquatic snails that pose an extreme threat to the cultivation of rice, which have not yet been found in Italy. The monitoring was made necessary by the presence of some Clusters and Pavilions whose décor included vats or pools dedicated to the cultivation of rice. During the event, visual inspections of the rice plants were conducted to check for the presence of snail eggs, which are highly visible due to their reddish color. The results of all the visual inspections were negative.

STAFF

The monitoring plan's design and its revision in progress in response to the assessment of a constant phytosanitary risk was entrusted to the Head of the "Phytosanitary Surveillance Team". She availed herself of the collaboration of two specifically trained technical resources and an inspector from the plant health service.

Important support was also provided by the Director of the phytosanitary laboratory and the entire technical staff.

In addition to carrying out the monitoring of the exhibition grounds, technical personnel were also employed in the reinforced surveillance dedicated to the areas adjacent to the exhibition grounds.

The table below details the days of activity dedicated by each technical resource to the monitoring project:

NAME	Total days dedicated	Days on site
Mariangela Ciampitti	120	36
Francesca Siena	90	8
Chiara Patti	233	54
Emma Zanotti	237	58
Laboratory personnel	n.a.	16

TRAINING OF EXHIBITION SITE GROUNDSKEEPERS

Personnel from the plant health service conducted two training sessions for the Expo groundskeepers to inform them of the purposes of the monitoring program and to solitict their cooperation. The meetings were held on 10th April 2015 at the ERSAF offices and on 7th July 2015 on site at the Expo 2015 residential facility (*campo base*).

DOCUMENTATION OF REFERENCE

http://ec.europa.eu/food/plant/plant_health_biosafety/europhyt/index_en.htm
https://www.eppo.int/QUARANTINE/Alert_List/alert_list.htm
http://www.aphis.usda.gov/wps/portal/aphis/home/
http://www.cabi.org/
http://www.efsa.europa.eu/en/panels/plh.htm
Erwin and Ribeiro 1996. Isolation by baiting with selective host, Chapter 2, pag. 18, Phytophthora diseases worldwide
Handbook of quarantine pest for Europe (CABI) 1998 2nd edition
ISPM FAO - INTERNATIONAL STANDARDS FOR PHYTOSANITARY MEASURES 1 to 29 (2007 edition)
ISPM No. 6 (1997) Guidelines for surveillance

ANNEXES

EXPO MILANO 2015

Regione Lombardia
Servizio Fitosanitario

Monitoraggio fitosanitario

Legenda

Trappole

- Trappola popillia
- Prisma Agrilus
- Cromotropica gialla
- Cromotropica blu
- Trappola Multifunnel
- Trappola Planotrap
- Pagoda/Delta trap
- Secchiello
- Rebel bianca
- Rebel amarillo
- Cromotropica blu tripidi
- Cromotropica gialla tripidi
- Cromotropica Anthonomus
- Captaspore
- Mele esca
- Pianta spia
- Rebel amarillo Rhagoletis
- Sesia

Riso 37 H10
Bangladesh, Cambogia, Myanmar, Repubblica Democratica Popolare del Laos, Sierra Leone, Sasmati Pavilion

Cacao e Cioccolato 45 I12
Camerun, Costa D'Avorio, Cuba, Gabon, Ghana, Sao Tomé & Principe

Caffè 58 H14
Burundi, El Salvador, Etiopia, Guatemala, Kenya, Repubblica Dominicana, Ruanda, Timor Est, Uganda, Yemen

Frutta e Legumi 61 G15
Benin, Gambia, Guinea, Guinea Equatoriale, Kirghizistan, Repubblica Democratica del Congo, Sri Lanka, Uzbekistan, Zambia

Spezie 64 G16
Afghanistan, Brunei Darussalam, Repubblica Unita della Tanzania, Vanuatu - Pacific Islands Forum (Isole Marshall, Isole Salomone, Kiribati, Nauru, Niue, Papua Nuova Guinea, Samoa, Tonga, Tuvalu)

Bio-Mediterraneo 97 D21
Albania, Algerie, Egitto, Grecia, Libano, Malta, Montenegro, San Marino, Serbia, Tunisia; World Expo Museum

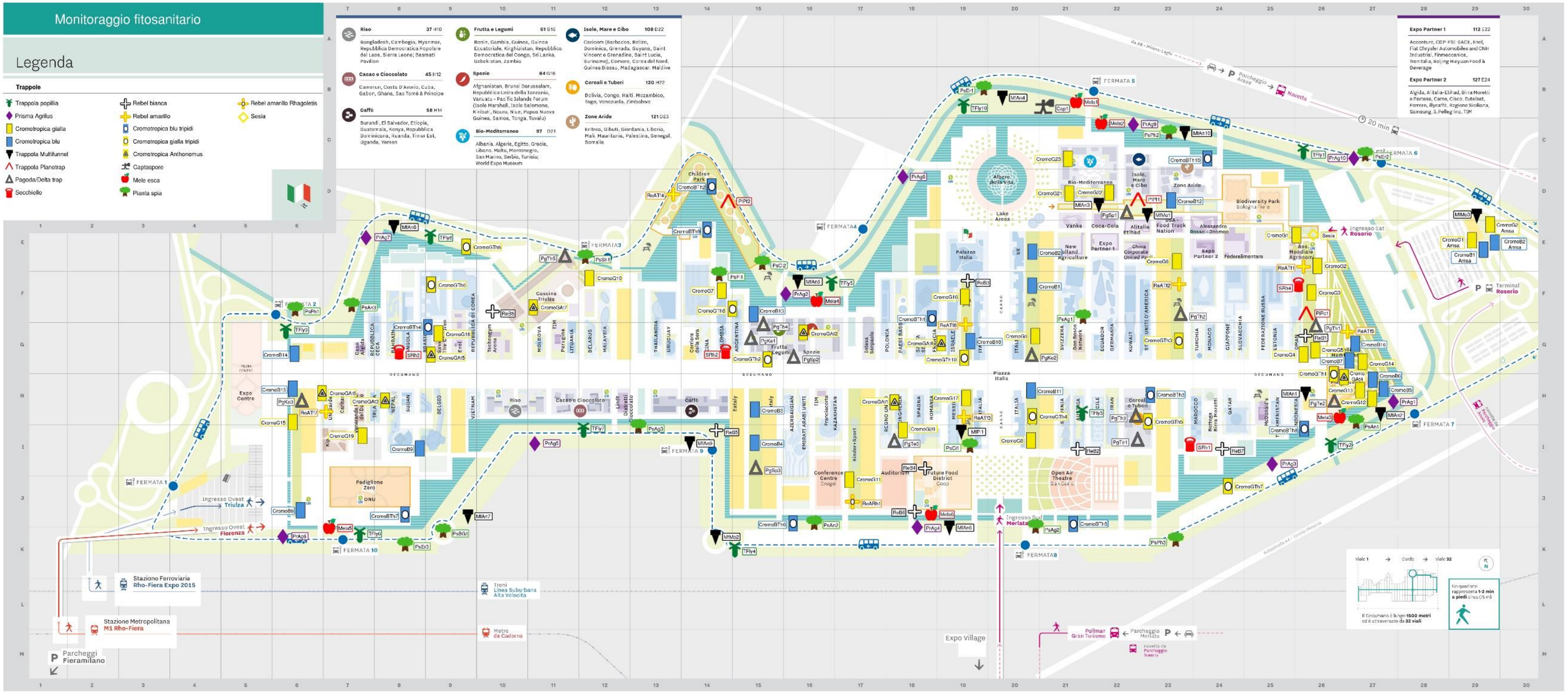
Isole, Mare e Cibo 108 D22
Caricom (Barbados, Belize, Dominica, Grenada, Guyana, Saint Vincent e Grenadine, Saint Lucia, Suriname), Comore, Corea del Nord, Guinea Bissau, Madagascar, Maldive

Cereali e Tuberi 120 H22
Bolivia, Congo, Haiti, Mozambico, Togo, Venezuela, Zimbabwe

Zone Aride 121 D23
Eritrea, Gibuti, Giordania, Liberia, Mali, Mauritania, Palestina, Senegal, Somalia

Expo Partner 1 112 E22
Accenture, CDP-FSI-SACE, Enel, Fiat Chrysler Automobiles and CNH Industrial, Finmeccanica, Trenitalia, Beijing Haiyan Food & Beverage

Expo Partner 2 127 E24
Algida, Alitalia-Etihad, Birra Moretti e Peroni, Cama, Cisco, Eutelsat, Ferrero, illycaffè, Regione Siciliana, Samsung, S.Pellegrino, TIM



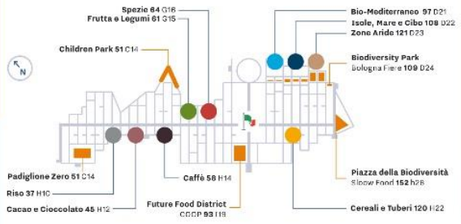
Partecipanti, Aree e Servizi – Ordine numerico

0 Padiglione Zero J 7	10 Caritas Internationalis H 7	20 Nepal H 8	30 Belgian Food & Events H 9	40 Perugina G 11	50 Uruguay G 13	60 Argentina G 15	70 Azerbaij. Garden of Biodiversity H 16	80 Polonia G 18	90 Israele G 19	100 Magnum Pleasure Store H 21	110 Coca-Cola E 22	120 Cluster Cereali e Tuberi H 22	130 Crucianì C F 24	140 Street Food 9 vd. retro I 24	150 Indonesia H 26
1 Expo Centre H 5	11 Van. Fabbr. del Duomo H 7	21 Sudan H 8	31 Belgio H 9	41 Lituania G 11	51 **Children Park** C 14	61 Cluster Frutta e Legumi G 15	71 Conference Centre Orogel I 16	81 Paesi Bassi G 18	91 **Italia** G 19	101 Area di Servizio G1 G 21	111 Alitalia Etihad E 22	121 Cluster Zona Aride D 23	131 Turchia G 24	141 Street Food 10 vd. retro I 24	151 Collina Mediterranea G 26
2 Area Self Intesa Sanpaolo G 6	12 Irlanda H 7	22 Street Food 8 vd. retro F 8	32 Area di Servizio B2 H 9	42 Belarus G 12	52 Street Food 6 vd. retro F 14	62 Kenya F 15	72 Street Food 7 vd. retro J 16	82 Santa Sede G 18	92 Messico H 19	102 Svizzera G 21	112 Expo Partner 1 E 22	122 Usa Food Truck Nation H 23	132 Monaco G 24	142 Ass. Mond. Agronomi F 25	152 **Piazza della Biodiversità**
3 Area di Servizio A H 6	13 Street Food 1 vd. retro H 7	23 Juice Bar J 8	33 Vietnam H 9	43 Malaysia G 12	53 Switch EXPerience G 14	63 Grom F 15	73 Filiale Intesa Sanpaolo G 17	83 Francia G 18	93 **Future Food Dist. Coop** I 19	103 Don Bosco Network G 21	113 China Corporate Un. Pav. E 22	123 Stati Uniti d'America G 23	133 Giappone G 24	143 Piazza Eventi Expo 3 F 25	**Slow Food** H 26
4 OVS G 7	14 Ireland "Bandstand" I 7	24 Maria Marinoni E 9	34 Belgium "A place to be" H 9	44 Area di Servizio C1 G 12	54 Street Food 6 vd. retro G 14	64 Cluster Spezie G 16	74 Kinder + Sport H 17	84 Regno Unito H 18	94 Lake Arena D 20	104 Area di Servizio G2 H 21	114 Piazza Expo G 22	124 Area di Servizio H1 G 23	134 Slovacchia G 24	144 Federazione Russa G 25	
5 Excelsior Milano G 7	15 La Piada & Le Bolle J 7	25 Brasile G 9	35 Dispensa Emilia E 10	45 Cluster Cacao e Cioccolato H 12	55 Cina G 14	65 Azerbaigian H 16	75 Area di Servizio F2 H 17	85 Ungheria H 18	95 Unione Europea E 20	105 Slovenia H 21	115 Ecuador G 22	125 Area di Servizio H2 H 23	135 Marocco H 24	145 Estonia G 25	
6 Casa Algida G 7	16 Davide Oldani J 7	26 Street Food 4 vd. retro G 9	36 Technogym Arena G 10	46 Lindt H 12	56 Corriere della Sera G 14	66 Emirati Arabi Uniti H 16	76 Sky I 17	86 Spagna H 18	96 Street Food 9 vd. retro C 21	106 Austria H 21	116 Germania G 22	126 Alessandro Rosso - Joomoo E 24	136 Bottega Birra Moretti H 24	146 McDonald's H 25	
7 Repubblica Ceca C 7	17 Street Food 2 vd. retro F 8	27 Save the Children G 9	37 Cluster Riso H 10	47 Distretti Cioccolato H 12	57 Colombia G 14	67 Franciacorta H 18	77 Case Ferrarini D 18	87 Romania H 18	97 Cluster Bio-Mediterraneo D 21	107 Open Air Theatre San Carlo J 21	117 Kuwait G 22	127 Expo Partner 2 E 24	137 Qatar	147 Turkmenistan H 25	
8 Kip - International School H 7	18 Bahrain G 8	28 Enel G 9	38 Cascina Triulza F 11	48 Beretta 1812 G 13	58 Cluster Caffè H 14	68 Kazakhstan H 16	78 Area Eventi Pad. Italia E 18	88 Coca-Cola Skydeck H 18	98 Auditorium I 18	108 Cluster Isole, Mare e Cibo D 22	118 Cile H 22	128 Federalimentare E 24	138 Qatar "Oryx Theatre" H 24	148 Juice Bar I 25	
9 UN Garden H 7	19 Angola G 8	29 Repubblica di Corea G 9	39 Moldova G 11	49 Thailandia G 13	59 Caffarel F 15	69 Piazza Eventi Expo 1 H 16	79 Place Stendhal G 18	89 Auditorium I 18	99 New Holland Agriculture E 21	109 **Biodiv. Park Bologna Fiere** D 24	119 Iran H 22	129 Piazza Eventi Expo 2 F 24	139 MSC Area H 24	149 Oman G 26	

Cluster e Aree Tematiche

Le Aree Tematiche

Le **Aree Tematiche** sono i luoghi fisici che interpretano il Tema **Nutrire il Pianeta, Energia per la Vita** attraverso percorsi espositivi ed elementi attrattivi coinvolgono direttamente i visitatori più piccoli e più grandi e diventano l'occasione per un'esperienza sensoriale ed educativa. Sono sviluppate da prestigiosi curatori internazionali e si trovano agli ingressi principali e in altri punti chiave del Sito Espositivo.



I Cluster

I Cluster sono una delle novità di Expo Milano 2015: i Paesi sono raggruppati secondo identità tematiche e filiere alimentari. In questo modo il tema **Nutrire il Pianeta, Energia per la Vita** viene sviluppato tenendo concreto lo spirito di questi spazi espositivi che rappresentano condivisione, dialogo e cooperazione.

I Cluster sono caratterizzati da aree comuni, che sviluppano attraverso spazi funzionali: (mercato, mostra, eventi, degustazioni) la filiera alimentare.

Il Cardo – Padiglione Italia

ITALIA
Vivaio Italia

Il Padiglione Italia mette in mostra le eccellenze italiane: la cultura e le tradizioni nazionali legate al cibo e all'alimentazione, caratterizzate dall'alta qualità delle materie prime e dei prodotti finali.

Si compone del Palazzo Italia, dei quattro edifici sul Cardo e della Lake Arena, per un totale di 14.000 metri quadri.

Padiglione Italia

a Le Casa dell'Identità Italiana Peck a Palazzo Italia.
b Gitterio
c Women for Expo
d Croce Rossa Italiana
e Calabria
Emilia Romagna
f Irpinia
Trentino
g Spazio ristoratori
h Ecco Pizza & Pasta
i Lombardia
j Gramaroldo
k Lavazza
l Coppini Arte Olearia
m Coldiretti
n Copagri
o Padiglione del Vino
p Piacenza
q Sicilia
r Alto Adige - Südtirol
s Martini & Rossi
t Birrificio Angelo Poretti
u Acqua S. Pellegrino
v Fch Food - Confindustria
z Gelateria Rigoletto

Campagna EPPO 2015 - Non Rischiare

Partecipanti e Aree – Ordine alfabetico

Organizzazioni Internazionali

Società Civile

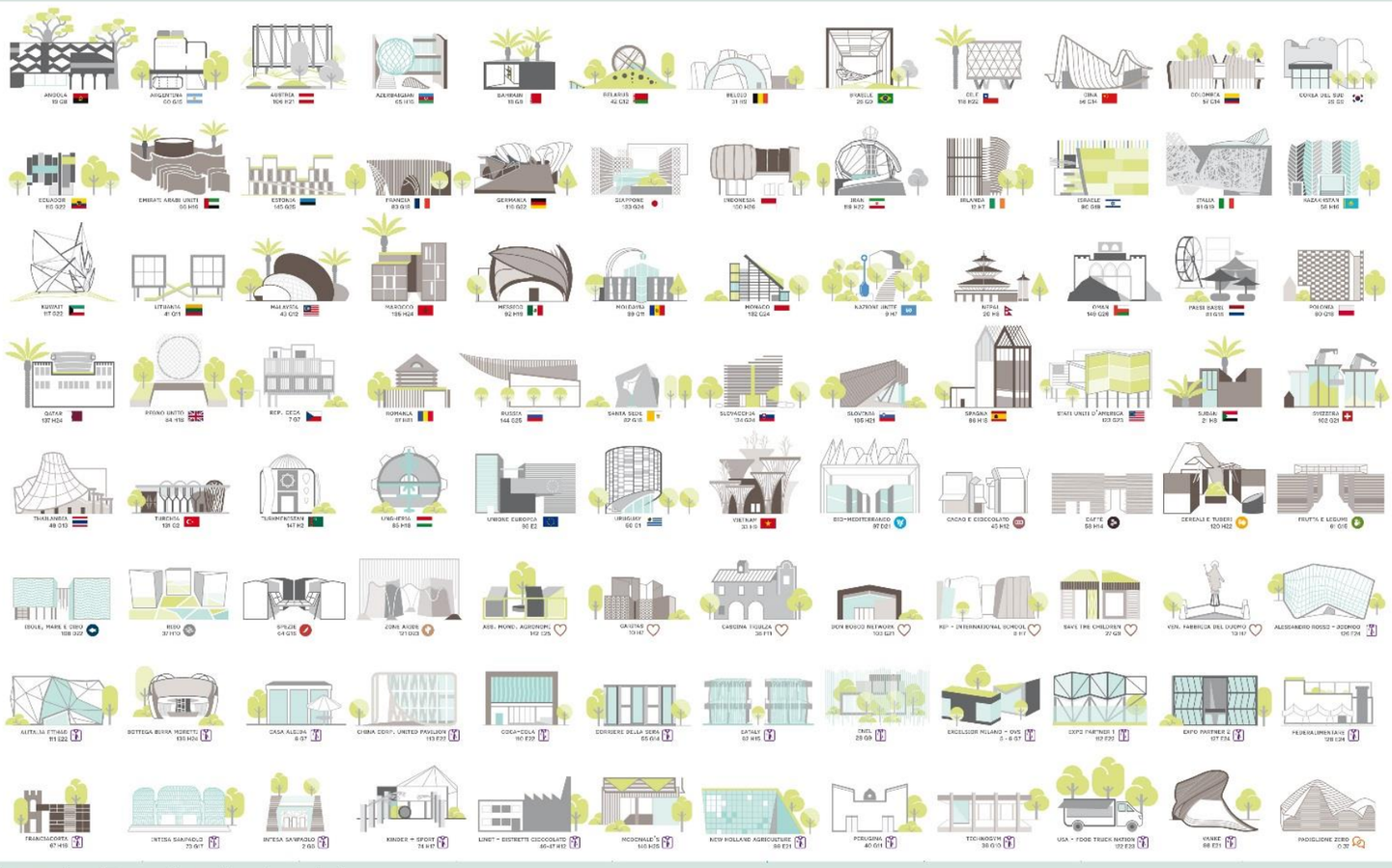
Aziende

Aree Tematiche

Aree Eventi

Prodotti Ufficiali

Un giro del mondo nell'architettura di Expo



Legenda - Trappole collocate nel sito

Simbologia	Tipo Trappola	Organismi monitorati in Expo	N.° trappole
	Multifunnel corti	*Anoplophora spp.*	10
		Pityophthorus juglandis	1
		Monochamus spp.	3
	Frismi viola	*Agrilus spp. (Agrilus planipennis; Agrilus anxius; Agrilus auroguttatus)*	10
	Secchielli	*Rhynchophorus spp.*	4
	Trappole verdi con all	*Popilia japonica*	10
	Pagoda Delta Trap	*Thaumatotibia leucotreta*	5
		Tecia solanivora	3
		Spodoptera littoralis	3
		Keiferia lycopersicella	3
		Tuta spp.	1
	Planotrap	*Planococcus ficus*	2
		Planococcus citri	1
	Rebell Amarillo	*Tetranichi (Bachrocera spp.; Ceratitis spp.)*	7
		Rhagoletis spp.	1
	Rebell Bianco	*Coleotteri (Byturus spp.); Tentredini (Hoplocampa spp.)*	8

Simbologia	Tipo Trappola	Organismi monitorati in Expo	N.° trappole
	Mela esca	*Rhythrophtora spp.*	6
	Capsapone	*Spora lungina*	1
	Cromotrappole Blu	*Thrips spp.*	10
		Bactrocera spp., Ceratitis spp. Rhagoletis spp., Strauzia longipennis; Toxoptera citricida; Oulema melanopus, Galerucella spp., Thrips spp.	20
	Cromotrappole Gialle	*Thrips spp.*	10
		Anthonomus eugenii	8
		Bactrocera spp.; Ceratitis spp.; Rhagoletis spp.; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Thrips spp; Diabrotica virgifera; Anastrepha spp.; Megacopta cribraria; Trioza erytreae; Bemisia tabaci.	25
	Piante spia	*Anoplophora spp.*	3
		Agrilus spp. (Agrilus planipennis, Agrilus anxius, Agrilus auroguttatus)	3
		Ernetinia aceriglovora	3
		Phytophthora ramorum	3
		Dunsapteleocrus e Gibberella	1
		Flavescenza e Xylella	1
		Sharka	1
		Citrus tristeza virus	2

Regione Lombardia
Servizio Fitosanitario

Aree di Servizio e ristorazione

Le Aree di Servizio

Sono dieci in tutto l'area espositiva, ci troverete una ricca offerta ristorativa. Per maggiori dettagli controllate le icone sulla mappa.

I Ristoranti

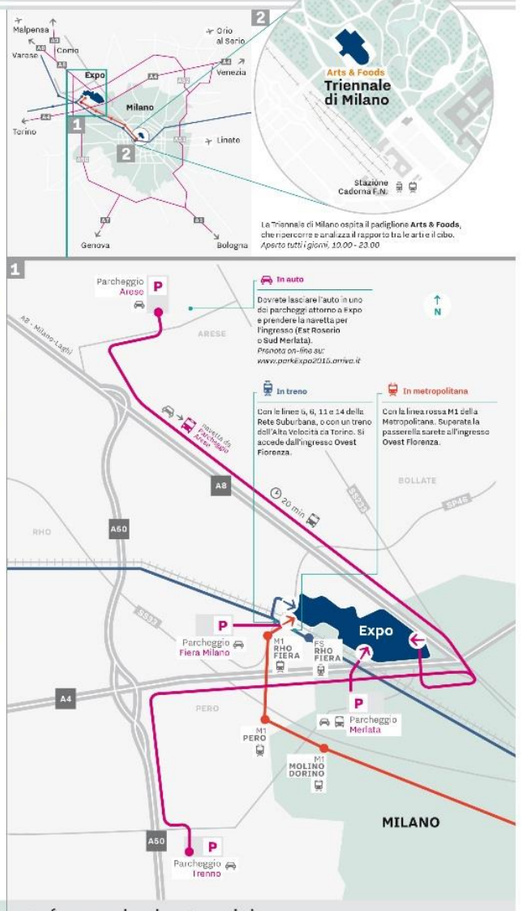
I Self-service

Bar e Caffettorie

I Chioschi

Cibo di Strada

Milano e l'Expo



Informazioni e Servizi

All'ingresso di Expo saranno svolti controlli di sicurezza secondo gli standard previsti negli aeroporti internazionali.

In Expo non ci sarà deposito bagagli. Spazi per deposito, limitati e a pagamento, saranno disponibili presso Fiera Milano a Rho.

Hai un biglietto a data aperta?
L'ingresso a Expo sarà consentito fino al raggiungimento della capacità massima giornaliera del sito espositivo. Scegli il giorno della tua visita su www.expo2015.org per assicurarti l'accesso nel giorno stesso.

I Servizi utili

75 I17
124 G23

Telefono Emergenza: 112
Informazioni: 020 2016

Nei punti informazioni sarà possibile acquistare i biglietti per gli spettacoli dell'Open Air Theatre San Carlo e prenotare le visite guidate.

© 2015 Expo 2015 SpA Aprile 2015
Direzione Visitor Experience and Exhibition Design
Edizione 1, maggio 2015
Per ristampe e diritti:
wayfinding@expo2015.org

Map & Wayfinding Design
Luigi Farrauto (100km Studio), Team Manager:
Andrea Noveli
Marco Palermo, Edoardo Nardella.
Valentina Marchioni (Illustrazioni)

ANNEX 2_CHECKPOINTS TABLE

Kind of traps	Monitored organisms	N°. Traps	Pheromons	pheromon/ attractive field life	n. pherom/ attractive for season	Monitoring period	Trap Code	Traps placement			Installation date	Pheromons replacement
								Coord. map	Zone	Plants		
Shorts Multifunnel	*Anoplophora spp.*	10	Kairomone 1	60 days	3 kit	may-sept	MfAn1	26H	green area with *Acer campestre*	*Acer campestre*	21-mag	Uninstalled
							MfAn2	28G	green perim. behind slow food	*Platanus* L.	21-mag	Uninstalled
			Kairomone 2	60 days			MfAn3	22D	Biodiversity Park	*Carpinus* sp.	27-mag	Uninstalled
							MfAn4	20B	green perim. behind life tree	*Platanus* L.	28-mag	Uninstalled
			Kairomone 3	60 days			MfAn5	16F	green perim. behind spices cluster	*Platanus* L.	28-mag	03-set / Uninstalled
							MfAn6	19J	green perim. behind future food district	*Platanus* L.	28-mag	03-set / Uninstalled
			Pheromone α	60 days			MfAn7	9J	green perim. Zero pavillion (corner)	*Platanus* L.	24-giu	Uninstalled
							MfAn8	8E	green perim. behind Angola pav.	*Platanus* L.	24-giu	Uninstalled
			Pheromone β	(2 pieces) 30 days			MfAn9	14I	green perim. behind restaurant area 96	*Platanus* L.	29-giu	Uninstalled
							MfAn10	23C	green perim. behind Arid areas cluster	*Acer platanoides*	29-giu	Uninstalled
	Pityophthorus juglandis	1	Pheromone	3 months-2 months (T>30°C)	1	may-oct	MfPi1	19I	Hortus 3 near Mexico pav.	*Juglans sp.*	28-mag	02-sept / Uninstalled
	Monochamus spp.	3	Pheromone gallowit K	6-8 weeks	3	middle may - end sept	MfMo1	23D	Biodiversity Park	*Pinus sp.*	27-mag	03-sept / Uninstalled
							MfMo2	14K	Roundabout into external perim. , behind Eataly	*Pinus sp.*	29-giu	08-set / Uninstalled
							MfMo3	E29	AMSA Area		29-lug	
Purple prism	*Agrilus spp. (Agrilus planipennis, Agrilus anxius, Agrilus auroguttatus)*	10	Pheromone leaf alcohol	3 months	2	middle apr/ early may-middle/end oct	PrAg1	H27	green perim. behind Biodiversity square/ Mediterranean hills	*Quercus sp.*	01-giu	02-sept / Uninstalled
							PrAg2	16F	green perim. behind Children Park (end)	*Fraxinus sp.*	01-giu	03-sept / Uninstalled
							PrAg3	25I	green perim. behind McDonald	*Quercus sp.*	25-giu	02-sept / Uninstalled
							PrAg4	18J	green perim. behind Auditorium	*Fraxinus sp.*	25-giu	02-sept / Uninstalled
							PrAg5	11I	green perim. behind Rice cluster	*Quercus sp.*	25-giu	02-sept / Uninstalled
			Pheromone manuka oil	2 months	3		PrAg6	6K	green perim. Behind Zero pav.	*Fraxinus sp.*	25-giu	02-sept / Uninstalled
							PrAg7	7E	green perim. behind Czech Rep. pav.	*Quercus sp.*	25-giu	02-sept / Uninstalled
							PrAg8	18D	green perim. behind Italy pav. (bridge 6 side)	*Quercus sp.*	25-giu	03-sept / Uninstalled
							PrAg9	22C	green perim. behind Island cluster	*Quercus sp.*	25-giu	03-sept / Uninstalled
							PrAg10	27C	green perim. Office side	*Fraxinus sp.*	25-giu	03-sept / Uninstalled
Buckets	*Rhyncophorus spp.*	4	Pheromone	10-15 weeks	3	apr-nov	SRh1	23I	Morocco pav.	*Phoenix spp.*	05-mag	Uninstalled
							SRh2	14G	Colombia pav.			Uninstalled
							SRh3	8G	Bahrein pav.			Uninstalled
							SRh4	26F	Oman pav.			Uninstalled
Green traps with wings	*Popilia japonica*	10	Pheromone	1 Full season	1	early spring - end summer	Tfly1	26D	NORD-EST-enclosure peak	enclosure	11-mag	Uninstalled
							Tfly2	26I	SUD-EST-enclosure peak	enclosure	11-mag	Uninstalled
							Tfly3	(20J) - 21H	green perim. behind Oper Air Theatre; 10/07 shifted in Austria pav.	*Fagus sp.*	10-giu	Uninstalled
							Tfly4	15K	green perim. behind Eataly	*Carpinus sp.*	10-giu	Uninstalled
							Tfly5	16F	green perim. behind spices cluster	*Fraxinus sp.*	10-giu	Uninstalled
							Tfly6	7J	green perim. behind Zero pav.	*Quercus sp.*	15-giu	Uninstalled
							Tfly7	12I	green perim. behind cocoa cluster	*Acer campestre*	15-giu	Uninstalled
							Tfly8	9E	green perim. behind Brazil pav.	*Platanus L.*	15-giu	Uninstalled
							Tfly9	6G	green perim. behind San Paolo	*Carpinus sp.*	24-giu	Uninstalled
							Tfly10	19B	green perim. behind life tree	*Carpinus sp.*	24-giu	Uninstalled
Pagoda Delta Trap	*Thaumatotibia leucotreta* (Koppert trap)	5	Pheromone	4-6 weeks	2	giu- aug o middle may-middle sept	PgTh1	26G	Hortus Oman side	*Citrus sp.*	15-giu	04-aug / Uninstalled
							PgTh2	23F	Hortus Turkey side	*Prunus sp.*	15-giu	04-aug / Uninstalled
							PgTh3	22H	Cereals and tubers Cluster	*Amaranthus sp.*	15-giu	04-aug / Uninstalled
							PgTh4	15G	Fruits and vegetables cluster	*Citrus sp.*	24-giu	Uninstalled
							PgTh5	11E	green area behind Cascina Triulza	*Punica granatum*	24-giu	04-aug / Uninstalled
	Tecia solanivora (Koppert trap)	3	Pheromone	30 days	3	giu-aug	PgTe1	22I	Cereals and tubers Cluster	*S. tuberosum*	15-giu	Uninstalled
							PgTe2	26H	Slow food pav.	*S. tuberosum*	24-giu	Uninstalled
							PgTe3	18H	Hungary pav.	*S. tuberosum*	13-lug	Uninstalled

Kind of traps	Monitored organisms	N°. Traps	Pheromons	pheromon/ attractive field life	n. pherom/ attractive for season	Monitoring period	Trap Code	Traps placement			Installation date	Pheromons replacement
								Coord. map	Zone	Plants		
Pagoda Delta Trap	*Spodoptera littoralis*	3	Pheromone	4-6 weeks	3	apr-sept	PgSp1	22D	Biodiversity Park	*Zea mais*	27-mag	28-lug / Uninstalled
							PgSp2	16G	Fruits and vegetables cluster	*S. lycopersicum*	27-mag	28-lug / Uninstalled
							PgSp3	15I	Hortus Eataly (Liguria side)	*Wisteria sp.*	27-mag	28-lug / Uninstalled
	Keiferia lycopersicella	3	Pheromone	4-6 weeks	2	middle giu - middle sept	PgKe1	15G	Fruits and vegetables cluster	*S. lycopersicum*	24-giu	17-aug / Uninstalled
							PgKe2	20G	Hortus Cardo side (Vino Italy)	*S. lycopersicum*	15-lug	17-aug / Uninstalled
							PgKe3	6H	Hortus structure services side, near Expo centre	*S. lycopersicum*	15-lug	26-aug / Uninstalled
	Sesia spp.	1	Pheromone		1		Sesia	26E	Biodiversity Park	*Malus sp.*	14-mag	04-aug / Uninstalled
Planotrap	*Planococcus fico*	1	Pheromone				PlPf1	22D	Biodiversity Park	*Ficus sp.*	26-mag	Uninstalled
	Planococcus fico	1	Pheromone				PlPf2	14D	Children park	*Ficus sp.*	27-mag	Uninstalled
	Planococcus citri	1	Pheromone				PlPc1	26F	Hortus front hill	*Citrus sp.*	21-mag	Uninstalled
Rebell Amarillo	*Tephritidae (Bactrocera spp.; Ceratitis spp.)*	7	/	/	/	may-oct	ReATf1	26E	Hortus behind "ass. mondiale agronomi"	*P. avium*	28-mag	Uninstalled
							ReATf2	23F	Hortus Turkey side	*P. avium*	28-mag	Uninstalled
							ReATf3	19H	Hortus near Mexico	*Juglans sp..*	28-mag	Uninstalled
							ReATf4	13D	Children park	*P. avium*	28-mag	Uninstalled
							ReATf5	27G	Mediterranean hills (top)	*Olea europaea*	13-lug	Uninstalled
							ReATf6	19G	Hortus behind Italy pav., Israel restaurant pav. side	*P. avium*	13-lug	Uninstalled
							ReATf7	7H	UN Garden	*P. avium*	13-lug	Uninstalled
	Rhagoletis spp.	1	Pheromone	4-6 weeks	3	end giu - end aug	ReARh1	17J	behind stand Kinder+sport	*P. avium*	13-lug	Uninstalled
Rebell White	*Coleoptera (Byturus spp.); Hymenoptera (Hoplocampa spp.)*	8	/	/	/	end giu - end aug	ReB1	26 G	pic nic area near Oman	*P. avium*	21-mag	Uninstalled
							ReB2	21 I	green area behind Austria pav.	*Rubus idaeus*	10-lug	Uninstalled
							ReB3	19 F	Hortus behind Italy pav., Israel restaurant pav. side	*Prunus sp.*	10-lug	Uninstalled
							ReB4	18I	Future food district, behind Spain pav.	*Malus sp.*	10-lug	Uninstalled
							ReB5	14I	green perim., behind coffee cluster	*Tilia sp.*	15-lug	Uninstalled
							ReB6	18J	Tub perimeter, Food future district side	*Malus sp.*	10-lug	Uninstalled
							ReB7	24I	flowerbed with *Morus* sp. and *Rubus* sp. shrub, behind Morocco pav.	*Rubus sp.*	20-lug	Uninstalled
							ReB8	10F	Flowerbed with *Populus* sp. and *Rubus* sp. shrub Korea pav. side/Cascina Triulza restaurant	*Rubus sp.*	20-lug	Uninstalled
Lures (Malus sp.)	*Phythophtora acerina et al.*	6	/	/	/	febb-dec	mela1	21B	green perim. behind bio-mediterranean cluster	*Malus sp.*	02-feb	Uninstalled
							mela2	22C	green perim. behind bio-mediterranean cluster		02-feb	Uninstalled
							mela3	26H	green perim. behind slow food		02-feb	Uninstalled
							mela4	16F	green perim. behind spice cluster, end of children park	*Malus sp.*	06-mag	Uninstalled
							mela5	7K	green perim. behind Zero pav.		06-mag	Uninstalled
							mela6	18J	Fitodepuration tub near Open air theatre		07-mag	Uninstalled
Captaspore	Fungal spores	1	/	/	/	febb-oct	Cap1	21 C	green perim., Arid areas cluster side.		02-feb	Uninstalled
Blue Chrome Traps	*Thrips spp.*	10	Kairomone	42 days	3	giu-sept	Cromo BTh1	19G	France pav.	*S. lycopersicum*	26-mag	03-aug/24-set Uninstalled
							Cromo BTh2	14D	Children park	*Citrus sp.*	26-mag	03-aug/24-set Uninstalled
							Cromo BTh3	23H	Cereals and tubers Cluster	*C. quinoa*	01-giu	03-aug/24-set Uninstalled
							Cromo BTh4	9G	Brazil pav.	*G. max/S. tuberosum*	01-giu	03-aug/24-set Uninstalled
							Cromo BTh5	21J	Parking on green perim. behind Open air theatre	flowerbed with *rosa* sp., *a. platanoides* near dumpster	15-lug	17-aug/28-set Uninstalled
							Cromo BTh6	16K	Parking on green perim. behind Eataly	flowerbed with *rosa* sp., *a. platanoides* near dumpster	15-lug	17-aug/28-set Uninstalled
							Cromo BTh7	8J	behind Zero pav.	Meadow with dumpsters	15-lug	disappeared
							Cromo BTh8	26I	Indonesia pav. side, behind services structure.	Meadow with dumpsters	15-lug	17-aug/24-set Uninstalled
							Cromo BTh9	14E	flowerbed area UN, behind Cina pav.	flowerbed with *Populus* sp. near dumpster	16-lug	17-aug/28-set Uninstalled
							Cromo BTh10	24C	Smoking pitch flowerbed behind Arid areas cluster	flowerbed with *P. granatum* near dumpsters	16-lug	24/09/2015 Uninstalled

Kind of traps	Monitored organisms	N°. Traps	Pheromons	pheromon/ attractive field life	n. pherom/ attractive for season	Monitoring period	Trap Code	Traps placement			Installation date	Pheromons replacement
								Coord. map	Zone	Plants		
Blue Chrome Traps	*Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; Agrilus auroguttatus; Thrips palmi.*	15	/	/	/	giu-sept	Cromo B1	20F	Hortus	*Malus sp.*	14-mag	Uninstalled
							Cromo B2	20E	Hortus	*P. persica*	14-mag	disappeared
							Cromo B3	15H	Hortus	*P. avium*	14-mag	disappeared
							Cromo B4	15I	Hortus	*Malus sp.*	14-mag	Uninstalled
							Cromo B5	27H	Mediterranean hills (behind slow food)	*Olea europaea*	19-mag	Uninstalled
							Cromo B6	27H	Mediterranean hills (behind slow food)	*Olea europaea*	19-mag	Uninstalled
							Cromo B7	27G	Mediterranean hills	*Olea europaea*	19-mag	Uninstalled
							Cromo B8	6J	Hortus	*Malus sp.*	19-mag	Uninstalled
							Cromo B9	8I	Hortus	*Pyrus sp.*	19-mag	disappeared
							Cromo B10	19G	Hortus	*P. persica*	19-mag	Uninstalled
							Cromo B11	20H	Hortus	*Prunus sp.*	19-mag	Uninstalled
							Cromo B12	23D	Biodiversity Park	*Citrus sp.*	26-mag	Uninstalled
							Cromo B13	15F	Fruits and vegetables cluster	*Malus sp.*	26-mag	Uninstalled
							Cromo B14	6G	green area	*Malus sp.*	26-mag	Uninstalled
							Cromo B15	6H	green area	*Prunus armeniaca*	26-mag	Uninstalled
							Cromo B16	G27	Mediterranean hills (top)	*Olea europaea*	19-mag	Uninstalled
							Cromo B1amsa	E29	amsa area	railing	29-lug	
							Cromo B2amsa	E29	amsa area	railing	29-lug	
Yellow Chrome Traps	*Thrips spp.*	10	Kairomone	42 days	3	giu-sept	Cromo GTh1	26H	Slow food pav.	*S. lycopersicum*	26-mag	03-aug/24-set Uninstalled
							Cromo GTh2	15G	Fruits and vegetables cluster	*S. lycopersicum*	26-mag	03-aug/24-set Uninstalled
							Cromo GTh3	23G	Hortus Turkey side	*P. persica*	28-mag	03-aug/24-set Uninstalled
							Cromo GTh4	20H	Hortus near "Martini"	*Vitis sp.*	28-mag	03-aug/24-set Uninstalled
							Cromo GTh5	23H	Cereals and tubers Cluster	*S. tuberosum*	01-giu	03-aug/24-set Uninstalled
							Cromo GTh6	9F	Brazil pav.	*Abelmoschus esculentus*	01-giu	03-aug/24-set Uninstalled
							Cromo GTh7	24J	Parking near green perim. behind Quatar pav.	flowerbed with *Rosa* sp., *A. platanoides* near dumpster	15-lug	17-aug/24-set Uninstalled
							Cromo GTh8	10F	green perim. behind Korea pav.	*Populus* sp. near dumpster	15-lug	17-aug/28-set Uninstalled
							Cromo GTh9	14F	Behind kitchen Argentine pav.	shrub with *Pittosporum* sp., palm trees	15-lug	17-aug/28-set Uninstalled
							Cromo GTh10	19G	Hortus near Israel pav.	*S. lycopersicum*	16-lug	17-aug Uninstalled
	Anthonomus eugenii	8	Pheromone	4 weeks (2 pieces)	4	giu-sept	Cromo GAt1	18H	Hungary pav.	*Capsicum L.*	29-giu	3-aug Uninstalled
							Cromo GAt2	16G	Spices Cluster	*Capsicum L.*	29-giu	3-aug Uninstalled
							Cromo GAt3	8H	Nepal pav.	*Capsicum L.*	29-giu	3-aug Uninstalled
							Cromo GAt4	26H	Slow food pav.	*Capsicum L.*	29-giu	4-aug Uninstalled
							Cromo GAt5	9G	Brazil pav.	*Capsicum L.*	14-lug	18-aug Uninstalled
							Cromo GAt6	7H	UN Garden	*Capsicum L.*	14-lug	18-aug Uninstalled
							Cromo GAt7	11F	Cascina Triulza garden	*Capsicum L.*	16-lug	18-aug Uninstalled
							Cromo GAt8	19G	France pav.	*Capsicum L.*	16-lug	18-aug Uninstalled
	Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; A. auroguttatus; Thrips palmi; Diabrotica virgifera; Anthonomus eugenii; Anastrepha spp.; Megacopta cribraria; Trioza eritreae; Bemisia tabaci.	20	/	/	/	may-oct	Cromo G1	26E	Biodiversity Park	*Vitis sp.*	11-mag	Uninstalled
							Cromo G2	26E	Hortus near "ass. mondiale agronomi"	*Citrus sp.*	11-mag	Uninstalled
							Cromo G3	26F	Hortus near Oman	*Prunus sp.*	11-mag	Uninstalled
							Cromo G4	26G	Hortus near Oman	*Wisteria sp.*	11-mag	Uninstalled
							Cromo G5	26G	Hortus near Oman	*Citrus sp. vaso*	11-mag	disappeared
							Cromo G6	23E	Hortus near Partner Expo 2	*Prunus sp.*	11-mag	Uninstalled
							Cromo G7	14F	Arbor behind Colombia pav.	*Vitis sp.*	14-mag	Uninstalled
							Cromo G8	20I	Hortus	*Pyrus sp.*	14-mag	Uninstalled
							Cromo G9	20G	Hortus	*P. persica*	14-mag	Uninstalled
							Cromo G10	12F	behind cascina Triulza	*Diospyros kaki*	14-mag	Uninstalled
							Cromo G11	17J	behind stand Kinder+sport	*Prunus sp.*	13-lug	Uninstalled

Kind of traps	Monitored organisms	N°. Traps	Pheromons	pheromon/ attractive field life	n. pherom/ attractive for season	Monitoring period	Trap Code	Traps placement			Installation date	Pheromons replacement
								Coord. map	Zone	Plants		
Yellow Chrome Traps	*Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; Agrilus auroguttatus; Thrips palmi; Diabrotica virgifera; Anthonomus eugenii; Anastrepha spp.; Megacopta cribraria; Trioza eritreae; Bemisia tabaci.*		/	/	/	may-oct	Cromo G12	27H	Mediterranean hills (behind slow food)	*Olea europaea*	19-mag	Uninstalled
							Cromo G13	27H	Mediterranean hills (behind slow food)	*Olea europaea*	19-mag	Uninstalled
							Cromo G14	27G	Mediterranean hills	*Olea europaea*	19-mag	Uninstalled
							Cromo G15	6I	Hortus	*Punica granatum*	19-mag	Uninstalled
							Cromo G16	19F	Hortus	*Prunus sp.*	19-mag	Uninstalled
							Cromo G17	19H	Hortus	*Citrus sp.*	19-mag	Uninstalled
							Cromo G18	9G	Save the Children pav.	*Malus sp.*	13-lug	disappeared
							Cromo G19	7I	green area behind "Fabbrica del Duomo"	*Citrus sp.*	14-lug	Uninstalled
							Cromo G20	18I	behind Hungary pav.	*P. avium*	13-lug	disappeared
							Cromo G21	21D	Bio-Mediterranean Cluster	*Olea europaea*	05-ago	Uninstalled
							Cromo G22	22D	Bio-Mediterranean Cluster	*Olea europaea*	05-ago	Uninstalled
							Cromo G23	21C	Bio-Mediterranean Cluster	*Olea europaea*	05-ago	disappeared
							Cromo G1amsa	E29	amsa area	railing	29-lug	
							Cromo G2amsa	E29	amsa area	railing	29-lug	
Spy plants	*Anoplophora spp.*	3	/	/	/	may-oct	PsAn1	27H	green perim. behind biodiversity square/Mediterranean hills	*Alnus* sp.	26-ago	
							PsAn2	16J	green perim. behind Conference centre	*Acer campestre*	26-ago	
							PsAn3	7F	green perim. behind "Casa Algida"	*Acer campestre*	26-ago	
	Agrilus spp. (Agrilus planipennis, A. anxius, A. auroguttatus)	3	/	/	/	may-oct	PsAg1	21F	behind Don Bosco pav.	*Betula* sp.	28-lug	
							PsAg2	20J	green perim. behind Open air theatre	*Quercus sp.*	26-ago	
							PsAg3	13H	green perim. behind "Distretto cioccolato"	*Fraxinus sp.*	26-ago	
	Erwinina amylovora	3	/	/	/	middle may-middle lug / middle set-middle oct	PsEr1	20B	green perim. behind albero della vita	*Crataegus* sp.(25 plants)	26-ago	
							PsEr2	27C	green perim. bus stop 6 side	*Crataegus* sp.	04-set	
							PsEr3	K8	green perim. behind Zero pav.	*Crataegus* sp.	04-set	
	Phytophthora ramorum	3	/	/	/		PsPh1	6F	green perim. railing side , near bus stop n.2	*Viburnum* sp.	03-set	
							PsPh2	23C	green perim. behind cluster arid areas	*Viburnum* sp.	03-set	
							PsPh3	23K	green perim. railing side, behind Morocco pav./structure service	*Viburnum* sp.	03-set	
	Bursaphelenchus and *Gibberella*	1	/	/	/		PsBG1	9K	Roundabout Zero pav. side	*Pinus mugo*	03-set	
	Flavescenza and Xylella	1	/	/	/		PsFl1	14F	Arbor behind Argentine	*Vitis sp.*	04-set	
	Sharka	1	/	/	/		PsSh1	12E	Cascina triulza	*Prunus domestica*	04-set	
	Citrus tristezza virus	2	/	/	/	all the year	PsCi1	19H	Hortus Mexico	*Citrus limon*	22-giu	
							PsCi2	15E	Children park, end part	*Citrus limon*	27-ago	

ANNEX 3_ORGANISMS OF GREEN PERIMETER

ORGANISMS OF GREEN PERIMETER					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Expo green perimeter (Apples)	Search *Phytophthora* sp.	Apples (Mela1)		16/02/2015	Presence of *Pythium litorale*
				02/03/2015	Presence of *Mucor* sp. and *Galactomyces geotrichum* sp.
				16/03/2015	Presence of *Pythium dissotocum*
				07/04/2015	Presence of *Clonostachys rosea*
				20/04/2015	Presence of *Trichoderma asperellum*
				27/04/2015	Presence of *Mucor* sp.
				30/07/2015	Presence of *Colletotrichum fiorinae*
				28/08/2015	Presence of *Colletotrichum acutatum*
				05/10/2015	Presence of *Pythium dissotocum*
				22/10/2015	Presence of *Colletotrichum acutatum*
		Apples (Mela2)		23/02/2015	Presence of *Alternaria alternata*
				16/03/2015	Presence of *Pythium dissotocum*
				13/04/2015	Presence of *Mucor* sp.
				27/04/2015	Presence of *Mucor* sp.
				07/04/2015	Presence of *Candida* sp.
				04/05/2015	Presence of *Fusarium* sp
				11/05/2015	Presence of *Fusarium tricinctum*
				18/05/2015	Presence of *Galactomyces* sp.
				28/08/2015	Presence of *Colletotrichum acutatum*
				17/09/2015	Presence of *Pythium dissotocum*
		Apples (Mela3)		02/03/2015	Presence of *Mucor* sp. and *Galactomyces geotrichum* sp.
				16/03/2015	Presence of *Pythium dissotocum*
				07/04/2015	Presence of *Galactomyces geotrichum*
				13/04/2015	Presence of *Mucor* sp.
				27/04/2015	Presence of *Mucor* sp.
				04/05/2015	Presence of *Mucor* sp.
				11/05/2015	Presence of *Phytophthora lacustris*
				22/06/2015	Presence of *Colletotrichum acutatum*
				28/08/2015	Presence of *Colletotrichum fiorinae*
				17/09/2015	Presence of *Colletotrichum acutatum*
				05/10/2015	Presence of *Pythium dissotocum*
				22/10/2015	Presence of *Colletotrichum acutatum*
		Apples (Mela4)		11/05/2015	Presence of *Pythium dissotocum*
				22/06/2015	Presence of *Colletotrichum acutatum*
				30/07/2015	Presence of *Colletotrichum acutatum*
				22/10/2015	Presence of *Colletotrichum acutatum*
		Apples (Mela5)		11/05/2015	Presence of *Trichoderma koningopsis*
				22/06/2015	Presence of *Colletotrichum fiorinae*
				30/07/2015	Presence of *Colletotrichum fiorinae*
				28/08/2015	Presence of *Colletotrichum fiorinae*
				17/09/2015	Presence of *Pythium dissotocum*
				05/10/2015	Presence of *Pythium dissotocum*
				22/10/2015	Presence of *Geotrichum candidum*
Expo green perimeter (Captaspore) (21C)	Search fungal spores	Spores		09/02/2015	Presence of conidia of *Alternaria* sp. and *Diplodia* sp.
				10/02/2015	Presence of conidia of *Alternaria* sp. and *Diplodia* sp.
				02/03/2015	Presence of *Leptosphaeria* sp. and *Cladosporium* sp.
				09/03/2015	Presence of *Cladosporium* sp, *Alternaria* sp., *Epicoccum* sp.
				16/03/2015	Presence of *Alternaria* sp.
				23/03/2015	Presence of *Sphaeropsis* sp. spores
				30/03/2015	Presence of *Alternaria* sp.
				07/04/2015	Presence of *Cladosporium* sp. and *Alternaria* sp.
				13/04/2015	Presence of *Diplodia* sp., *Cladosporium* sp. and *Alternaria* sp. spores.
				20/04/2015	Presence of *Alternaria* sp., *Tilletia* sp., *Cladosporium* sp. spores.
				27/04/2015	Presence of *Alternaria* sp. and *Tilletia* sp. spores.
				11/05/2015	Presence of *Alternaria alternata* , *Diplodia* sp. and *Cladosporium* sp.
				18/05/2015	Presence of *Alternaria* sp. , *Epicoccum* sp., *Galactomyces* sp.
				22/06/2015	Presence of *Alternaria* sp.,*Epicoccum* sp., *Cladosporium* sp., *Sphaeropsis* sp.
				29/06/2015	Presence of *Alternata alternata* , *Epicoccum* sp., *Cladosporium* sp. and *Plasmopara* sp. oospores
				07/07/2015	Presence of *Alternaria alternata* , *Epicoccum* sp., *Cladosporium* sp. spores and *Plasmopara* oospores.
				14/07/2015	Presence of *Alternaria alternata* , *Epicoccum* sp., *Cladosporium* sp. spores

ORGANISMS OF GREEN PERIMETER					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Expo green perimeter (Captaspore) (21C)	Search fungal spores	Spores		20/07/2015	Presence of *Trichothecium roseum* , *Alternaria alternata* , *Cladosporium* sp., *Epicoccum* sp., and *Plasmopara* sp. oospores.
				27/07/2015	Presence of *Cladosporium* sp., *Plasmopara* sp., *Alternata alternata* , *Epicoccum* sp. spores.
				03/08/2015	Presence of *Epicoccum* sp., *Trichotecium roseum* , *Alternaria alternata* spores.
				10/08/2015	Presence of *Alternata alternata* , *Cercospora* sp., *Plasmopara* sp., *Cladosporium* sp., *Epicoccum* sp. spores.
				17/08/2015	Presence of *Alternaria alternata* , *Cladosporium* sp., *Epicoccum* sp. spores.
				25/08/2015	Presence of *Alternaria* sp. spores.
				31/08/2015	Presence of *Alternaria* sp., *Epicoccum* sp., *Plasmopara* sp., *Cladosporium* sp. spores.
				07/09/2015	Presence of *Alternaria* sp. , *Epicoccum* sp..
				14/09/2015	Presence of *Alternaria* sp., *Epicoccum* sp., *Drechslera* sp., *Cladosporium* sp. spores.
				21/09/2015	Presence of *Alternaria* sp., *Cladosporium* sp., *Drechslera* sp., *Epicoccum* sp., *Pestalotiopsis* sp. spores
				28/09/2015	Presence of *Alternaria* sp., *Epicoccum* sp., *Drechslera* sp., *Diplodia* sp., *Cladosporium* sp. spores.
				05/10/2015	Presence of *Epicoccum* sp. spores.
				13/10/2015	Presence of *Cladosporium* sp., *Epicoccum* sp., *Alternaria* sp. spores.
				19/10/2015	Presence of *Epicoccum* sp. and *Alternaria* sp. spores.
				26/10/2015	Presence of *Epicoccum* sp., and *Alternaria* sp. spores.
				30/10/2015	Presence of *Epicoccum* sp. and *Alternaria* sp. spores.
Expo green perim. (Tfly6)(7J)	Search for *Popillia japonica*	Entomological samples		23/07/2015	Presence of an exemplar on *Lucilia sericata* and *Rhaphigaster nebulosa*
Expo green perim.(Tfly5) (16F)	Search for *Popillia japonica*	Entomological samples		23/07/2015	Presence of an exemplar on *Cetonia aurata* , *Corythucha ciliata* , and two exemplar on *Halyomorpha halys*
Expo green perimeter (Tfly7) (12I)	Search for *Popillia japonica*	Entomological samples		09/07/2015	Presence of *Diabrotica v. virgifera*
		Entomological samples		05/08/2015	Presence of an exemplar on *Halyomorpha halys* , *Paracorsia repandalis* , and *Adalia decempunctata*
Expo green perim. (Tfly8)(9E)	Search for *Popillia japonica*	Entomological samples		09/07/2015	Presence of *Formica* sp.
Expo green perim.(Tfly9) (6G)	Search for *Popillia japonica*	Entomological samples		05/08/2015	Presence of an exemplar on *Harpalus rufipes*
Expo green perim. (Tfly10)(19B)	Search for *Popillia japonica*	Entomological samples		09/07/2015	Presence of *Rhaphigaster* sp.
		Entomological samples		26/08/2015	Presence of an exemplar on *Cetonia aurata*
Expo green perim. (ReB5) (14I)	Search Tentredini	Entomological samples		31/08/2015	Presence of *Mystacides azureus*
Expo green perim. (PrAg1) (27H)	Search for *Agrilus* spp.	Entomological samples		02/09/2015	Presence of *Deraeocoris lutescens* and *Scymnus auritus*
Expo green perim. (PrAg3) (25I)	Search for *Agrilus* spp.	Entomological samples		11/08/2015	Presence of an exemplar on *Bruchus brachialis* , and an exemplar on *Metcalfa pruinosa*
Expo green perim. (PrAg5) (11I)	Search for *Agrilus* spp.	Entomological samples		02/09/2015	Presence of *Hydroptila angulata* , *Hylaeus* sp. (*dilatatus* or *angularis*), *Deraeocoris* sp., *Adalia bipunctata* and *Ophraella communa*
Expo green perim. (PrAg7) (7E)	Search for *Agrilus* spp.	Entomological samples		02/09/2015	Presence of *Orientus ishidae*
Expo green perim. (PrAg8) (18D)	Search for *Agrilus* spp.	Entomological samples		03/09/2015	Presence of *Stethorus punctillum* and *Harmonia axyridis*
Expo green perim. (PrAg10) (27C)	Search for *Agrilus* spp.	Entomological samples		03/09/2015	Presence of *Ophraella communa* and *Orietus ishidae*
Expo green perim. (MfAn2) (28G)	Search for *Anoplophora* spp.	Entomological samples		29/09/2015	Presence of *Halyomorpha halis* , *Vespa crabro* and *Chlorophorus varius varius*
Expo green perim. - Parking (CromoBTh5) (21J)	Search for *Thrips*	Entomological samples		07/09/2015	Presence of *Nomophila noctuella* , *Thrips tabaci* , *Athalia rosae rosaee* and *Frankliniella occidentalis*
				28/09/2015	Presence of hundreds of *Thrips tabaci* specimens
				27/11/2015	Presence of *Frankliniella occidentalis*
Expo green perim. - Parking (CromoBTh6) (16K)	Search for *Thrips*	Entomological samples		07/09/2015	Presence of thrips *Frankliniella intonsa* species. Presence of *Megalonotus sabulicola* , *Choreutis nemorana* , *Helicoverpa armigera,Ophraella communa* , *Fieberiella florii* and *Japananus hyalinus.*
				28/09/2015	Presence of one hundred of thrips *Aeolothrips intermedius* , *Fieberiella florii* and *Japananus hyalinus*
				02/11/2015	Presence of *Tenothrips* sp. (prob. *frici*), *Japananus hyalinus* , *Anoecia* sp.

ORGANISMS OF GREEN PERIMETER					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Expo green perim. - Parking (CromoGTh7) (24J)	Search for *Thrips*	Entomological samples		27/11/2015	Presence of *Frankliniella occidentalis* and *Fieberiella florii*
Expo green perim. - row of trees behind Korea (CromoGTh8) (10F)	Search for *Thrips*	Entomological samples		08/09/2015	Presence of *Frankliniella intonsa* , *Nomophila noctuella* , *Ophraella communa* , *Cicadella viridis* , *Hylaeus* sp., and *Agraylea sexmaculata*
				28/09/2015	Presence of dozens of thrips *Frankliniella occidentalis* . Presence of *Dictya* sp. specimens.
				29/10/2015	Presence of *Frankliniella occidentalis* , *Psammotettix confinis* , *Hydroptila vectis* , *Adalia bipunctata*
Expo green perim.- roundabout behind coffee cluster (14K)	*Xylella fastidiosa*	*Spartium* sp.	2	22/09/2015	Absent
Mediterranean hill	*Xylella fastidiosa*	*Olea europaea*	81	06/05/2015	Absent
		Rosmarinus sp.	2	22/09/2015	Absent
		Spartium sp.	2	22/09/2015	Absent
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoB5)		27/05/2015	*Thrips tabaci*
				29/10/2015	Presence of *Frankliniella intonsa* , *Thaumatomyia notata* , Psychodidae, and *Ischnopterapion virens*
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoB16)		27/05/2015	*Thrips tabaci*
				22/07/2015	Presence of *Frankliniella occidentalis*
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoB16)		18/08/2015	Presence of an exemplar on *Autographa gamma* , an exemplar on *Laodelphax striatellus* , *Spodoptera exigua* , and *Deraeocoris nebulosus*
				17/09/2015	Presence of *Dictya* sp.
				29/10/2015	Presence of *Thrips tabaci* and *Ophraella communa*
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoG12)		22/07/2015	Presence of *Haplothrips aculeatus*
				18/08/2015	Presence of an exemplar on *Harmonia axyridis* , two exemplars on *Ophraella communa* , and an exemplar on *Laodelphax striatellus*
				17/09/2015	Presence of *Daraeocoris nebulosus* , *Hydroptila vectis* , *Cicadella viridis* , *Ophraella communa* , *Coenosia attenuata* , *Thaumatomyia notata* , and *Laodelphax striatellus*
				26/10/2015	Presence of *Acrolepiopsis assectella* , *Zygina rahmni* , *Ophraella communa* and *Stethorus* sp.
				29/10/2015	Presence of *Frankliniella occidentalis* , *Cicadella viridis* , *Dictya* sp. and psychodidae specimens
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoG13)		22/07/2015	Presence of *Frankliniella intonsa*
				18/08/2015	Presence of two exemplars on *Ophraella communa* and *Galerucella luteola*
				17/09/2015	Presence of *Aphelinus* sp., *Dictya* sp., *Coenosia attenuata* , and *Thaumatomyia notata*
				29/10/2015	Presence of *Frankliniella occidentalis*
Mediterranean hill	Search Fruit flies and *Thrips*	*Olea europaea* (CromoG14)		22/07/2015	Presence of *Thrips tabaci*
				18/08/2015	Presence of an exemplar on *Ophraella communa* , some *Aphis craccivora* specimens
				17/09/2015	Presence of *Spodoptera exigua* , *Stegobium paniceum* , *Frankliniella occidentalis, Thrips hawaiiensise* , and *Dictya* sp.
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoB6)		27/05/2015	Presence of *Thrips tabaci* and *Anaphothrips obscurus*
				18/08/2015	Presence of an exemplar on *Spodoptera exigua* , *Laodelphax striatellus* , and some *Coenosia attenuata* specimens
				17/09/2015	Presence of *Cricotopus* sp., *Haplothrips* sp., and *Dictya* sp.
				29/10/2015	Presence of *Frankliniella intonsa*
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoB7)		27/05/2015	Presence of *Thrips trehernei*
				22/07/2015	Presence of *Thrips tabaci*
				18/08/2015	Presence of an exemplar on *Aelia acuminata* , and *Laodelphax striatellus*
				17/09/2015	Presence of *Laodelphax striatellus* and *Nysius* sp.
				29/10/2015	Presence of *Frankliniella occidentalis*
	Search diptera tephritidae	*Olea europaea* (ReATf5)		09/09/2015	Presence of *Altica oleracea* , *Ophraella communa* , and *Cicadella viridis*
				23/10/2015	Presence of *Ophraella communa* , *Coccinella decempunctata* , *Dictyophora europaea* , *Altica oleracea* , *Euleia* sp., *Trixagus meybohmi*
Hortus (6G)	Search Fruit flies and *Thrips*	*Malus* sp. (CromoB14)		22/06/2015	Presence about ten of Thrips not determinable
				23/07/2015	Presence of *Haplothrips aculeatus*
				30/10/2015	Presence of *Hydroptila vectis* , some Psychodidae specimens and *Anoecia* sp.

ORGANISMS OF GREEN PERIMETER					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Hortus (6H-J)	Search Fruit flies and *Thrips*	*Malus* sp. (CromoB8)		22/06/2015	Presence of *Thrips tabaci*
				23/07/2015	Presence of *Frankliniella occidentalis* , *Tenothrips frici* and *Aeolothrips intermeofus*
				27/08/2015	Presence of *Hydroptila vectis*
				30/10/2015	Presence of Psychodidae specimens
	Search Fruit flies and *Thrips*	*Prunus armeniaca* (CromoB15)		22/06/2015	Presence about ten of Thrips not determinable
				23/07/2015	Presence of *Frankliniella occidentalis* and *Aeolothrips intermedius*
				27/08/2015	Presence of *Ophraella communa*
				30/10/2015	Presence of caddis-fly and *Anoecia* sp.
	Search Fruit flies and *Thrips*	*P. granatum* (CromoG15)		23/07/2015	Presence of *Frankliniella occidentalis*
				27/08/2015	Presence of *Euleia* sp. and *Acanthiophilus helianthi*
				30/10/2015	Presence of *Zygina lunaris* , *Siphoninus phyllireae, Hydroptila vectis* and whiteflies
	Search for *Keiferia licopersicella*	*S. lycopersicum* (PgKe3)		29/09/2015	Presence of *Ophraella communa* and *Propylaea* 14-*punctata*
Hortus (15H-J) beside Eataly	Search aphids	*Wisteria* sp.		24/06/2015	Presence of *Aphis craccivora*
	Identification moth	*Wisteria* sp.		24/06/2015	Presence of *Autographa gamma*
	Search Fruit flies and *Thrips*	*P. avium* (CromoB3)		22/07/2015	Presence of *Thrips tabaci*
	Search Fruit flies and *Thrips*	*Malus sp.* (CromoB4)		22/07/2015	Presence of *Thrips tabaci*
				02/11/2015	Presence of *Coccotrypes dactyliperda* , *Hydroptila vectis* , *Dictya* sp., *Stephanitis pyri*
Hortus (19 F-G) beside Israel	Search *Thrips*	Entomological samples (CromoGTh10)		17/08/2015	Presence of *Frankliniella occidentalis*
				07/09/2015	Presence of dozens of *Tenothrips frici* . Presence of *Aphis craccivora.*
				24/09/2015	Presence about ten thrips of *Frankliniella occidentalis* and *Haplothrips aculeatus* species
				29/10/2015	Presence of *Frankliniella occidentalis* , *Cicadella viridis* and psychodidae specimens
	Search Fruit flies and *Thrips*	*P. persica* (CromoB10)		25/08/2015	Presence of *Coenosia attenuata* , *Sepedon sphegea* and *Spodoptera exigua*
				29/09/2015	Presence of *Lygus* sp. and psychodidae specimens
				29/10/2015	Presence of *Thrips tabaci* and *Ophraella communa*
	Search Fruit flies and *Thrips*	*Prunus sp.* (CromoG16)		29/09/2015	Presence of *Zygina rhamni* , *Dictya* sp., *Scymnus* sp. and *Stethorus* sp.
				29/10/2015	Presence of *Rhopalosiphum padi* , *Euleia* sp. and psychodidae specimens
	Search diptera tephritidae	*P. avium* (ReATf6)		29/09/2015	Presence of *Ophraella communa* , *Athalia rosae rosae* , *Scymnus* sp. and *Stethorus* sp.
				23/10/2015	Presence of *Hishimonus hamatus* , *Typhlocyba* sp., Psychodidae specimens, *Macrosteles* sp., Ephemeroptera specimens, and *Scymnus* sp.
Hortus (19H-I) beside Mexico	*Xylella fastidiosa*	*Olea europaea*	13	06/05/2015	Absent
	Search Fruit flies and *Thrips*	*Citrus* sp. (CromoG17)		29/09/2015	Presence of *Laodelphax striatellus* , *Dyctia* sp. and Psychodidae specimens
				30/10/2015	Presence of *Hydroptila angulata* , Psychodidae and whiteflies specimens
	Search Tephritidae	*Juglans* sp. (ReATf3)		23/10/2015	Presence of *Chaetocnema tibialis* , *Zyginidia pullula* , *Scymnus* sp.and Tingidae specimens
Hortus (20E-G) beside Cardo	Search Fruit flies and *Thrips*	*Malus* sp. (CromoB1)		20/07/2015	Presence of *Frankliniella occidentalis*
				30/10/2015	Presence of *Drosophila suzukii* and psychodidae specimens
		P. persica (CromoB2)		29/09/2015	Presence of *Scymnus* sp. and *Stethorus* sp.
		P. persica (CromoG9)		25/08/2015	Presence of *Lucilia sericata*
				29/09/2015	Presence of *Scymnus* sp., of *Stethorus* sp. and *Japananus hyalinus*
				30/10/2015	Presence of *Euleia* sp.
	Search for *Keiferia licopersicella*	*S. lycopersicum* (PgKe2)		17/08/2015	Presence of *Ophraella communa*
				29/09/2015	Presence of Psychodidae specimens
Hortus (20H-I)	Search Fruit flies and *Thrips*	*Prunus* sp. (CromoB11)		27/05/2015	Presence of *Thrips hawaiiensis* and *Thrips major*
				25/08/2015	Presence of *Coenosia attenuata* and *Ophraella communa*
				29/09/2015	Presence of *Cicadella viridis*
				30/10/2015	Presence of *Drosophila suzukii, Drosophila* sp. (prob. *simulans*)
		Pyrus sp. (CromoG8)		25/08/2015	Presence of *Ophraella communa*
				29/09/2015	Presence of *Cicadella viridis* , *Thaumatomya notata* , *Scymnus* sp. and *Stethorus* sp.
				30/10/2015	Presence of *Zyginidia pullula* , *Zygina rhamni* and psychodidae specimens

ORGANISMS OF GREEN PERIMETER					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Hortus (20H-I)	Search *Thrips*	*Vitis* sp. (CromoGTh4)		03/08/2015	Presence of dozens di thrips of *Frankliniella occidentalis.*
				07/09/2015	Presence of dozens of thrips of *Frankliniella occidentalis* and *Thrips tabaci*
				30/10/2015	Presence of *Thaumatomyia notata* , *Zygina* sp., *Zyginidia pullula, Ophraella communa*
Hortus (23E-G) beside Turkey	Search Fruit flies and *Thrips*	*Prunus* sp. (CromoG6)		22/07/2015	Presence of *Frankliniella intonsa*
				25/08/2015	Presence of *Lygus pratensis* , *Harmonia axyridis* and *Ophraella communa.*
				29/10/2015	Presence of *Rhopalosiphum padi, Zygina rhamni*
	Search Thrips	*P. persica* (CromoGTh3)		22/07/2015	Absent of Thrips specimens, Presence of *Diabrotica virgifera virgifera*
				03/08/2015	Presence of less then 10 thrips of *Frankliniella intonsa* .
				07/09/2015	Presence of thrips of *Haplothrips aculeatus* and *Ophraella communa*
				29/10/2015	Presence of *Frankliniella occidentalis*
	Search diptera tephritidae	*P. avium* (ReATf2)		09/09/2015	Presence of *Euleia* sp., *Scymnus* sp. and *Stethorus* sp.
				23/10/2015	Presence of *Typhlocyba* sp., *Hydroptila angulans* and *Scymnus sp.*
	Search for *Thaumatotibia leucotreta*	*Prunus* sp. (PgTh2)		17/09/2015	Presence of *Pammene albuginana*
	Xylella fastidiosa	*P. avium*	2	22/09/2015	Absent
Hortus (26E-G)	Search Fruit flies and *Thrips*	*Citrus* sp. (CromoG2)		20/07/2015	Presence of *Thrips tabaci* and *Aeolothrips intermedius*
				18/08/2015	Presence of *Zyginidia pullula* and an exemplar on *Laodelphax striatellus*
				15/09/2015	Presence of *Dictya* sp.
				29/10/2015	Presence of *Cicadella viridis,* Lycaenidae specimens*, Zygina* sp., and *Empoasca pteridis*
	Search Fruit flies and *Thrips*	*P. avium* (CromoG3)		17/07/2015	Presence of *Frankliniella occidentalis*
				18/08/2015	Presence of an exemplar on *Metcalfa pruinosa* and *Tephritis formosa*
				15/09/2015	Presence of *Dictya* sp.
				29/10/2015	Presence of *Zyginidia pullula* , *Thaumatomyia notata*
	Search Fruit flies and *Thrips*	*Wisteria* sp. (CromoG4)		17/07/2015	Presence of *Frankliniella intonsa*
				18/08/2015	Presence of an exemplar on *Halyomorpha halys* , *Heliothis peltigera* , and an exemplar on *Thaumatomyia notata*
				15/09/2015	Presence of *Plutella xylostella* and *Halyomorpha halys*
				29/10/2015	Presence of *Frankliniella occidentalis* , *Cicadella viridis* , *Zygina rhamni*
	Search Fruit flies and *Thrips*	*Citrus* sp. (CromoG5)		18/08/2015	Presence of *Zyginidia pullula*
				15/09/2015	Presence of *Ophraella communa* and *Dyctya* sp.
Hortus (26E-G)	Search Tephritidae	*P. avium* (ReATf1)		28/07/2015	Presence of *Scaphoideus titanus* , *Scymnus* sp. and *Stethorus* sp.
				09/09/2015	Presence of *Ophraella communa* , and *Hylaeus* sp.
				23/10/2015	Presence of *Ophraella communa, Metcalfa pruinosa,* and *Scymnus* sp.
	Search for *Thaumatotibia*	*Citrus* sp. (PgTh1)		17/09/2015	Presence of *Grapholita funebrana* and *Grapholita janthinana*
	Xylella fastidiosa	*P. avium*	2	22/09/2015	Absent
Area AMSA	Search Fruit flies and *Thrips*	Dumpsters (CromoG1amsa)		04/09/2015	Presence of *Zyginidia pullula* and *Cicadella viridis*
				05/11/2015	Presence of *Zyginidia pullula* and *Psammotettix* sp.
		Dumpsters (CromoG2amsa)		04/09/2015	Presence of *Typhaeaster Korea* , *Zyginidia pullula* , *Cicadella viridis* and *Ophraella communa*
		Dumpsters (CromoB1amsa)		04/09/2015	Presence of *Spodoptera exigua* and *Zyginidia pullula*
		Dumpsters (CromoB2amsa)		04/09/2015	Presence of *Spodoptera exigua* , of *Taylorilygus apicalis* , *Megalonotus* sp. (*sabulicola* or *chiragra*), *Zyginidia pullula* and *Ophraella communa*

ANNEX 4_ORGANISMS OF CLUSTERS

ORGANISMS OF CLUSTERS					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Biomediterranean Cluster	*Xylella fastidiosa*	*Olea europaea*	16	10/07/2015	Absent
			20	14/07/2015	Absent
			9	06/08/2015	Absent
		Myrtus communis	2	22/09/2015	Absent
	Search Fruit flies and *Thrips*	*Olea europaea* (CromoG23)		04/09/2015	Presence of *Xyleborinus saxeseni* , *Zyginidia pullula* , and *Clogmia albipunctata*
		Olea europaea (CromoG21)		29/09/2015	Presence of *Kalotermes flavicollis* , and psychodidae specimens
				26/10/2015	Presence of *Frankliniella occidentalis* , *Zyginidia pullula* , *Tomosvaryella kuthyi* , and psychodidae specimens
		Olea europaea (CromoG22)		26/10/201	Presence of *Zyginidia pullula* and psychodidae specimens
Coffee Cluster	*Xylella fastidiosa*	*Coffea arabica*	80	27/04/2015	Absent
	Identification of suspect insects	*Coffea arabica*	3	14/10/2015	Presence of *Pseudococcus viburni*
	Identification of suspect insects	*Coffea arabica*	1	14/10/2015	Presence of *Pseudococcus longispinus*
Cereals and tubers Cluster	Search for *Thrips*	*Chenopodium quinoa* (CromoBTh3)		04/06/2015	Presence of *Thrips tabaci* and *Frankliniella occidentalis*
				15/06/2015	Presence of *Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis,* and *Aeolothrips intermedius*
				10/07/2015	Presence of *Thrips tabaci*
				22/07/2015	Presence of *Frankliniella occidentalis* and *Lygus* sp.
				03/08/2015	Presence of dozens of *Thrips* of *Frankliniella occidentalis* and *Frankliniella intonsa.*
				08/09/2015	Presence of *Frankliniella occidentalis*
				24/09/2015	*Presence* about ten of *Thrips* of *Frankliniella intonsa* and *Thrips tabaci*
				30/10/2015	Presence of *Drosophila* sp.*, Eucallipterus tiliae, Omalium caesum,* and *Rhopalosiphum rufiabdominalis*
	Search leaf miners and *Thrips*	*Chenopodium quinoa* (Mined leaves)		15/06/2015	Presence of *Pegomya* sp.
	Search for *Thrips*	*Solanum tuberosum* (CromoGTh5)		15/06/2015	Presence of *Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis,* and *Aeolothrips intermedius*
				22/07/2015	Presence of *Frankliniella occidentalis*
				03/08/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis.*
				07/09/2015	Presence of dozens of *Thrips* of *Frankliniella occidentalis*
				24/09/2015	Presence around twenty *Thrips* of *Frankliniella intonsa* and *Microcephalothrips abdominalis*
				30/10/2015	Presence of *Thrips tabaci* and *Frankliniella occidentalis*
	Search for leaf miners	*Chenopodium quinoa* (parasitized caterpillar)		15/06/2015	Presence of *Autographa gamma* parasitized by *Euplectrus* sp.
	Search for *Tecia solanivora*	*Solanum tuberosum* (PgTe1)		04/08/2015	Presence of *Heliothis peltigerae* and *Monopis imella*
	Search for *Thaumatotibia*	*Sorghum vulgare* (PgTh3)		17/09/2015	Presence of *Monopis imella*
Fruits and vegetables cluster	Search Fruit flies and *Thrips*	*Malus* sp. (CromoB13)		22/06/2015	Presence of *Anaphothrips obscurus*
				23/07/2015	Presence of *Frankliniella occidentalis*
				25/08/2015	Presence of *Chorosoma schillingi*
				26/10/2015	Presence of *Hydroptila angulata, Calathus* sp.*, Stephanitis pyri*

ORGANISMS OF CLUSTERS					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Fruits and vegetables cluster	Search for *Thrips*	*Solanum lycopersicum* (CromoGTh2)		22/06/2015	Presence of *Thrips tabaci*
				23/07/2015	Presence of *Frankliniella occidentalis*
				03/08/2015	Presence of dozens of *Thrips* of *Frankliniella occidentalis.*
				08/09/2015	Presence of *Frankliniella occidentalis, Tuta absoluta* and *Anoecia* sp.
				24/09/2015	Presence about ten of *Thrips* of *Thrips tabaci*
				29/10/2015	Presence of *Frankliniella occidentalis*
	Search for *Keiferia lycopersicella*	*Solanum lycopersicum* (PgKe1		12/08/2015	Presence of *7 exemplar on Tuta absoluta*
				29/09/2015	Presence of *Tuta absoluta*
	Search for *Thaumatotibia*	*Citrus limon* (PgTh4)		17/09/2015	Presence of *Tuta absoluta* and *Cadra cautella*
	Search *Spodoptera* sp.	*Solanum lycopersicum* (PgSp2)		06/10/2015	Presence of *Ophraella communa* and *Hylaeus* sp.
Rise Cluster	*Xylella fastidiosa*	*Olea europaea*	4	04/06/2015	Absent
Spice Cluster	Search *Anthonomus*	*Capsicum* L. (CromoGAt2)		03/08/2015	Presence of *Eurydema ventralis* , *Ophraella communa* , and *Lygus pratensis.*
				11/09/2015	Presence of *Ophraella communa* , *Cicadella viridis* , *Orius majusculus* , *Nysius* sp., and *Hylaeus* sp.
				28/09/2015	Presence of *Lygus* sp.
Arid areas Cluster (back) (CromoBTh10) (24C)	Search for *Thrips*	Dumpsters		09/09/2015	Presence of *Frankliniella intonsa*
				24/09/2015	Presence of one hundred *Thrips* of *Frankliniella occidentalis.*
				27/11/2015	Presence of *Frankliniella occidentalis*

ANNEX 5_ORGANISMS OF PAVILIONS

ORGANISMS OF PAVILIONS					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Angola Pavilion	*Xylella fastidiosa*	*Olea europaea*	10	14/10/2015	Absent
	Search for harmful organisms subjected to phytosanitary restrictions	Banana soil		19/10/2015	Absent
Argentine Pavilion	*Xylella fastidiosa*	*Olea europaea*	5	06/05/2015	Absent
	Search *Thrips*	Entomological samples (CromoGTh9)		17/08/2015	Presence of *Frankliniella occidentalis*
				08/09/2015	Presence of *Agraylea sexmaculata* and *Psychoda alternata*
				28/09/2015	Absence of Thrips, presence of psychodidae specimens
				29/10/2015	Presence of psychodidae specimens
Austria Pavilion (back)	Search sawflies (Tenthredinidae)	Entomological samples (ReB2)		31/08/2015	Presence of *Hylaeus leptocephalus, Ophraella communa* and *Aproaerema anthyllidella*
Bahrain Pavilion	*Xylella fastidiosa*	*Olea europaea*	5	05/05/2015	Absent
	Rhynchophorus ferrugineus	Entomological samples (SRh3)		22/07/2015	Presence of *Oryctes nasicornis* (female)
Belgium Pavilion (back)	Search Fruit flies and Thrips	*Pyrus* sp (CromoB9)		25/08/2015	Presence of *Metcalfa pruinosa*
Birra Moretti Pavilion	*Xylella fastidiosa*	*Olea europaea*	2	06/05/2015	Absent
Brazil Pavilion	*Xylella fastidiosa*	*Coffea arabica*	12	23/04/2015	Absent
	Identification of suspect insects	Cocoons on *Brassica* sp.		01/06/2015	*Plutella xylostella*
	Identification of suspect insects	Chrysalis *Pieris* on *Brassica* sp.		01/06/2015	*Pieris rapae*
	Search *Colletotrichum demantium*	*Glycine max* leaves		04/06/2015	Presence of *Colletotrichum dematium* acervuli and spores; Presence of *Frankliniella occidentalis*
	Search *Thrips*	*Chenopodium quinoa* leaves		04/06/2015	Presence of *Frankliniella occidentalis*
	Search *Thrips*	*Solanum tuberosum* and *Glycine max* (CromoBTh4)		04/06/2015	Presence of *Frankliniella occidentalis*
				15/06/2015	Presence of *Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis* and *Aeolothrips intermedius*
				10/07/2015	Presence of *Frankliniella occidentalis*
				23/07/2015	Presence of *Frankliniella occidentalis*
				03/08/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis.*
				08/09/2015	Presence of *Tuta absoluta, Frankliniella occidentalis* and *Cicadella viridis*
				30/10/2015	Presence of *Thrips tabaci, Frankliniella occidentalis* and *Trechus quadristriatus*
	Search *Thrips*	*Abelmoschus esculentus* (CromoGTh6)		15/06/2015	Presence of *Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis,* and *Aeolothrips intermedius*
				10/07/2015	Presence of *Frankliniella occidentalis*
				23/07/2015	Presence of *Frankliniella occidentalis* and *Gypsonoma aceriana*
				08/09/2015	Presence of *Frankliniella occidentalis* , *Tuta absoluta, Agraylea sexmaculata* and *Mystacides azureus*
				30/10/2015	Presence of *Zyginidia pullula,* whiteflies and *Trechus quadristriatus*
	Search leaf miners and *Thrips*	*Glycine max*		27/07/2015	*Frankliniella occidentalis*
	Identification of suspect insects	*Coffea arabica*	3	14/10/2015	Presence of *Saissetia coffeae*
	Search phytoparasitic nematodes	Banana soil		19/10/2015	Absent
China Pavilion (back)	Search Thrips	Dumpsters(CromoBTh9)		08/09/2015	Presence of *Frankliniella occidentalis, Hydropsyche sp.* and *Sepedon sphegea*
				28/09/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis.*
				27/11/2015	Presence of *Frankliniella occidentalis*
	Xylella fastidiosa	*Olea europaea*	6	28/09/2015	Absent
	Search *Bursaphelenchus xylophilus* nematodes	Mulching		19/10/2015	Absent

ORGANISMS OF PAVILIONS					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Colombia Pavilion	*Xylella fastidiosa*	*Coffea arabica*	26	29/07/2015	Absent
	Search Fruit flies and *Thrips*	*Vitis* L. (CromoG7) back		20/07/2015	Presence of *Anaphothrips obscurus*
				27/08/2015	Presence of *Coenosia attenuata* of *Ophraella communa*
				28/09/2015	Presence of Dictya sp. specimens, caddis-fly, psychodidae specimens and *Cadra cautella*
				26/10/2015	Presence of *Hydroptila angulata* and psychodidae specimens
	Search for harmful organisms subjected to phytosanitary restrictions	Banana soil		19/10/2015	Absent
Coppini Olea Pavilion	*Xylella fastidiosa*	*Olea europaea*	2	04/06/2015	Absent
Biological Pavilion	*Xylella fastidiosa*	*Olea europaea*	1	14/07/2015	Absent
Eco Pasta&Pizza Pavilion	*Xylella fastidiosa*	*Olea europaea*	4	06/05/2015	Absent
United Arab Emirates Pavilion	*Xylella fastidiosa*	*Olea europaea*	1	07/05/2015	Absent
		Myrtus communis	2	22/09/2015	Absent
Enel Pavilion	*Xylella fastidiosa*	*Olea europaea*	1	06/05/2015	Absent
France Pavilion	*Xylella fastidiosa*	*Olea europaea*	2	05/05/2015	Absent
	Search *Thrips*	*Solanum lycopersicum* (CromoBTh1)		22/06/2015	*Thrips tabaci*
				23/07/2015	Presence of *Ophraella communa, Pieris sp., Frankliniella intonsa, Aeolothrips intermedius* and *Frankliniella occidentalis*
				07/09/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis* and *Nysius graminicola* specimens, *Cacoecimorpha pronubana* and *Sitotroga cerealella*
				24/09/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis.*
	Search for *Anthonomus eugenii*	*Capsicum* L. (CromoGAt8)		18/08/2015	Presence of 1 *Corythucha ciliata* , and 3 *Orius majusculus* specimens
				06/10/2015	Presence of *Halyomorpha halys, Sitotroga cerealella,* and *Monopis imella*
Indonesia Pavilion (back)	Search *Thrips*	Dumpsters (CromoBTh8)		17/08/2015	Presence of *Frankliniella occidentalis*
				07/09/2015	Presenze about ten of *Thrips* of *Frankliniella occidentalis, Xyleborinus saxeseni, Thaumatomya notata* and *Hydroptila* sp.
				24/09/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis* and *Thrips tabaci*
Iran Pavilion	*Xylella fastidiosa*	*Olea europaea*	5	05/05/2015	Absent
	Checking on walnut leaves	*Juglans* sp.		15/06/2015	Acaric erythrophic *Aceria erineus* and *Marssonina juglandis* spores
	Search *Bursaphelenchus xylophilus* nematodes	Mulching		19/10/2015	Absent
Kazakhstan Pavilion	Search for harmful organisms subjected to phytosanitary restrictions	Mulching		19/10/2015	Absent
Malaysia Pavilion	Search *Bursaphelenchus xylophilus* nematodes	Mulching		19/10/2015	Absent
Morocco Pavilion	*Xylella fastidiosa*	*Olea europaea*	15	05/05/2015	Absent
	Xylella fastidiosa	*Nerium oleander*	20	05/05/2015	Absent
	Search sawflies (Tenthredinidae)	Entomological samples (ReB7)		31/08/2015	Presence of *Athalia rosae rosae, Ophraella communa* and *Halyomorpha halys*
	Search *Bursaphelenchus xylophilus* nematodes	Mulching		16/10/2015	Absent
Nepal Pavilion	Search for *Anthonomus eugenii*	*Capsicum* L. (CromoGAt3)		03/08/2015	Presence of *Ophraella communa.*
				06/10/2015	Presence of *Ophraella communa* and *Lyctus africanus*
San Pellegrino Pavilion	*Xylella fastidiosa*	*Olea europaea*	2	04/06/2015	Absent
Save the children Pavilion	*Xylella fastidiosa*	*Olea europaea*	1	05/05/2015	Absent
Sicily Pavilion	*Xylella fastidiosa*	*Olea europaea*	1	06/05/2015	Absent
Slow food Pavilion	Search *Thrips*	*Solanum lycopersicum* (CromoGTh1)		22/07/2015	Presence of *Frankliniella occidentalis*
				03/08/2015	Presence of one hundred Thrips of *Frankliniella occidentalis* .Presence of *Diabrotica v. virgifera*

ORGANISMS OF PAVILIONS					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Slow food Pavilion	Search *Thrips*	*Solanum lycopersicum* (CromoGTh1)		07/09/2015	Presence about ten of *Thrip,* some belonging to of *Microcephalothrips abdominalis* . Presence of *Lygus pratensis, Ophraella communa, Chrysotus n.* sp. *choricus grp.*
				24/09/2015	Presence about twenty of *Thrips* of *Thrips tabaci*
				29/10/2015	Presence of *Frankliniella occidentalis*
	Search *Anthonomus*	*Capsicum* L. (CromoGAt4)		04/08/2015	Presence of *Diabrotica v. virgifera* , *Ophraella communa* , *Lygus pratensis* , *Stictocephala bisonia* and *Altica oleracea* specimens
				14/09/2015	Presence of *Ophraella communa* , and *Halyomorpha halys*
				06/10/2015	Presence of *Ophraella communa, Cicadella viridis* and *Luperomorpha xanthodera*
EU Pavilion	*Xylella fastidiosa*	*Olea europaea*	1	06/05/2015	Absent
Hungary Pavilion	*Search Anthonomus*	*Capsicum* L. (CromoGAt1)		03/08/2015	Presence of *Nysius graminicola, Lygus pratensis* and *Duponchelia fovealis*
				09/09/2015	Presence of *Ophraella communa, Cicadella viridis*
				29/09/2015	Presence of *Galerucella luteola* , *Cicadella viridis* , *Corticarina sp* ., *Alloxysta* sp. and *Cryptophagus pilosus*
	Search Fruit flies and Thrips	*Prunus avium* (CromoG20)		25/08/2015	Presence of *Ophraella communa* , *Fieberiella florii* , *Scymnus* sp., and *Stethorus* sp.
	Search Tecia solanivora	*Solanum tuberosum* (PgTe3)		03/09/2015	Presence of *Clepsis peritana* and *Scrobipalpa ocellatella*
Zero Pavilion (meadow at the back)	Search *Thrips*	Dumpsters (CromoBTh7)		09/09/2015	Presence of *Haplothrips leucanthemi*
				24/09/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis*
Cascina Triulza	Search for *Anthonomus eugenii*	*Capsicum* L. (CromoGAt7)		18/08/2015	Presence of 3 *Cicadella viridis* , and 1 *Nysius senecionis* specimens
				14/09/2015	Presence of *Cicadella viridis* , *Nysius* sp. and *Dictya* sp.
	Search Fruit flies e *Thrips*	*Diospyros kaki* (CromoG10)		20/07/2015	Presence of *Frankliniella intonsa* , *Aeolothrips intermedius* and *Haplothrips* sp.
				27/08/2015	Presence of *Eutettix* sp.or *Allygus atomarius*
				02/11/2015	Presence of *Zygina nivea, Ophraella communa,* and *Dictya sp.*
	Search sawflies (Tenthredinidae)	Entomological samples (ReB8)		31/08/2015	Presence of *Cryptochironomus supplicans* , *Mystacides azureus, Athalia rosae rosae, Ophraella communa, Cicadella viridis* , *Laodelphax striatellus* and *Nomophila noctuella*
	Search *Thaumatotibia leucotreta*	Entomological samples (PgTh5)		24/09/2015	Presence of *Pammene albuginana*
				23/10/2015	*Presence of Hydroptila angulata* and *Corticarina sp.*
Children park	Search *Thrips*	*Citrus limon* (CromoBTh2)		22/06/2015	Presence of *Thrips flavus*
				23/07/2015	Presence of *Thrips tabaci* and *Frankliniella intonsa*
				03/08/2015	Presence about ten of *Thrips* of *Frankliniella occidentalis.*
				08/09/2015	Presence of *Frankliniella intonsa* and *Semidalis* sp.(prob. *aleyrodiformis*)
				28/09/2015	Presence about ten of *Thrips* of *Thrips physapus*
				30/10/2015	Presence of *Anoecia* sp., *Hydroptila vectis*
Children park	Search for *Planococcus ficus*	*Ficus carica* (PlPf2)		29/07/2015	Presence of *Sigara striata*
	Search Diptera Tephritidae	*Prunus avium* (ReATf4)		05/08/2015	Presence of *Adalia bipunctata* , 1 *Rhagoletis cerasi* , 1 *Euscelidius variegatus* and 1 *Philaenus spumarius* specimens
				23/10/2015	Presence of *Psammotettix* sp. *(prob. confinis), Sitona hispidulus, Trichoptera, Orientus ishidae, Scymnus* sp., *Stethorus* sp., *Anoecia* sp., *Thaumatomyia notata* and *Euleia* sp.
	Search CTV	*Citrus limon* (PsCi2)		17/09/2015	Presence of *Dialeurodes citri*
	Search mealybugs (Planococcus citri)	*Citrus medica*		08/09/2015	Presence of *Planococcus citri*

ORGANISMS OF PAVILIONS					
Tested areas	**Analyzes carried out**	**Analyzed material**	**N. tested plants**	**Date**	**Results**
Fabbrica del Duomo (back)	Search Fruit flies and *Thrips*	*Citrus limon* (CromoG19)		27/08/2015	Presence of *Ophraella communa*
				30/10/2015	Presence of psychodidae specimens
Future food district	Search sawflies (Tenthredinidae)	*Malus* sp. (ReB4)		31/08/2015	Presence of *Eristalinus taeniops, Lygus pratensis, Ophraella communa* and *Oplodontha viridula.*
		Malus sp. (ReB6)		31/08/2015	Presence of *Eristalinus sp.* and *Ophraella communa*
Biodiversity Park	*Xylella fastidiosa*	*Olea europaea*	8	06/05/2015	Absent
		Rosmarinus officinalis	2	22/09/2015	Absent
		Olea europaea	12	05/10/2015	Absent
	Search for *Monochamus* sp.	*Pinus* sp. (MfMo1)		03/09/2015	Presence of *Apis mellifera* , *Polistes* sp., *Vespula* sp., and *Ips sexdentatus*
	Search Fruit flies and *Thrips*	*Vitis* L. (CromoG1)		17/07/2015	Presence of *Frankliniella occidentalis*
				18/08/2105	Presence of *Protapion fulvipes* and *Dioxyna sp.* (prob. *bidentis*)
				15/09/2015	Presence of *Orius majusculus* , *Scymnus* sp. and *Stethorus* sp.
				26/10/2015	Presence of *Acrolepiopsis assectella, Zygina rahmni, Ophraella communa,* and *Stethorus sp.*
		Citrus limon (CromoB12)		25/08/2015	Presence of *Chorosoma schillingi*
				29/09/2015	Presence of *Ophraella communa*
				26/10/2015	Presence of *Psammotettix* sp. (prob. *confinis*), *Anoecia* sp., psychodidae specimens and *Trixagus meybohmi*
	Search for *Planococcus ficus*	*Ficus carica* (PlPf1)		26/10/2015	Presence of *Laodelphax striatellus* , psychodidae specimens and *Macrosteles* sp.
	Search for *Sesia* sp.	*Malus* sp. (Sesia)		23/10/2015	Presence of *Corticarina sp., Ophraella communa,* and *Scymnus sp.*
	Search for *Spodoptera* sp.	*Zea mais* (PgSp1)		06/10/2015	Presence of *Metcalfa pruinosa* specimens
Stand Kinder+Sport	Search *Rhagoletis* sp.	*Prunus avium* (ReaRh1)		31/08/2015	Presence of *Hydroptila vectis* , *Hydropsyche modesta* , *Adalia bipunctata* , *Fieberiella florii* and *Ophraella communa*
	Search Fruit flies and *Thrips*	*Prunus* sp. (CromoG11)		02/11/2015	Presence of *Hydroptila vectis, Ophraella communa,* and *Zygina* sp.
UN Garden	Search Diptera Tephritidae	*Prunus avium* (ReATf7)		05/08/2015	Presence of 1 *Metcalfa pruinosa* , 7 *Ophraella communa* , 1 *Cicadella viridis* and 1 *Spodoptera exigua specimens*
				09/09/2015	Presence of *Eristalinus taeniops,* and *Ophraella communa*

ANNEX 6_ACCESS TO THE SITE

DATE	CIAMPITTI	CAVAGNA	SIENA	TANTARDINI	PATTI	ZANOTTI	REASON
20_06_2014	1	1					inspection with Selleri
13_01_2015	1	1					inspection with Cristina Martone
2_02_2015	1	1					installation of spore caps and apple baits mela1 and2
9_02_2015	1	1	1	1			check spore caps and apple baits mela1 and mela2
10_02_2015	1	1	1	1			check spore caps
16_02_2015	1		1		1	1	check spore caps and apple baits mela1, 2 and 3
23_02_2015	1		1		1	1	check spore caps and apple baits mela1, 2 and 3
02_03_2015	1				1	1	check spore caps and apple baits mela1, 2 and 3
09_03_2015					1	1	check spore caps and apple baits mela1, 2 and 3
16_03_2015					1	1	check spore caps and apple baits mela1, 2 and 3
23_03_2015					1	1	check spore caps and apple baits mela1, 2 and 3
30_03_2015					1	1	check spore caps and apple baits mela2 and mela3, new installation of apple bait mela1
01_04_2015	1	1			1	1	inspection to check the progress of planting
07_04_2015					1	1	check spore caps and apple baits mela1, mela2 and mela3
13_04_2015					1	1	check spore caps and apple baits mela1, mela2 and mela3
20_04_2015					1	1	check spore caps and apple baits mela1, mela2 and mela3
27_04_2015	1				1	1	check spore caps, apple baits mela1, mela2 and mela3 and sampling of olive trees samples on French pavillion for Xylella fastidiosa
04_05_2015	1				1	1	check spore caps and apple baits mela1, mela2 and mela3
05_05_2015	1	1			1	1	sampling of olive and oleander tree samples for Xylella fastidiosa and installation of buckets traps for Rhynchophorus
06_05_2015					1	1	sampling of olive tree samples for Xylella fastidiosa and installation of apple bait mela4 and mela5
06_05_2015					1	1	sampling of olive tree samples for Xylella fastidiosa and installation of apple bait mela4 and mela5
07_05_2015	1				1	1	sampling of olive and coffea trees samples for Xylella fastidiosa and installation of apple bait mela6
08_05_2015	1						
11_05_2015	1				1	1	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6. Installation of 2 Popillia traps and 6 chromotropic traps
14_05_2015					1	1	check apple baits mela1, mela2, mela3, mela4, and mela5. Installation of 1 trap for Sesia and 8 chromotropic traps
18_05_2015					1	1	check spore caps, apple baits mela1, mela2, mela3, mela4, and mela5. Installation of apple bait mela6.
19_05_2015					1	1	check buckets traps for Rhynchophorus and installation of 14 chromotropic traps
21_05_2015	1				1	1	installation of Planococcus citri trap, 1 white rebel trap and 2 multi-funnel traps for Anoplophora
25_05_2015					1	1	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6.
26_05_2015					1	1	installation of 4 chromotropic traps for Thrips, 1 plano trap for Planococcus and installation of 3 chromotropic traps
27_05_2015					1	1	installation of 2 Multi-funnel traps 1 for Monochamus and 1 for Anoplophora, 3 delta traps for Spodoptera, 1 plano trap for Planococus, check 16 chromotropic traps and reinstallation of 2 chromotropic traps
28_05_2015					1	1	installation of 4 multi-funnel traps, 3 for Anoplophora and 1 for Pityophthorus, 4 amarillo rebel traps for tephritidae and 2 chromotropic traps for Thrips

DATE	CIAMPITTI	CAVAGNA	SIENA	TANTARDINI	PATTI	ZANOTTI	REASON
01_06_2015	1	1			1	1	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6. Installation of 2 prism traps for Agrilus and 4 chromotropic traps for Thrips. Generic material sample for foliar mining check.
04_06_2015	1				1	1	Sampling of olive trees samples for Xylella fastidiosa, sampling of soy and quinoa samples for Thrips check, generic material sample for foliar mining check, check and replacement of 3 chromotropic traps
08_06_2015			1		1		check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6.
10_06_2015					1		installation of 3 Popillia japonica traps
15_06_2015	1				1	1	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6. check Popillia traps TFly2. installation of 3 Popillia traps. Sampling of 4 plant material samples. installation of 3 pagodas traps for Thaumatotibia and 1 pagodas trap for Tecia. check rhynchophorus buckets trap SRh2. check and replacemen 4 chromotropic traps for Thrips
22_06_2015					1	1	check spore caps and sampling apple baits mela1, mela2, mela3, mela4, mela5 and mela6. check 5 rebell traps, installation of 2 chromotropic traps, replacemen of 9 chromotropic traps, check 2 planotrap for planococcus, sampling of 1 Citrus sample for Citrus tristeza virus, replacemen of 3 chromotropic traps for thrips, check 3 pagodas traps for Spodoptera, check Tfly1 for Popillia and reinstallation of 1 chromotropic traps. Generic sampling of 2 aphids on wisteria and 1 mole samples.
22_06_2015					1	1	check spore caps and sampling apple baits mela1, mela2, mela3, mela4, mela5 and mela6. check 5 rebell traps, installation of 2 chromotropic traps, replacemen of 9 chromotropic traps, check 2 planotrap for planococcus, sampling of 1 Citrus sample for Citrus tristeza virus, replacemen of 3 chromotropic traps for thrips, check 3 pagodas traps for Spodoptera, check Tfly1 for Popillia and reinstallation of 1 chromotropic traps. Generic sampling of 2 aphids on wisteria and 1 mole samples.
24_06_2015					1	1	installation of di 2 multi-funnel for anoplofora, check 2 trap popillia Tfly, check and replacemen di 3 pagode for spodoptera, installation of pagoda for 2 taumatotibia, installation of 1 pagoda for Tecia, installation of di 1 pagoda for keiferia, check and replacemen di 1 trap cromotropica.
25_06_2015					1	1	installation of 1 multi-funnel trap for monochamus spp., installation of 8 prism traps for Agrilus spp.
29_06_2015					1	1	Sampling and replacemen of spore caps. installation of 2 multi-funnel traps for Anoplophora spp., 1 multi-funnel trap for Monochamus spp. and installation of 4 chromotropic traps for Anthonomus spp.
07_07_2015	1				1	1	check spore caps
09_07_2015	1				1	1	check 10 traps for popillia with sampling of 4 samples, shifted Popillia trap Tfly3 in Austria pavillion, check and sampling of 1 planotrap for Planococcus
10_07_15					1	1	Sampling of 6 olive samples for Xylella fastidiosa; check apple bait Mela1, Mela2, Mela3, Mela6; check and replacemen traps 4 Thrips CromoBTh and CromoGTh; Installation of 4 white rebell traps for sawflies, sampling of generic sample of thrips Var11

DATE	CIAMPITTI	CAVAGNA	SIENA	TANTARDINI	PATTI	ZANOTTI	REASON
13_07_15					1	1	installation of 1 pagodas traps for tecia; installation of di 3 rebell Amarillo traps for tephritidae and 1 rebell Amarillo for Rhagoletis; installation of 4 yellow cromotropic traps
14_07_15	1	1			1	1	check spore caps; installation of 1 yellow chromotropic trap and 2 yellow chromotropic traps for anthomonus, sampling of 7 olive samples for Xylella fastidiosa
15_07_15	1				1	1	installation of 7 chromotropic thrips traps; installation of 2 pagodas traps for Keiferia, repositioning of 1 white rebell trap; check of 2 popillia traps
16_07_15					1	1	installation of 3 chromotropic traps for Thrips; installation of 2 chromotropic traps for Anthonomus. Check of 2 Agrilus traps.
17_07_15	1	1				1	replacement of 4 chromotropic traps.
20_07_15					1		Sampling and replacemen of spore caps. installation of 2 white rebel traps for tephritidae, replacemen 6 simple chromotropic traps and 1 chromotropic trap for thrips, reinstallation of 4 chromotropic traps, check 2 traps for Rhynchophorus and 3 for Popillia.
22_07_15					1		Replacement of 8 chromotropic simple traps and 4 chromotropic traps for Thrips, reinstallation of 4 chromotropic traps, check 1 trap for Rhynchophorus, 3 pagodas traps for Thaumatotibia, 1 pagodas traps for Spodoptera, 2 for Tecia and 2 amarillo rebel trapsfor tephritidae.
23_07_15					1		Replacement of 5 chromotropic simple traps and 5 chromotropic traps for Thrips, check 1 pagodas trap for Thaumatotibia, 1 pagodas trap for Spodoptera and 6 Popillia traps with sampling of 2 samples.
27_07_15					1	1	Reinstallation of 6 apple baits, sampling and replacemen spore caps, repositioning of multi-funnel trap for Anoplophora, sampling of thrips material, check 5 TFly traps for Popillia
28_07_15					1	1	Reinstallation of buckets trap for Rhynchophorus , check and cleaning of 1 rebell amarillo traps, check of 1 white rebell trap and 1 amarillo rebell trap, detection of 1 spy plant for Agrilus
29_07_15	1				1	1	Reinstallation of 1 yellow chromotropic trap, change pheromons lure for 3 spodoptera pagodas traps, replacemen of 1 white rebell trap, sampling of 23 coffea plants, placement of 1 multi-funnel traps for monochamus, 2 yellow chromotropic traps and 2 blue chromotropic trap, entomological material sample for planococcus ficus on plano trap
30_07_15					1	1	check apple baits mela1, mela2, mela3, mela4, mela5 and mela6.
03_08_15						1	Sampling and replacemen spore caps, change pheromone lures and replacemen of 9 thrips chromotropic traps, check 2 thrips traps, change pheromone lures and replacemen of 3 anthonomus traps
04_08_2015						1	check traps and change pheromone lures on 2 pagodas traps for Tecia, sampling of PgTe1 sample, check and replacement anthonomus trap, change pheromone lure on sesia trap, check and replacemen pheromone lures on 3 thaumatotibia traps, repositioning CromoG11 and yellow thrips chromotropic trap CromoGTh9 pheromone lure because disappeared;
05_08_2015						1	installation of 3 yellow chromotropic traps, check and sampling of 10 Popillia Tfly traps, check 1 amarillo rebell trap for Rhagoletis, check and sampling of 2 amarillo rebell traps, check 1 yellow chromotropic trap for Anthonomus (disappeared), check blue chromotropic trap for Thrips

DATE	CIAMPITTI	CAVAGNA	SIENA	TANTARDINI	PATTI	ZANOTTI	REASON
06_08_2015	1	1				1	sampling of 8 olive samples from Biomediterranean cluster for Xylella fastidiosa
10_08_2015						1	Sampling and replacemen spore caps; replacemen of sesia trap
11_08_2015	1					1	check of 2 prism traps for Agrilus
12_08_2015						1	Check of 5 yellow chromotropic traps for Anthonomus and 1 replacement, check of 3 chromotropic traps for Thrips, 1 yellow chromotropic trap for fruit flies, check and replacement of 1 pagoda trap for tecia. Check of 1 multifunell trap for anoplophora, 1 white rebell trap, check and replacement of 1 pagoda trap for Keiferia.
17_08_2018					1	1	Sampling and replacemen spore caps. Check 4 buckets traps for Rhynchophorus, replacement of 4 blue chromotropic traps, 4 yellow chromotropic traps and pheromone lures for Thrips, check 2 pagodas traps for Keiferia with replacemen of pheromon lures and 1 of trap.
18_08_2015					1	1	Replacement of 4 chromotropic traps for Anthonomus and pheromone lures. Replacemen of 8 yellow chromotropic traps and 4 blue chromotropic traps.
20_08_2015					1	1	
25_08_2015					1	1	Replacement of 6 apple baits, 11 chromotropic traps, and reinstallation of 2 chromotropic traps CromoB3 and CromoG11, only check for CromoB4 and CromoB2, check and replacemen spore caps, check Tfly trap for Popillia.
26_08_2015	1	1			1	1	New installation of pagoda trap for Keiferia; check 9 multi-funnel traps for Anoplophora; check 1 multi-funnel trap for Monochamus, 6 TFly traps for Popillia; Finding 3 spy plants for Anoplophora, 2 spy plants for Agrilus; 1 spy plant for Erwinia.
27_08_2015					1	1	Replacement of 7 chromotropic traps, new installation of 1 chromotropic trap, finding 1 spy plant for Citrus tristezza virus.
28_08_2015					1	1	check apple baits mela1, mela2, mela3, mela4, mela5 and mela6.
31_08_2015	1				1	1	Sampling and replacemen spore caps, uninstall 6 white rebel traps and 1 amarillo rebel traps for Rhagoletis
01_09_2015	1						check treatments of Qatar pavillion
02_09_2015					1	1	uninstall 2 white rebel traps and 1 pagoda trap for Tecia, new installation of multi-funnel trap for Pityophthorus with pheromon lures replacemen, check and replacemen of pheromone lures for 6 prism traps for Agrilus with sampling of 3 traps.
03_09_2015					1	1	uninstall di 2 pagodas traps for Tecia, check 3 multi-funnel traps for Anoplophora and replacemen 2 pheromone lures, check and replacemen pheromone lure of Monochamus trap with sampling, replacemen 4 pheromone lures of Agrilus prism traps with sampling. Finding 3 spy plants for Phythophthora and 1 for Bursaphelencus and Gibberella.
04_09_2015						1	check and replacemen of 2 blue chromotropic traps, 1 Monochamus traps with replacement of pheromone lure, and sampling. Check 5 yellow chromotropic traps with replacemen of 4 traps. Finding 2 spy plants for Erwinia, 1 spy plant for Sharka and 1 for flavescence dorée of grapevine.

DATE	CIAMPITTI	CAVAGNA	SIENA	TANTARDINI	PATTI	ZANOTTI	REASON
07_09_2015					1	1	Sampling and replacemen spore caps. Check 2 buckets traps for Rhynchophorus, check 6 and replacemen 5 yellow chromotropic traps and 4 blue chromotropic traps for Thrips. Check 1 Popillia trap, 1 Planococcus trap and 1 Thaumathotibia trap.
08_09_2015					1	1	Check 2 buckets traps for Rhynchophorus, check and replacemen 4 yellow chromotropic traps and 4 blue chromotropic traps for Thrips, check 3 Popillia traps, and 1 Planococcus trap, replacemen Monochamus traps pheromone lure, and sampling citrus leaves for cochineal.
09_09_2015					1	1	check and replacemen 2 chromotropic traps for Thrips; check and sampling 4 rebell amarillo traps and 4 yellow chromotropic traps for Thrips; check 1 Rebell Amarillo trap; new installation of rebell Amarillo trap (with chromotropic trap); check 1 planotrap for Planococcus; check 4 chromotropic traps for anthonomus with sampling.
11_09_2015	1				1	1	new installation of 2 traps for Anthonomus; check 2 rebell amarillo traps; check and sampling 1 chromotropic traps for Anthomonus
14_09_2015					1	1	Sampling and replacement spore caps. Replacement 5 apple baits for Phytophthora, check and replacemen 2 traps for Anthonomus.
15_09_2015					1		check and replacemen 5 yellow chromotropic traps, check 1 Planococcus trap and 1 for Thaumathotibia. New installation of 1 apple bait for Phytophthora
17_09_2015			1		1		Replacement 4 blue and 3 yellow chromotropic traps, new installation of 1 yellow chromotropic trap for Thrips, check 6 apple baits for Phytophthora, 2 traps for Popillia, disinstallation of 3 pagodas and replacemen of 1 for Thaumatotibia, Sampling of Arbutus branches for Phytophthora, and Citrus leaves from spy plant for Citrus tristezza virus.
18_09_2015	1		1		1		Meeting with press.
21_09_2015						1	Meeting with press, sampling and replacemen spore caps
22_09_2015					1	1	Sampling of rosemary, myrtle, cherry and broom branches for Xylella fastidiosa, and 1 sample for Erwinia.
24_09_2015					1	1	Replacement 6 yellow and 5 blue chromotropic traps for Thrips and their pheromone lures, change only pheromone lure for 2 blue and 2 yellow chromotropic traps for Thrips, new installation of 1yellow chromotropic trap, check 1 planotrap for Planococcus ficus, 1 trap for popilia and replacemen 1 trap for Thaumatotibia.
25_09_2015	1	1					Meeting with Bosnian Workgroup
28_09_2015					1	1	Sampling and replacement spore caps. Replacemen 4 blue and 2 yellow chromotropic traps for Thrips and their pheromone lures. Check 1 blue and 2 yellow chromotropic traps for Thrips, replacemen 1 yellow chromotropic trap. Sampling olive branches for Xylella fastidiosa and uninstall 1 trap for Anthonomus with sampling.
29_09_2015					1	1	Uninstall 3 multi-funnel traps for Anoplophora and sampling material of them, uninstall 1 multi-funnel traps for Monochamus, pagodas traps for Keiferia, with sampling of each of them, check 1 Popillia trap, uninstall 1 trap for Anthonomus with sampling, check and replacemen 4 blue and 6 yellow chromotropic traps, only check for 2 yellow and 1 blue chromotropic trap, check and replacemen 1 amarillo rebel trap for Tephritidae, new installation of 1 yellow and 1 blue chromotropic trap and 1 yellow chromotropic trap for Thrips.

DATE	CIAMPITTI	CAVAGNA	SIENA	TANTARDINI	PATTI	ZANOTTI	REASON
02_10_2015						1	Replacement 6 apple baits for Phytophthora
05_10_2015					1	1	Sampling and replacement spore caps, check apple baits mela1, mela2, mela3, mela4, mela5 and mela6, uninstall 4 multi-funnel traps for Anoplophora, sampling of 4 olive trees samples for Xylella fastidiosa.
06_10_2015						1	Uninstalling 5 CromoG traps for Anthonomus and 3 pagodas traps for Spodoptera
13_10_2015					1		Sampling and replacement spore caps.
14_10_2015	1				1	1	Sampling 6 coffea trees samples and 2 mulch bark samples
16_10_2015					1	1	Sampling 2 mulch bark samples.
19_10_2015					1	1	Sampling and replacement spore caps, 3 mulch bark samples and 4 soil samples for banana nematodes. Replacement 6 apple baits for Phytophthora and check 1 trap for Popillia
22_10_2015					1	1	Sampling 6 apple baits for Phytophthora
23_10_2015					1	1	Sampling and uninstall 7 Rebel traps for Tephritidae, 2 traps for Thaumatotibia and 1 trap for Sesia
26_10_2015					1	1	Sampling and replacement spore caps, uninstall 2 Popillia traps, sampling and uninstall 2 blue and 4 yellow chromotropic traps, disappeared 1 yellow chromotropic trap (CromoG23), sampling and uninstall 1 Planococcus trap
28_10_2015	1	1				1	Uninstall 5 Popillia traps
29_10_2015					1	1	Uninstall 4 buckets traps for rhynchophorus and 2 traps for Popillia, sampling and uninstall 1 blue and 6 yellow chromotropic traps for Thrips, 5 blue and 8 yellow chromotropic traps, 1 Planococcus trap, and disappeared 1 yellow chromotropic trap (CromoG5)
30_10_2015	1		1		1	1	Uninstall and sampling 10 chromotropic traps for fruit flies, 6 for Thrips, 1 multi-funnel trap for Anoplophora and 1 planotrap for planococcus; Uninstall 1 multi-funnel trap for pityophthorus and 1 multi-funnel trap for Monochamus. 2 yellow (CromoG18, CromoG20) and 1 blue (CromoB2) chromotropic traps was disappeared; Uninstall 3 apple baits; Uninstall and sampling spore caps.
02_11_2015					1	1	check and uninstall 1 blue and 2 yellow chromotropic traps and 1 blue chromotropic trap for thrips. Check and uninstall 1 Popillia trap, 1 blue chromotropic trap (CromoB3) was disappeared
05_11_2015					1	1	check and uninstall 2 multi-funnel traps for anoplophora, check and replacement 4 Amsa chromotropic traps, installation of 2 chromotropic traps for thrips, 1 yellow and 2 blue chromotropic traps was disappeared (CromoB1_amsa, CromoB2_amsa, CromoG2_amsa)
10_11_2015					1	1	uninstall and sampling 8 prism traps for Agrilus
16_11_2015					1	1	uninstall and sampling 1 prism trap for Agrilus
27_11_2015						1	uninstall and sampling 1 prism trap for Agrilus, 1 yellow and 4 blue chromotropic traps for Thrips. Check for CromoBTh7 (disappeared)
18_01_2016					1	1	Installation 3 apple baits for Phytophthora.
25_01_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
01_02_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
08_02_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
15_02_2016						1	Sampling and replacement 2 apple baits for Phytophthora, and new installation of 1apple bait.

DATE	CIAMPITTI	CAVAGNA	SIENA	TANTARDINI	PATTI	ZANOTTI	REASON
22_02_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
01_03_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
07_03_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
14_03_2016						1	Sampling and replacement 3 apple baits for Phytophthora, check 3 spy plants for Erwinia amylovora.
21_03_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
29_03_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
04_04_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
11_04_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
18_04_2016						1	Sampling and replacement 3 apple baits for Phytophthora.
TOTALE GIORNATE	36	14	8	2	87	105	

ANNEX 7_ACTIVITIES

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
20_06_2014	inspection with Selleri			
13_01_2015	inspection with Cristina Martone			
2_02_2015	installation of spore caps and apple baits mela1 and2	/		
9_02_2015	check spore caps and apple baits mela1 and mela2	CAP1 and Guisa1 (x mela)	Cap_0902: Microscope observations	Presence of conidia of *Alternaria* sp. and *Diplodia* sp.
			Guisa1: Search *Phytophthora* sp.	Absence of oomycetes
10_02_2015	check spore caps	CAP1 and Guisa1 (x mela)	Cap1_1002: Microscope observations	Presence of *Alternaria* sp. conidia and *Diplodia* sp.
			Guisa1: Search *Phytophthora* sp.	Absence of oomycetes.
16_02_2015	check spore caps and apple baits mela1, 2 and 3	mela1_1602; mela2_1602; cap1_1602	Cap1_1602: Microscope observations	Absence of fungal spores.
			mela1_1602: Search *Phytophthora* sp.	Absence of *Phytopththora* ; insulation and presence of *Pythium litorale* confirmed by sequencing.
			mela2_1602: Search *Phytophthora* sp.	Absence of oomiceti.
23_02_2015	check spore caps and apple baits mela1, 2 and 3	mela2_2302; cap_2302	Cap_2302: Microscope observations	Absence of fungal spores.
			mela2_2302: Search *Phytophthora* sp.	Absence of oomiceti, presence of *Alternaria alternata* (confirmed by sequencing)
02_03_2015	check spore caps and apple baits mela1, 2 and 3	mela2_0203; mela3_0203; cap_0203	Cap1_0203: Microscope observations	Presence of *Leptosphaeria* sp. spores, and *Cladosporium* sp.
			mela1_0203	Insulation and presence of *Mucor* sp., and *Galactomyces geotrichum* confirmed by sequencing
			mela3_0203	Insulation and presence of Mucor sp., and Galactomyces geotrichum confirmed by sequencing.
09_03_2015	check spore caps and apple baits mela1, 2 and 3	mela3_0903; cap_0903	Cap1_0903: Microscope observations	Presence of *Cladosporium* sp, *Alternaria* sp., *Epicoccum* sp.
			mela3_0903: Search *Phytophthora* sp.	Absent
16_03_2015	check spore caps and apple baits mela1, 2 and 3	mela1_1603; mela2_1603; mela3_1603; cap_1603	Cap1_1603:Microscope observations	Presence of *Alternaria* sp.
			mela1_1603:Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela2_1603:Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela3_1603:Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
23_03_2015	check spore caps and apple baits mela1, 2 and 3	mela3_2303; cap1_2303	Cap1_2303: Microscope observations	Presence of *Sphaeropsis* sp. spores
			mela3_2303: Search *Phytophthora* sp.	Absent
30_03_2015	check spore caps and apple baits mela2 and mela3, new installation of apple bait mela1	mela2_3003; mela3_3003 cap1_3003	Cap1_3003: Microscope observations	Presence of *Alternaria* sp.
			mela2_3003:Search *Phytophthora* sp.	Absent
			mela3_3003:Search *Phytophthora* sp.	Absent
01_04_2015	inspection to check the progress of planting	/		
07_04_2015	check spore caps and apple baits mela1, mela2 and mela3	mela1_0704; mela2_0704; mela3_0704; cap1_0704	Cap1_0704: Microscope observations	Presence of *Cladosporium* sp. and *Alternaria* sp.
			mela1_0704: Search *Phytophthora* sp.	Presence of *Clonostachys rosea*
			mela2_0704: Search *Phytophthora* sp.	Presence of *Candida* sp.
			mela3_0704: Search *Phytophthora* sp.	Presence of *Galactomyces geotrichum*
13_04_2015	check spore caps and apple baits mela1, mela2 and mela3	mela1_1304; mela2_1304; mela3_1304; cap1_1304	Cap1_1304: Microscope observations	Presence of *Diplodia* sp. spores, *Cladosporium* sp., *Alternaria* sp.
			mela1_1304: Search *Phytophthora* sp.	Absent
			mela1_1304: Search Phytophthora sp.	Presence of *Mucor* sp.
			mela1_1304: Search Phytophthora sp.	Presence of *Mucor* sp.
20_04_2015	check spore caps and apple baits mela1, mela2 and mela3	mela1_2004; mela2_2004; mela3_2004; cap1_2004	Cap1_2004: Microscope observations	Presence of *Alternaria* sp. spores, *Tilletia* sp., *Cladosporium* sp.
			mela1_2004:Search *Phytophthora* sp.	Presence of *Trichoderma asperellum*
			mela2_2004:Search *Phytophthora* sp.	Absent
20_04_2015	check captaspore ed esche mela 1, mela 2 and mela 3	mela1_2004; mela2_2004; mela3_2004; cap1_2004	mela3_2004:Search *Phytophthora* sp.	Absent
27_04_2015	check spore caps, apple baits mela1, mela2 and mela3 and sampling of olive trees samples on French pavillion for Xylella fastidiosa	mela1_2704; mela2_2704; mela3_2704; cap1_2704, 4373957 xyl, 4373958 xyl	Cap1_2704: Microscope observations	Presence of *Alternaria* sp. spores e *Tilletia* sp.
			mela1_2704:Search *Phytophthora* sp.	Presence of *Mucor* sp.
			mela2_2704:Search *Phytophthora* sp.	Presence of *Mucor* sp.
			mela3_2704:Search *Phytophthora* sp.	Presence of *Mucor* sp.
			Sample for Xylella: 4373957	Absent
			Sample for Xylella: 4373958	Absent
04_05_2015	check spore caps and apple baits mela1, mela2 and mela3	mela1_0405; mela2_0405; mela3_0405; cap1_0405	Cap1_0405: Microscope observations	Presence of *Alternaria* sp. spores, *Cladosporium* sp.
			mela1_0405: Search *Phytophthora* sp.	Absent
			mela2_0405: Search *Phytophthora* sp.	Presence of *Fusarium* sp.
			mela3_0405: Search *Phytophthora* sp.	Presence of *Mucor* sp.

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
05_05_2015	sampling of olive and oleander tree samples for Xylella fastidiosa and installation of buckets traps for Rhynchophorus	SRh1; SRh2; SRh3; SRh4; 4373966; 4373965; 3021117; 4095362; 3021118; 4373967	Sample for Xylella: 4373966	Absent
			Sample for Xylella: 4373965	Absent
			Sample for Xylella: 3021117	Absent
			Sample for Xylella: 4095362	Absent
			Sample for Xylella: 3021118	Absent
			Sample for Xylella: 4373967	Absent
06_05_2015	sampling of olive tree samples for Xylella fastidiosa and installation of apple bait mela4 and mela5	mela4; mela5; 4373869; 4373872; 4373865; 4373858; 4373866; 4373859; 4373874; 4373864; 4373863; 4373867; 4373873; 4373857; 4373856; 4373855; 4373854; 4373862; 4373861; 4373860; 4373870; 4373868; 4094729	Sample for Xylella: 4373869	Absent
			Sample for Xylella: 4373872	Absent
			Sample for Xylella: 4373865	Absent
			Sample for Xylella: 4373858;	Absent
			Sample for Xylella: 4373866	Absent
			Sample for Xylella: 4373859	Absent
06_05_2015	sampling of olive tree samples for Xylella fastidiosa and installation of apple bait mela4 and mela5	mela4; mela5; 4373869; 4373872; 4373865; 4373858; 4373866; 4373859; 4373874; 4373864; 4373863; 4373867; 4373873; 4373857; 4373856; 4373855; 4373854; 4373862; 4373861; 4373860; 4373870; 4373868; 4094729	Sample for Xylella: 4373874	Absent
			Sample for Xylella: 4373864	Absent
			Sample for Xylella: 4373863	Absent
			Sample for Xylella: 4373867	Absent
			Sample for Xylella: 4373873	Absent
			Sample for Xylella: 4373857	Absent
			Sample for Xylella: 4373856	Absent
			Sample for Xylella: 4373855	Absent
			Sample for Xylella: 4373854	Absent
			Sample for Xylella: 4373862 (Birra Moretti)	Absent
			Sample for Xylella: 4373861	Absent
			Sample for Xylella: 4373860	Absent
			Sample for Xylella: 4373870	Absent
			Sample for Xylella: 4373868	Absent
			Sample for Xylella: 4094729	Absent
07_05_2015	sampling of olive and coffea trees samples for Xylella fastidiosa and installation of apple bait mela6	mela6; 4373882; 4373871; 4373876; 4373875; 4373877; 4373878; 4373879; 4373881	Sample for Xylella: 4373882	Absent
			Sample for Xylella: 4373871	Absent
			Sample for Xylella: 4373876	Absent
			Sample for Xylella: 4373875	Absent
			Sample for Xylella: 4373877	Absent
			Sample for Xylella: 4373878	Absent
			Sample for Xylella: 4373879	Absent
			Sample for Xylella: 4373881	Absent
08_05_2015				
11_05_2015	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6. Installation of 2 Popillia traps and 6 chromotropic traps	mela1_1105; mela2_1105; mela3_1105; mela4_1105; mela5_1105; mela6_1105; cap1_1105; Tfly1; Tfly2; CromoG1; CromoG2; CromoG3; CromoG4; CromoG5; CromoG6	Cap1_1105: Microscope observations	Presence of *Alternaria alternata* , *Diplodia* sp., *Cladosporium* sp.
			mela1_1105: Search *Phytophthora* sp.	Absent
			mela2_1105: Search *Phytophthora* sp.	Presence of *Fusarium tricinctum*
			mela3_1105: Search *Phytophthora* sp.	Presence of *Phytopthora lacustris*
			mela4_1105: Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela5_1105: Search *Phytophthora* sp.	Presence of *Trichoderma koningopsis*
			mela6_1105: Search *Phytophthora* sp.	Absent
14_05_2015	check apple baits mela1, mela2, mela3, mela4, and mela5. Installation of 1 trap for Sesia and 8 chromotropic traps	mela1_1405; mela2_1405; mela3_1405; mela4_1405; mela5_1405; PgSe; CromoG7; CromoG8; CromoG9; CromoG10; CromoB1; CromoB2; CromoB3; CromoB4		
18_05_2015	check spore caps, apple baits mela1, mela2, mela3, mela4, and mela5. Installation of apple bait mela6.	cap1_1805; mela1_1805; mela2_1805; mela3_1805; mela4_1805; mela5_ 1805; mela6_1805	Cap1_1805: Microscope observations	Presence of *Alternaria* sp., *Epicoccum* sp..
			mela1_1805:Search *Phytophthora* sp.	Absent
			mela2_1805:Search *Phytophthora* sp.	Presence of *Galactomyces* sp.
			mela3_1805:Search *Phytophthora* sp.	Absent
			mela4_1805:Search *Phytophthora* sp.	Absent
			mela5_1805:Search *Phytophthora* sp.	Absent
19_05_2015	check buckets traps for Rhynchophorus and installation of 14 chromotropic traps	CromoG11; CromoG12; CromoG13; CromoG14; CromoG15; Cromo G16; CromoG17; CromoB5; CromoB6; CromoB7; CromoB8; CromoB9; CromoB10; CromoB11		
21_05_2015	installation of Planococcus citri trap, 1 white rebel trap and 2 multi-funnel traps for Anoplophora	Plpc1; Reb1; MfAn1; MfAn2		

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
25_05_2015	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6.	mela1_2505; mela2_2505; mela3_2505; mela4_2505; mela5_ 2505; mela6_2505; cap1_2505;		
26_05_2015	installation of 4 chromotropic traps for Thrips, 1 plano trap for Planococcus and installation of 3 chromotropic traps	CromoTh1, PlPf1, CromoB12, CromoBTh1, CromoGth2, CromoB12, CromoGTh2, CromoB13, CromoB14, CromoB15		
27_05_2015	installation of 2 Multi-funnel traps 1 for Monochamus and 1 for Anoplophora, 3 delta traps for Spodoptera, 1 plano trap for Planococus, check 16 chromotropic traps and reinstallation of 2 chromotropic traps	MfMo1, MfAn3, PgSp1, PgSp2, PgSp3, PlPf2, CromoG1_2705, CromoG2_2705, CromoG3_2705, CromoG4_2705, CromoG6_2705, CromoG8_2705, CromoG9_2705, CromoG12_2705, CromoG13_2705, CromoG14_2705, CromoB1_2705, CromoB2_2705, CromoB5_2705, CromoB6_2705, CromoB7_2705, CromoB11_2705, CromoB16_2705, CromoG5, CromoB2	CromoG1_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG2_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG3_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG4_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG6_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG8_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG9_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG12_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG13_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoG14_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoB1_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoB2_2705: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
			CromoB5_2705: visual analysis and stereo-microscopic observation	Presence of Thrips, less than 10 individuals, of *Thrips tabaci* species
			CromoB6_2705: visual analysis and stereo-microscopic observation	Presence a dozen of Thrips, of *Thrips tabaci* and *Anaphothrips obscurus* species
			CromoB7_2705: visual analysis and stereo-microscopic observation	Presence a dozen of Thrips, of *Thrips trehernei* species
			CromoB11_2705: visual analysis and stereo-microscopic observation	Presence a dozen of Thrips, of *Thrips hawaiiensis* and *Thrips major* species
			CromoB16_2705: visual analysis and stereo-microscopic observation	Presence of a dozen of Thrips, of *Thrips tabaci* species
28_05_2015	installation of 4 multi-funnel traps, 3 for Anoplophora and 1 for Pityophthorus, 4 amarillo rebel traps for tephritidae and 2 chromotropic traps for Thrips	MfAn4, MfAn5, MfAn6, MfPi1, ReATf1, ReaTf2, ReATf3, ReATf4, CromoGTh3, CromoGTh4		
01_06_2015	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6. Installation of 2 prism traps for Agrilus and 4 chromotropic traps for Thrips. Generic material sample for foliar mining check.	mela1_0106; mela2_0106; mela3_0106; mela4_0106; mela5_ 0106; mela6_0106; cap1_0106; PgAr1; PgAr2; CromoBTh3; CromoBTh4; CromoGTh5; CromoGTh6; Var2_0106; Var3_0106	Var2_0106: breeding and identification	*Pieris rapae*
			Var3_0106: breeding and identification	*Plutella xylostella*
04_06_2015	Sampling of olive trees samples for Xylella fastidiosa, sampling of soy and quinoa samples for Thrips check, generic material sample for foliar mining check, check and replacement of 3 chromotropic traps	4094708; 4094710; 4373959;4373960;4373961; CromoBTh3_0406; CromoBTh4_0406; CromoGTh5_0406; Var1_0406; Var4_0406	CromoBTh3_0406:visual analysis and stereo-microscopic observation	Presence of a dozen of Thrips, of *Thrips tabaci* and *Frankliniella occidentalis* species
			CromoBTh4_0406:visual analysis and stereo-microscopic observation	Presence of hundreds of Thrips of *Frankliniella occidentalis* species
			CromoGTh5_0406:visual analysis and stereo-microscopic observation	Absence of Thrips
			Var1_0406:breeding and identification	Unidentified for parasitization of bred specimens
			Var4_0406:breeding and identification	Absence
			Sample of Soy 4094708: Search *Colletotrichum dematium* and Thrips identification	Presence of *Colletotrichum dematium* acervuli and spores, agent of soy anthrax; Thrips of *Frankliniella occidentali* s species
			Sample of Quinoa for Thrips: 4094710 Molecular analysis	Thrips of *Frankliniella occidentalis* species
			Sample for Xylella: 4373959	Absent
			Sample for Xylella: 4373960	Absent
			Sample for Xylella: 4373961	Absent

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
08_06_2015	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6.	mela1_0806; mela2_0806; mela3_0806; mela4_0806; mela5_ 0806; mela6_0806; cap1_0806;		
10_06_2015	installation of 3 Popillia japonica traps	Tfly3; Tfly4; Tfly5		
15_06_2015	check spore caps and apple baits mela1, mela2, mela3, mela4, mela5 and mela6. check Popillia traps TFly2. installation of 3 Popillia traps. Sampling of 4 plant material samples. installation of 3 pagodas traps for Thaumatotibia and 1 pagodas trap for Tecia. check rhynchophorus buckets trap SRh2. check and replacemen 4 chromotropic traps for Thrips	Cap1_1506;mela1_1506; mela2_1506; mela3_1506; mela4_1506; mela5_ 1506; mela6_1506; Tfly2; Tfly6, Tfly7, Tfly8; Var5_1506, Var6_1506, Var7_1506, Var8_1506; PgTh1, PgTh2, PgTh3; PgTe1; SRh2; CromoGTh5_1506, CromoGTh6_1506, CromoBTh3_1506, CromoBTh4_1506.	Var5_1506: Quinoa leaves with con foliage mine	*Pegomya* sp.
			Var6_1506:Quinoa leaves with caterpilla parasitized	Presence of *Autographa gamma,* parasitized by *Euplectrus* sp.
			Var7_1506: Juglans, Iran pavillion, visual analysis and stereo-microscopic observation	Presence of erythrophic mite *Aceria erineus* (Nal.). Presence of *Marssonina juglandis* (lib.) Magn. spores
			Var8_1506: Juglans fruits	Absence of insects
			CromoGTh5_1506: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis, Aeolothrips intermedius*
			CromoGTh6_1506: visual analysis and stereo-microscopic observation	Presence of Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis, Aeolothrips intermedius
			CromoBTh3_1506: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis, Aeolothrips intermedius*
			CromoBTh4_1506: visual analysis and stereo-microscopic observation	Presence of Frankliniella occidentalis, Frankliniella intonsa, Frankliniella tenuicornis, Aeolothrips intermedius
22_06_2015	check spore caps and sampling apple baits mela1, mela2, mela3, mela4, mela5 and mela6. check 5 rebell traps, installation of 2 chromotropic traps, replacemen of 9 chromotropic traps, check 2 planotrap for planococcus, sampling of 1 Citrus sample for Citrus tristeza virus, replacemen of 3 chromotropic traps for thrips, check 3 pagodas traps for Spodoptera, check Tfly1 for Popillia and reinstallation of 1 chromotropic traps. Generic sampling of 2 aphids on wisteria and 1 mole samples.	Cap1_2206; mela1_2206; mela2_2206; mela3_2206; mela4_2206; mela5_ 2206; mela6_2206; ReATf1, ReATf2, ReATf3, ReATf4, ReB1; CromoG16, CromoB10; CromoB3_2206, CromoB4_2206, CromoB8_2206, CromoB13_2206, CromoB14_2206, CromoB15_2206, CromoG7_2206, CromoG15_2206, CromoG17_2206, PlPf1, PlPf2, 4373962, CromoBTh1_2206, CromoBTh2_2206, CromoGTh2_2206, PgSp1, PgSp2, PgSp3, Tfly1, CromoB9; Var9_2206, Var10_2206	Cap1_2206: Microscope observations	Presence of *Alternaria* sp.,*Epicoccum* sp., *Cladosporium* sp., *Spaheropsis* sp.
			mela1_2206:Search *Phytophthora* sp.	Absent
			mela2_2206:Search *Phytophthora* sp.	Absent
			mela3_2206:Search *Phytophthora* sp.	Presence of *Colletotrichum acutatum*
			mela4_2206:Search *Phytophthora* sp.	Presence of *Colletotrichum acutatum*
			mela5_2206:Search *Phytophthora* sp.	Presence of *Colletotrichum fiorinae*
			mela6_2206:Search *Phytophthora* sp.	Absent
			CromoB3_2206:visual analysis and stereo-microscopic observation	Absence of Thrips
22_06_2015	check spore caps and sampling apple baits mela1, mela2, mela3, mela4, mela5 and mela6. check 5 rebell traps, installation of 2 chromotropic traps, replacemen of 9 chromotropic traps, check 2 planotrap for planococcus, sampling of 1 Citrus sample for Citrus tristeza virus, replacemen of 3 chromotropic traps for thrips, check 3 pagodas traps for Spodoptera, check Tfly1 for Popillia and reinstallation of 1 chromotropic traps. Generic sampling of 2 aphids on wisteria and 1 mole samples.	Cap1_2206; mela1_2206; mela2_2206; mela3_2206; mela4_2206; mela5_ 2206; mela6_2206; ReATf1, ReATf2, ReATf3, ReATf4, ReB1; CromoG16, CromoB10; CromoB3_2206, CromoB4_2206, CromoB8_2206, CromoB13_2206, CromoB14_2206, CromoB15_2206, CromoG7_2206, CromoG15_2206, CromoG17_2206, PlPf1, PlPf2, 4373962, CromoBTh1_2206, CromoBTh2_2206, CromoGTh2_2206, PgSp1, PgSp2, PgSp3, Tfly1, CromoB9; Var9_2206, Var10_2206	CromoB4_2206:visual analysis and stereo-microscopic observation	Absence of Thrips
			CromoB8_2206:visual analysis and stereo-microscopic observation	Presence of less than 10 Thrips of *Thrips tabaci* species
			CromoB13_2206:visual analysis and stereo-microscopic observation	Presence of a dozen of Thrips, of *Anaphothrips obscurus* species
			CromoB14_2206:visual analysis and stereo-microscopic observation	Presence of a dozen of Thrips
			CromoB15_2206:visual analysis and stereo-microscopic observation	Presence of a dozen of Thrips
			CromoG7_2206:visual analysis and stereo-microscopic observation	Absence of Thrips and Diptera Tephritidae
			CromoG15_2206:visual analysis and stereo-microscopic observation	Absence of Thrips and Diptera Tephritidae
			CromoG17_2206:visual analysis and stereo-microscopic observation	Absence of Thrips and Diptera Tephritidae
			CromoBTh1_2206:visual analysis and stereo-microscopic observation	Presence of hundreds of Thrips of *Thrips tabaci* species
			CromoBTh2_2206:visual analysis and stereo-microscopic observation	Presence of fifty Thripsof *Thrips flavus* species
			CromoGTh2_2206:visual analysis and stereo-microscopic observation	Presence of hundreds of Thrips of *Thrips tabaci* species
			Sample for Citrus tristezza virus:4373962	Absent
			Var9_2206: glycine sample for aphid control	Presanza of A*phis craccivora*
			Var10_2206:moths on glycine	Presence of *Autographa gamma*

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
24_06_2015	installation of of 2 multi-funnel for anoplofora, check 2 trap popillia Tfly, check and replacemen of 3 pagode for spodoptera, installation of pagoda for 2 taumatotibia, installation of 1 pagoda for Tecia, installation of of 1 pagoda for keiferia, check and replacemen of 1 trap cromotropica.	MfAn7, MfAn8, TFly9, TFly10, PgSp1, PgTh4, PgSp2, PgTe2, PgSp3, PgKe1, CromoG10, PgTh5	CromoG10: check fruit flies	Negativo
			PgSp1: check Spodoptera	Negativo
			PgSp2: check Spodoptera	Negativo
			PgSp3: check Spodoptera	Negativo
25_06_2015	installation of 1 multi-funnel trap for monochamus spp., installation of 8 prism traps for Agrilus spp.	MfMo1, PgAg3, PgAg4, PgAg5, PgAg5, PgAg6, PgAg7, PgAg8, PgAg9, PgAg10		
29_06_2015	Sampling and replacemen of spore caps. installation of 2 multi-funnel traps for Anoplophora spp., 1 multi-funnel trap for Monochamus spp. and installation of 4 chromotropic traps for Anthonomus spp.	Cap1_2906; MfMo2, MfAn9, MfAn10, CromoGAt1, CromoGAt2, CromoGAt3, CromoGAt4	Cap1_2906: Microscope observations	Presence of *Alternata alternata, Epicoccum* sp., *Cladosporium* sp. and Oospore of *Plasmopara* sp.
07_07_2015	check spore caps	cap1_0607	Cap1_0707: Microscope observations	Presence of *Alternaria alternata* spores, *Epicoccum* sp., *Cladosporium* sp., Oospore of *Plasmopara* sp.
09_07_2015	check 10 traps for popillia with sampling of 4 samples, shifted Popillia trap Tfly3 in Austria pavillion, check and sampling of 1 planotrap for Planococcus	Tfly1, Tfly2, Tfly3, Tfly4, Tfly5_0907, Tfly6, Tfly7_0907, Tfly8_0907, Tfly9, Tfly10_0907; PlPc1_0907	Tfly5_0907: check popillia visual analysis and stereo-microscopic observation	Negativo
			Tfly7_0907: check popillia visual analysis and stereo-microscopic observation	Negativo, Presence of *Diabrotica v. virgifera*
			Tfly8_0907: check popillia visual analysis and stereo-microscopic observation	*Formica* sp. (prob. 98,4%)
			Tfly10_0907: check popillia visual analysis and stereo-microscopic observation	Negativo, Presence of *Rhaphigaster* sp.
			PlPc1_0907: check planococcus	NEGATIVE (no insect)
10_07_15	Sampling of 6 olive samples for Xylella fastidiosa; check apple bait Mela1, Mela2, Mela3, Mela6; check and replacemen traps 4 Thrips CromoBTh and CromoGTh; Installation of 4 white rebell traps for sawflies, sampling of generic sample of thrips Var11	Mela1_1007, mela2_1007, mela3_1007, mela6_1007; campione 3020376, 3020377, 3020378, 3020379, 3020380, 3020381; CromoBTh3_1007, CromoBTh4, CromoGTh5, CromoGTh6; ReB2, ReB3, ReB4, ReB5; Var11_1007	Sample for Xylella: 3020376	Absent
			Sample for Xylella: 3020377	Absent
			Sample for Xylella: 3020378	Absent
			Sample for Xylella: 3020379	Absent
			Sample for Xylella: 3020380	Absent
			Sample for Xylella: 3020381	Absent
			CromoBTh3_1007: visual analysis and stereo-microscopic observation	Presence of hundreds of Thrips of *Thrips tabaci* species
			CromoBTh4_1007: visual analysis and stereo-microscopic observation	Presence of hundreds of Thrips of *Frankliniella occidentalis* species
			CromoGTh5_1007: visual analysis and stereo-microscopic observation	Absence of Thrips and Diptera Tephritidae
			CromoGTh6_1007: visual analysis and stereo-microscopic observation	Presence of hundreds of Thrips of *Thrips tabaci* species
			Var11_1007: check secco Thrips	NEGATIVE (no insect)
13_07_15	installation of 1 pagodas traps for tecia; installation of of 3 rebell Amarillo traps for tephritidae and 1 rebell Amarillo for Rhagoletis; installation of 4 yellow cromotropic traps	PgTe3, ReARh1, ReATf5, ReATf6, ReATf7, CromoG11,CromoG18, CromoG20, CromoG10		
14_07_15	check spore caps; installation of 1 yellow chromotropic trap and 2 yellow chromotropic traps for anthomonus, sampling of 7 olive samples for Xylella fastidiosa	Cap1_1407, CromoG19, CromoGAt5, CromoGAt6, 4373915, 4373916, 4373917, 4373918, 4373919, 4373920, 4373921	Cap1_1407: Microscope observations	Presence of *Alternaria alternata* spores, *Epicoccum* sp., *Cladosporium* sp.
			Sample for Xylella: 4373915	Absent
			Sample for Xylella: 4373916	Absent
			Sample for Xylella: 4373917	Absent
			Sample for Xylella: 4373918	Absent
			Sample for Xylella: 4373919	Absent
			Sample for Xylella: 4373920	Absent
			Sample for Xylella: 4373921	Absent

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
15_07_15	installation of 7 chromotropic thrips traps; installation of 2 pagodas traps for Keiferia, repositioning of 1 white rebell trap; check of 2 popillia traps	CromoGTh7, CromoBTh5, CromoBTh6, ReB5, CromoBTh7, CromoGTh8, PgKe2, CromoBTh8, PgKe3, CromoGTh9. TFly5 and Tfly2.		
16_07_15	installation of 3 chromotropic traps for Thrips; installation of 2 chromotropic traps for Anthonomus. Check of 2 Agrilus traps.	CromoGTh10, CromoBTh9, CromoBTh10, CromoGAt7, CromoGAt8, PgAg9, PgAg4.		
17_07_15	Replacement of 4 chromotropic traps.	CromoG1_1707, CromoG3_1707, CromoG4_1707, CromoG5_1707	CromoG1_1707: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoG3_1707: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoG4_1707: visual analysis and stereo-microscopic observation	*Frankliniella intonsa*
			CromoG5_1707: visual analysis and stereo-microscopic observation	/
20_07_15	Sampling and replacemen of spore caps. installation of 2 white rebel traps for tephritidae, replacemen 6 simple chromotropic traps and 1 chromotropic trap for thrips, reinstallation of 4 chromotropic traps, check 2 traps for Rhynchophorus and 3 for Popillia.	Cap1_2007; ReB7, ReB8, CromoB1_2007, CromoB10_2007, CromoG2_2007, CromoG7_2007, CromoG10_2007, CromoG16_2007, CromoGTh4_2007, CromoB12, CromoG8, CromoB11, CromoB2, SRh1, SRh4, TFly1, TFlly4, TFly8.	Cap1_2007: Microscope observations	Presence of *Trichothecium roseum, Alternaria alternata, Cladosporium* sp., *Epicoccum* sp., *Plasmopara* sp. oospore
			CromoB1_2007: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoB10_2007: visual analysis and stereo-microscopic observation	/
			CromoG2_2007: visual analysis and stereo-microscopic observation	*Thrips tabaci, Aeolothrips intermedius*
			CromoG7_2007: visual analysis and stereo-microscopic observation	*Anaphothrips obscurus*
			CromoG10_2007: visual analysis and stereo-microscopic observation	*Frankliniella intonsa, Aeolothrips intermedius, Haplothrips* sp.
			CromoG16_2007: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoGTh4_2007: visual analysis and stereo-microscopic observation	Absence Thrips and Diptera Tephritidae
22_07_15	Replacement of 8 chromotropic simple traps and 4 chromotropic traps for Thrips, reinstallation of 4 chromotropic traps, check 1 trap for Rhynchophorus, 3 pagodas traps for Thaumatotibia, 1 pagodas traps for Spodoptera, 2 for Tecia and 2 amarillo rebel trapsfor tephritidae.	CromoB3_2207, CromoB4_2207, CromoB7_2207, CromoB16_2207, CromoG6_2207, CromoG12_2207, CromoG13_2207, CromoG14_2207, CromoBth3_2207, CromoGTh1_2207, CromoGTh3_2207, CromoGTh5_2207; CromoB6, CromoB5, CromoG17, CromoB9; SRh3; PgTh1, PGTh2, PgTh3; PgSp3; PgTe1, PgTe2; ReATf3, ReATf6.	SRh3_2207/ check for *Rhynchophorus ferrugineus* visual analysis and stereo-microscopic observation	Absence of *Rhynchophorus ferrugineus.* Presence of *Oryctes nasicornis*
			CromoB3_2207: visual analysis and stereo-microscopic observation	*Thrips tabaci*
			CromoB4_2207: visual analysis and stereo-microscopic observation	*Thrips tabaci*
			CromoB7_2207: visual analysis and stereo-microscopic observation	*Thrips tabaci*
			CromoB16_2207: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoG6_2207: visual analysis and stereo-microscopic observation	*Frankliniella intonsa*
			CromoG12_2207: visual analysis and stereo-microscopic observation	*Haplothrips aculeatus*
			CromoG13_2207: visual analysis and stereo-microscopic observation	*Frankliniella intonsa*
			CromoG14_2207: visual analysis and stereo-microscopic observation	*Thrips tabaci*
			CromoBTh3_2207: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis, Lygus sp.*
			CromoGTh1_2207: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoGTh3_2207: visual analysis and stereo-microscopic observation	*no Thrips, Diabrotica virgifera virgifera*
			CromoGTh5_2207: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
23_07_15	Replacement of 5 chromotropic simple traps and 5 chromotropic traps for Thrips, check 1 pagodas trap for Thaumatotibia, 1 pagodas trap for Spodoptera and 6 Popillia traps with sampling of 2 samples.	CromoB8_2307, CromoB13_2307, CromoB14_2307, CromoB15_2307, CromoG15_2307, CromoGTh2_2307, CromoGTh6_2307, CromoBTh1_2307, CromoBTh2_2307, CromoBTh4_2307, PgTh4, PgSp2, TFly1, TFly5_2307, TFly8, TFly9, TFly6_2307, TFly7.	TFly5_2307/check popillia visual analysis and stereo-microscopic observation	Absence of *Popillia japonica* , Presence of 1 *Cetonia aurata* and 1 *Corythucha ciliata* and 2 H*alyomorpha Halys*
			TFly6_2307/check popillia visual analysis and stereo-microscopic observation	Absence of *Popillia japonica* , Presence of 1 *Lucilia sericata* and 1 *Rhaphigaster nebulosa*
			CromoB8_2307: visual analysis and stereo-microscopic observation	Presence of Frankliniella occidentalis, Tenothrips frici and Aeolothrips intermedius
			CromoB13_2307: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoB14_2307: visual analysis and stereo-microscopic observation	*Presence of Haplothrips aculeatus*
			CromoB15_2307: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis* and *Aeolothrips intermedius*
			CromoG15_2307: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoBTh1_2307:visual analysis and stereo-microscopic observation	*Ophraella communa, Pieris* sp., *Frankliniella intonsa, Aeolothrips intermedius, Frankliniella occidentalis*
			CromoBTh2_2307:visual analysis and stereo-microscopic observation	*Thrips tabaci, Frankliniella intonsa*
			CromoBTh4_2307:visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoGTh2_2307:visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoGTh6_2307:visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis* and *Gypsonoma aceriana*
27_07_15	Reinstallation of 6 apple baits, sampling and replacemen spore caps, repositioning of multi-funnel trap for Anoplophora, sampling of thrips material, check 5 TFly traps for Popillia	Cap1_2707, MfAn1, Var12_2707, TFly1, TFly2, TFly3, TFly4, TFly10	Cap1_2707: Microscope observations	Presence of *Cladosporium* sp. spores, *Plasmopara* sp., *Alternata alternata, Epicoccum* sp.
			Var12_2707 check Thrips biomolecular analysis	Presence of Thrips, of *Frankliniella occidentalis*
28_07_15	Reinstallation of buckets trap for Rhynchophorus , check and cleaning of 1 rebell amarillo traps, check of 1 white rebell trap and 1 amarillo rebell trap, detection of 1 spy plant for Agrilus	SRh1, ReATf1_2807, PS1, ReB6, ReATf3	ReATf1_2807: check Tephritidae	*Scaphoideus titanus, Scymnus sp., Stethorus sp.*
29_07_15	Reinstallation of 1 yellow chromotropic trap, change pheromons lure for 3 spodoptera pagodas traps, replacemen of 1 white rebell trap, sampling of 23 coffea plants, placement of 1 multi-funnel traps for monochamus, 2 yellow chromotropic traps and 2 blue chromotropic trap, entomological material sample for planococcus ficus on plano trap	CromoG5, PgPs1, PgPs2, PgPs3, PgTe1, ReB1_2907, 4373906, 4373905, 4373904, 4373903, 4373902, 4373963, 437394, 4094716, MfMo3, CromoG1amsa, CromoB1amsa, CromoG2amsa, CromoG1amsa, PlPf2_2907	Sample for Xylella: 4373902	Absent
			Sample for Xylella: 4373903	Absent
			Sample for Xylella: 4373904	Absent
			Sample for Xylella: 4373905	Absent
			Sample for Xylella: 4373906	Absent
			Sample for Xylella: 4373907	Absent
			Sample for Xylella: 4373963	Absent
			Sample for Xylella: 4373964	Absent
			Sample for Xylella: 4094716	Absent
			ReB1_2907: check sawflies	/
			PlPf2_2907: check per Planococcus	*Presence of Sigara striata*
30_07_15	check apple baits mela1, mela2, mela3, mela4, mela5 and mela6.	mela1_3007; mela2_3007; mela3_3007; mela4_3007; mela5_3007; mela6_3007	mela1_3007:Search *Phytophthora* sp.	Presence of *Colletotrichum florinae*
			mela2_3007:Search *Phytophthora* sp.	Absent
			mela3_3007:Search *Phytophthora* sp.	Absent
			mela4_3007:Search *Phytophthora* sp.	Presence of *Colletotrichum acutatum*
			mela5_3007:Search *Phytophthora* sp.	Presence of *Colletotrichum florinae*

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
03_08_15	Sampling and replacemen spore caps, change pheromone lures and replacemen of 9 thrips chromotropic traps, check 2 thrips traps, change pheromone lures and replacemen of 3 anthonomus traps	Cap1_0308, CromoGTh1_0308, CromoGTh2_0308, CromoGTh3_0308, CromoGTh4_0308, CromoGTh5_0308; CromoGTh6, CromoBTh1; CromoBTh2_0308, CromoBTh3_0308, CromoBTh4_0308, CromoGAt1_0308, CromoGAt2_0308, CromoGAt3_0308	Cap1_0308: Microscope observations	Presence of *Epicoccum* sp. spores, *Trichotecium roseum, Alternaria alternata.*
			CromoGTh1_0308: visual analysis and stereo-microscopic observation	Presence of hundreds of Thrips of *Frankliniella occidentalis* species. Presence of *Diabrotica v. virgifera*
			CromoGTh2_0308: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* species
			CromoGTh3_0308: visual analysis and stereo-microscopic observation	Presence of less than 10 Thrips of *Frankliniella intonsa* species
			CromoGTh4_0308: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips *Frankliniella occidentalis* species
			CromoGTh5_0308: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* species
			CromoBTh2_0308: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* species
03_08_15	Sampling and replacemen spore caps, change pheromone lures and replacemen of 9 thrips chromotropic traps, check 2 thrips traps, change pheromone lures and replacemen of 3 anthonomus traps	Cap1_0308, CromoGTh1_0308, CromoGTh2_0308, CromoGTh3_0308, CromoGTh4_0308, CromoGTh5_0308; CromoGTh6, CromoBTh1; CromoBTh2_0308, CromoBTh3_0308, CromoBTh4_0308, CromoGAt1_0308, CromoGAt2_0308, CromoGAt3_0308	CromoBTh3_0308: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* and *Frankliniella intonsa* species
			CromoBTh4_0308: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* species
			CromoGAt1_0308: check Anthonomus, visual analysis and stereo-microscopic observation	Absence of Anthonomus spp., Presence of *Nysius graminicola, Lygus pratensis* and *Duponchelia fovealis*
			CromoGAt2_0308: check Anthonomus, visual analysis and stereo-microscopic observation	Absence of Anthonomus spp., Presence of *Eurydema ventralis, Ophraella communa,* and *Lygus pratensis.*
			CromoGAt3_0308: check Anthonomus, visual analysis and stereo-microscopic observation	Absence of Anthonomus spp., Presence of Ophraella communa.
04_08_2015	check traps and change pheromone lures on 2 pagodas traps for Tecia, sampling of PgTe1 sample, check and replacement anthonomus trap, change pheromone lure on sesia trap, check and replacemen pheromone lures on 3 thaumatotibia traps, repositioning CromoG11 and yellow thrips chromotropic trap CromoGTh9 pheromone lure because disappeared;	PgTe1_0408, PgTe2, CromoGAt4_0408, trap sesia, PgTh1, PgTh2, Pgth3, CromoG11, CromoGTh9	CromoGAt4_0408: check Anthonomus, visual analysis and stereo-microscopic observation	Absence of Anthonomus spp., Presence of *Diabrotica v. virgifera, Ophraella communa, Lygus pratensis* ,1 *Stictocephala bisonia* and 1 of *Altica oleracea*
			Var13_0408: check *Tecia* spp., breeding and identification	Absence of *Tecia* spp., Presence of 1 *Heliothis peltigera* and *Monopis imella*
05_08_2015	installation of 3 yellow chromotropic traps, check and sampling of 10 Popillia Tfly traps, check 1 amarillo rebell trap for Rhagoletis, check and sampling of 2 amarillo rebell traps, check 1 yellow chromotropic trap for Anthonomus (disappeared), check blue chromotropic trap for Thrips	CromoG21, CromoG22, CromoG23, TFly1, TFly2, TFly3, TFly4, TFly5, TFly6, TFly7, TFly8, TFly9, TFly10, ReARh1, CromoBTh6, CromoAt6, ReATf7, ReATf4, Var14_0508, Var15_0508, Var16_0508, Var17_0508	Var14_0508 (trap Tfly7) visual analysis and stereo-microscopic observation	*Popillia japonica* : Absent. Presence of 1 *Halyomorpha halys* 1 *Paracorsia repandalis* 1 *Adalia decempunctata*
			Var15_0508 (trap ReATf7) visual analysis and stereo-microscopic observation	Tephritidae: absents. Presence of 1 *Metcalfa pruinosa* 7 *Ophraella communa* 1 *Cicadella viridis* 1 *Spodoptera exigua*
			Var16_0508 (trap TFly9) visual analysis and stereo-microscopic observation	*Popillia japonica* : Absent. Presence of 1 *Harpalus rufipes*
			Var17_0508 (trap ReATf4) visual analysis and stereo-microscopic observation	Tephritidae: presents. Presence of 1 *Rhagoletis cerasi* , 1 E*uscelidius variegatus* , 1 *Philaenus spumarius* and 1 *Adalia bipunctata*
06_08_2015	sampling of 8 olive samples from Biomediterranean cluster for Xylella fastidiosa	4096420, 4096421, 4096422, 4096423, 4096424, 4096425, 4096426, 4096427, 4096428	Sample for Xylella: 4096420	Absent
			Sample for Xylella: 4096421	Absent
			Sample for Xylella: 4096422	Absent
			Sample for Xylella: 4096423	Absent
			Sample for Xylella: 4096424	Absent
			Sample for Xylella: 4096425	Absent
			Sample for Xylella: 4096426	Absent
			Sample for Xylella: 4096427	Absent
			Sample for Xylella: 4096428	Absent

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
10_08_2015	Sampling and replacemen spore caps; replacemen of sesia trap	Cap1_1008, Sesia	Cap1_1008: Microscope observations	Presence of *Alternata alternata* spores, *Cercospora* sp., *Plasmopara* sp., *Cladosporium* sp., *Epicoccum* sp.
11_08_2015	check of 2 prism traps for Agrilus	PrAg3_1108, PrAg8	PgAg3_1108: Breeding, visual analysis and stereo-microscopic observation	*Agrilus* spp. Absent; Presence of 1 *Bruchus brachialis* and 1 *Metcalfa pruinosa*
12_08_2015	Check of 5 yellow chromotropic traps for Anthonomus and 1 replacement, check of 3 chromotropic traps for Thrips, 1 yellow chromotropic trap for fruit flies, check and replacement of 1 pagoda trap for tecia. Check of 1 multifunell trap for anoplophora, 1 white rebell trap, check and replacement of 1 pagoda trap for Keiferia.	CromoGAt1, CromoGAt5, CromoAt6, CromoAt7_1208, CromoAt8; CromoBTh4, CromoGTh6, CromoBTh8, CromoG20, PgTe3_1208, MfAn9, ReB5, PgKe1_1208	PgKe1_1208: Breeding, visual analysis and stereo-microscopic observation	*Keiferia* spp.: Absent. Presence of 7 *Tuta Absoluta*
17_08_2018	Sampling and replacemen spore caps. Check 4 buckets traps for Rhynchophorus, replacement of 4 blue chromotropic traps, 4 yellow chromotropic traps and pheromone lures for Thrips, check 2 pagodas traps for Keiferia with replacemen of pheromon lures and 1 of trap.	Cap1_1708; SRh1, SRh2, SRh3, SRh4, CromoBTh5_1708, CromoBTh6_1708, CromoBTh8_1708, CromoBTh9_1708, CromoGTh7_1708, CromoGTh8_1708, CromoGTh9_1708, CromoGTh10_1708, PgKe1, PgKe2_1708, PgKe3	Cap1_1708: Microscope observations	Presence of *Alternaria alternata* spores, *Cladosporium* sp., *Epicoccum* sp.
			CromoBTh5_1708: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoBTh6_1708: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoBTh8_1708: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoBTh9_1708: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoGTh7_1708: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoGTh8_1708: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoGTh9_1708: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis*
			CromoGTh10_1708: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis*
			PgKe2: visual analysis and stereo-microscopic observation	*Ophraella communa*
18_08_2015	Replacement of 4 chromotropic traps for Anthonomus and pheromone lures. Replacemen of 8 yellow chromotropic traps and 4 blue chromotropic traps.	CromoGAt5_1808, CromoGAt6_1808, CromoGAt7_1808, CromoGAt8_1808; CromoG1_1808, CromoG2_1808, CromoG3_1808, CromoG4_1808, CromoG5_1808, CromoG12_1808, CromoG13_1808, CromoG14_1808; CromoB5_1808, CromoB6_1808, CromoB7_1808, CromoB16_1808.	CromoG1_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Protapion fulvipes* and *Dioxyna* sp. (prob. bidentis)
			CromoG2_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Zyginidia pullula* 1 *Laodelphax striatellus*
			CromoG3_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Metcalfa pruinosa* and *Tephritis formosa*
			CromoG4_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Halyomorpha halys* 1 *Heliothis peltigera* 1 *Thaumatomyia notata1*
			CromoG5_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 Zyginidia pullula
			CromoG12_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Harmonia axyridis* 2 *Ophraella communa* 1 *Laodelphax striatellus*
			CromoG13_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 2 Ophraella communa 2 Galerucella luteola
			CromoG14_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Ophraella communa* and *Aphis craccivora*

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
18_08_2015	Replacement of 4 chromotropic traps for Anthonomus and pheromone lures. Replacemen of 8 yellow chromotropic traps and 4 blue chromotropic traps.	CromoGAt5_1808, CromoGAt6_1808, CromoGAt7_1808, CromoGAt8_1808; CromoG1_1808, CromoG2_1808, CromoG3_1808, CromoG4_1808, CromoG5_1808, CromoG12_1808, CromoG13_1808, CromoG14_1808; CromoB5_1808, CromoB6_1808, CromoB7_1808, CromoB16_1808.	CromoB6_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Spodoptera exigua* 1 *Laodelphax striatellus* and *Coenosia attenuata*
			CromoB7_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 Aelia acuminata and 1 Laodelphax striatellus
			CromoB16_1808 Breeding, visual analysis and stereo-microscopic observation	Presence: 1 *Autographa gamma* 1 *Laodelphax striatellus* 1 S*podoptera exigua* and *Deraeocoris nebulosus*
			CromoGAt7_1808 Breeding, visual analysis and stereo-microscopic observation	*Anthonomus* spp.: Absent. Presence: 3 *Cicadella viridis* 1 *Nysius senecionis*
			CromoGAt8_1808 Breeding, visual analysis and stereo-microscopic observation	*Anthonomus* spp.: Absent. Presence: 1 *Corythucha ciliata* 3 *Orius majusculus*
20_08_2015				
25_08_2015	Replacement of 6 apple baits, 11 chromotropic traps, and reinstallation of 2 chromotropic traps CromoB3 and CromoG11, only check for CromoB4 and CromoB2, check and replacemen spore caps, check Tfly trap for Popillia.	Mela1_2508; Mela2_2508; Mela3_2508; Mela4_2508; Mela5_2508; Mela6_2508;CromoG6_2508, CromoB12_2508; CromoG9_2508; CromoB11_2508; CromoG8_2508; CromoG17_2508; CromoB10_2508; CromoG20_2508; CromoB13_2508; CromoB9_2508; CromoB3, CromoG11; CromoB2, CromoB1; CromoB4;TFly2 , Cap1_2508	Cap1_2508: Microscope observations	Presence of *Alternaria* sp. spores
			CromoG6_2508: visual analysis and stereo-microscopic observation	Absence of Diptera Tephritidae, Presence of *Lygus pratensis, Harmonia axyridis* and *Ophraella communa* .
			CromoG8_2508: visual analysis and stereo-microscopic observation	Presence of *Ophraella communa*
			CromoG9_2508:visual analysis and stereo-microscopic observation	Presence of *Lucilia sericata*
			CromoG16_2508: visual analysis and stereo-microscopic observation	Absence of Diptera Tephritidae
			CromoG17_2508:visual analysis and stereo-microscopic observation	Absence of Diptera Tephritidae
			CromoG20_2508: visual analysis and stereo-microscopic observation	Presence of *Ophraella communa* , of *Fieberiella florii, Scymnus* sp., and *Stethorus* sp.
			CromoB9_2508: visual analysis and stereo-microscopic observation	Presence of Metcalfa pruinosa
			CromoB10_2508: visual analysis and stereo-microscopic observation	Presence of *Coenosia attenuata, Sepedon sphegea* and *Spodoptera exigua*
			CromoB11_2508: visual analysis and stereo-microscopic observation	Presence of *Coenosia attenuata* and *Ophraella communa*
			CromoB12_2508: analisi visiva and osservazione stereo-microscopio	Presence of *Chorosoma schillingi*
			CromoB13_2508:visual analysis and stereo-microscopic observation	Presence of *Chorosoma schillingi*
26_08_2015	New installation of pagoda trap for Keiferia; check 9 multi-funnel traps for Anoplophora; check 1 multi-funnel trap for Monochamus, 6 TFly traps for Popillia; Finding 3 spy plants for Anoplophora, 2 spy plants for Agrilus; 1 spy plant for Erwinia.	PgKe3, MfAn1, MfAn2, MfAn4,MfAn10, TFly1, PsAg2, PsAn1, MfAn6; MfMo2; TFly4; MfAn9; TFly7; PsAg3, MfAn7, MfAn8; PsAn3, MfAn5; TFly5; TFly10_2608; PsEr1; TFly9.	Tfly10_2608 visual analysis and stereo-microscopic observation	*Popillia japonica* : Absent. Presence: 1 *Cetonia aurata*
27_08_2015	Replacement of 7 chromotropic traps, new installation of 1 chromotropic trap, finding 1 spy plant for Citrus tristezza virus.	CromoB8_2708, CromoG15_2708, CromoB15_2708, CromoB14_2708, CromoG19_2708, CromoG10_2708, CromoG7_2708; CromoG18; PsCi2	CromoB8_2708: visual analysis and stereo-microscopic observation	*Hydroptila vectis*
			CromoB14_2708: visual analysis and stereo-microscopic observation	NEGATIVE (no result)
			CromoB15_2708: visual analysis and stereo-microscopic observation	*Ophraella communa*
			CromoG7_2708: visual analysis and stereo-microscopic observation	*Coenosia attenuata, Ophraella communa*
			CromoG10_2708: visual analysis and stereo-microscopic observation	Presence of *Eutettix* sp. or *Allygus atomarius*
			CromoG15_2708: visual analysis and stereo-microscopic observation	Presence of *Euleia* sp., and *Acanthiophilus helianthi*
			CromoG19_2708: visual analysis and stereo-microscopic observation	*Ophraella communa*
28_08_2015	check apple baits mela1, mela2, mela3, mela4, mela5 and mela6.	mela1_2808; mela2_2808; mela3_2808; mela4_2808; mela5_2808; mela6_2808	mela1_2808:Search *Phytophthora* sp.	Presence of *Colletotrichum acutatum*
			mela2_2808:Search *Phytophthora* sp.	Presence of *Colletotrichum acutatum*
			mela3_2808:Search *Phytophthora* sp.	Presence of *Colletotrichum fiorinae*
			mela4_2808:Search *Phytophthora* sp.	Absent
			mela5_2808:Search *Phytophthora* sp.	Presence of *Colletotrichum fiorinae*
			mela6_2808:Search *Phytophthora* sp.	Absent

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
31_08_2015	Sampling and replacemen spore caps, uninstall 6 white rebel traps and 1 amarillo rebel traps for Rhagoletis	Cap1_3108; ReB2_3108, ReB4_3108, ReB5_3108, ReB6_3108, ReB7_3108, ReB8_3108; ReARh1_3108	ReB2_3108:visual analysis and stereo-microscopic observation	Presence of *Hylaeus leptocephalus, Aproaerema anthyllidella, Ophraella communa*
			Cap1_3108: Microscope observations	Presence of *Alternaria* sp spores, *Epicoccum* sp., *Plasmopara* sp., *Cladosporium* sp.
			ReB3_3108:visual analysis and stereo-microscopic observation	NEGATIVE (no insect)
			ReB4_3108:visual analysis and stereo-microscopic observation	Presence of *Eristalinus taeniops , Lygus pratensis, Ophraella communa, Oplodontha viridula*
			ReB5_3108:visual analysis and stereo-microscopic observation	Presence of *Mystacides azureus*
			ReB6_3108:visual analysis and stereo-microscopic observation	*Eristalinus* sp., *Ophraella communa*
			ReB7_3108:visual analysis and stereo-microscopic observation	Presence of *Athalia rosae rosae, Ophraella communa, Halyomorpha halys*
			ReB8_3108:visual analysis and stereo-microscopic observation	Presence of *Cryptochironomus supplicans, Mystacides azureus, Nomophila noctuella, Athalia rosae rosae, Ophraella communa, Cicadella viridis, Laodelphax striatellus*
			ReARh1_3108:visual analysis and stereo-microscopic observation	Presence of *Hydroptila vectis, Hydropsyche modesta, Adalia bipunctata, Fieberiella florii, Ophraella communa*
01_09_2015	check treatments of Qatar pavillion			
02_09_2015	uninstall 2 white rebel traps and 1 pagoda trap for Tecia, new installation of multi-funnel trap for Pityophthorus with pheromon lures replacemen, check and replacemen of pheromone lures for 6 prism traps for Agrilus with sampling of 3 traps.	ReB1_0209, ReB3_0209; PgTe1_0209; MfPi1; PrAg1_0209, PrAg3, PrAg4, PrAg5_0209; PrAg6, PrAg7_0209	ReB1_0209: check sawflies, visual analysis and stereo-microscopic observation	NEGATIVE (no insect)
			ReB3_0209: check sawflies, visual analysis and stereo-microscopic observation	NEGATIVE (no insect)
			PrAg1_0209: check *Agrilus* sp.	Absence of *Agrilus* sp., Presence of *Deraeocoris lutescens* , and *Scymnus auritus.*
			PrAg5_0209: check *Agrilus* sp.	Absence of Agrilus sp., Presence of Hydroptila angulata, Hylaeus sp.(dilatatus o angularis), Deraeocoris sp., Adalia bipunctata, Ophraella communa
			PrAg7_0209: check *Agrilus* sp.	*Orientus ishidae*
			PgTe1_0209: check *Tecia* sp.	NEGATIVE (no insect)
03_09_2015	uninstall of 2 pagodas traps for Tecia, check 3 multi-funnel traps for Anoplophora and replacemen 2 pheromone lures, check and replacemen pheromone lure of Monochamus trap with sampling, replacemen 4 pheromone lures of Agrilus prism traps with sampling. Finding 3 spy plants for Phythophthora and 1 for Bursaphelencus and Gibberella.	PgTe2_0309, PgTe3_0309; MfAn3, MfAn5, MfAn6, MfMo1_0309; PrAg2, PrAg8_0309, PrAg9, PrAg10_0309; PsPh1, PsPh2, PsPh3, PsBG1.	PgTe2_0309: check *Tecia* sp.	NEGATIVE (no insect)
			PgTe3_0309: check *Tecia* sp.	*Clepsis peritana, Scrobipalpa ocellatella*
			PrAg8_0309: check *Agrilus* sp.	Absence of *Agrilus* sp., Presence *Stethorus punctillum, Harmonia axyridis*
			PrAg10_0309: check *Agrilus* sp.	Absence of Agrilus sp., Presence of Ophraella communa, Orientus ishidae
			MfMo1_0309: check *Monochamus sp.*	*Apis mellifera, Polistes sp., Vespula sp., Ips sexdentatus*
04_09_2015	check and replacemen of 2 blue chromotropic traps, 1 Monochamus traps with replacement of pheromone lure, and sampling. Check 5 yellow chromotropic traps with replacemen of 4 traps. Finding 2 spy plants for Erwinia, 1 spy plant for Sharka and 1 for flavescence dorée of grapevine.	PsEr2, PsEr3, PsFl1, PsSh1, CromoG1amsa_0409, CromoB1amsa_0409, CromoG2amsa_0409, CromoB2amsa_0409, CromoG21, CromoG22, CromoG23_0409	CromoG1amsa_0409: visual analysis and stereo-microscopic observation	*Zyginidia pullula, Cicadella viridis*
			CromoG2amsa_0409: visual analysis and stereo-microscopic observation	*Typhaea stercorea, Zyginidia pullula, Cicadella viridis, Ophraella communa*
			CromoB1amsa_0409: visual analysis and stereo-microscopic observation	*Spodoptera exigua, Zyginidia pullula*
			CromoB2amsa_0409: visual analysis and stereo-microscopic observation	*Spodoptera exigua, Taylorilygus apicalis, Megalonotus sp. (sabulicola/chiragra), Zyginiadia pullula, Ophraella communa*
			CromoG23_0409: visual analysis and stereo-microscopic observation	*Xyleborinus saxeseni, Zyginidia pullula, Clogmia albipunctata*

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
07_09_2015	Sampling and replacemen spore caps. Check 2 buckets traps for Rhynchophorus, check 6 and replacemen 5 yellow chromotropic traps and 4 blue chromotropic traps for Thrips. Check 1 Popillia trap, 1 Planococcus trap and 1 Thaumathotibia trap.	Cap1_0709;SRh1, SRh2; CromoGTh1_0709, CromoGTh3_0709, CromoGTh4_0709, CromoGTh5_0709, CromoGTh10_0709; CromoBTh1_0709, CromoBTh5_0709, CromoBTh6_0709, CromoBTh8_0709; CromoGTh7; Tfly2; PlPc1; PgTh3	Cap1_0709: Microscope observations	Presence of *Alternaria* sp.*, Epicoccum* sp.
			CromoGTh1_0709: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips someone of *Microcephalothrips abdominalis* species. Presence of *Lygus pratensis* and *Ophraella communa* . *Chrysotus n* . sp. choricus grp.
			CromoGTh3_0709: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Haplothrips aculeatus* species. Presence of *Ophraella communa* .
			CromoGTh4_0709: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* and *Thrips tabaci* species
			CromoGTh5_0709: visual analysis and stereo-microscopic observation	Presence of decine of Thrips della specie Frankliniella occidentalis
			CromoGTh10_0709: visual analysis and stereo-microscopic observation	Presence ofdozen of Thrips of *Tenothrips frici* species. Presence of *Aphis craccivora*
			CromoBTh1_0709: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* species, *Nysius graminicola, Cacoecimorpha pronubana* and *Sitotroga cerealella*
			CromoBTh5_0709: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Thrips tabaci and Frankliniella occidentalis* species, *Nomophila noctuella* , and *Athalia rosae rosae*
			CromoBTh6_0709: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella intonsa* species. Presence of *Megalonotus sabulicola* , *Choreutis nemorana, Helicoverpa armigera, Ophraella communa, Fieberiella florii* and *Japananus hyalinus* .
			CromoBTh8_0709: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* species, *Xyleborinus saxeseni, Thaumatomya notata* and *Hydroptila* sp.
08_09_2015	Check 2 buckets traps for Rhynchophorus, check and replacemen 4 yellow chromotropic traps and 4 blue chromotropic traps for Thrips, check 3 Popillia traps, and 1 Planococcus trap, replacemen Monochamus traps pheromone lure, and sampling citrus leaves for cochineal.	SRh2, SRh3; CromoGTh2_0809, CromoGTh6_0809, CromoGTh8_0809, CromoGTh9_0809; CromoBTh2_0809, CromoBTh3_0809, CromoBTh4_0809, CromoBTh9_0809; Tfly1, Tfly4, Tfly8; PlPf2; MfMo2; Var18_0809	CromoGTh2_0809:visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis, Tuta absoluta, Anoecia* sp.
			CromoGTh6_0809:visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis, Tuta absoluta, Agraylea sexmaculata, Mystacides azureus*
			CromoGTh8_0809:visual analysis and stereo-microscopic observation	Presence of Frankliniella intonsa, Nomophila noctuella, Ophraella communa, Cicadella viridis, Hylaeus sp., Agraylea sexmaculata
			CromoGTh9_0809:visual analysis and stereo-microscopic observation	Presence of *Agraylea sexmaculata, Psychoda alternata*
			CromoBTh2_0809:visual analysis and stereo-microscopic observation	Presence of *Frankliniella intonsa, Semidalis* sp. (prob. aleyrodiformis)
			CromoBTh3_0809:visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoBTh4_0809:visual analysis and stereo-microscopic observation	Presence of *Tuta absoluta, Frankliniella occidentalis, Cicadella viridis*
			CromoBTh9_0809:visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis, Hydropsyche sp., Sepedon sphegea*
			Var18_0809: check *Planococcus* sp.	Presence of *Planococcus citri*
09_09_2015	check and replacemen 2 chromotropic traps for Thrips; check and sampling 4 rebell amarillo traps and 4 yellow chromotropic traps for Thrips; check 1 Rebell Amarillo trap; new installation of rebell Amarillo trap (with chromotropic trap); check 1 planotrap for Planococcus; check 4 chromotropic traps for anthonomus with sampling.	CromoBTh10_0909; CromoBTh7_0909; ReATf1_0909; ReATf5_0909; ReATf2_909; ReATf7_0909; ReATf6; ReATf3 (sostituita con una cromotropica); PlPf1; CromoGAt1_0909; CromoGAt6; CromoGAt8; CromoGAt3	CromoBTh7_0909:visual analysis and stereo-microscopic observation	Presence of *Haplothrips leucanthemi*
			CromoBTh10_0909:visual analysis and stereo-microscopic observation	Presence of *Frankliniella intonsa*
			ReATf1_0909: check Tephritidae	*Ophraella communa* , *Hylaeus* sp.
			ReATf2_0909: check Tephritidae	Presence of *Euleia* sp., *Scymnus* sp., *Stethorus* sp.

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
09_09_2015	check and replacemen 2 chromotropic traps for Thrips; check and sampling 4 rebell amarillo traps and 4 yellow chromotropic traps for Thrips; check 1 Rebell Amarillo trap; new installation of rebell Amarillo trap (with chromotropic trap); check 1 planotrap for Planococcus; check 4 chromotropic traps for anthonomus with sampling.	CromoBTh10_0909; CromoBTh7_0909; ReATf1_0909; ReATf5_0909; ReATf2_909; ReATf7_0909; ReATf6; ReATf3 (sostituita con una cromotropica); PlPf1; CromoGAt1_0909; CromoGAt6; CromoGAt8; CromoGAt3	ReATf5_0909: check Tephritidae	Presence of *Altica oleracea, Ophraella communa, Cicadella viridis*
			ReaTf7_0909: check Tephritidae	Presence of *Eristalinus taeniops, Ophraella communa*
			CromoGAt1_0909: check Anthonomus sp.	*Ophraella communa* , *Cicadella viridis*
11_09_2015	new installation of 2 traps for Anthonomus; check 2 rebell amarillo traps; check and sampling 1 chromotropic traps for Anthomonus	CromoGAt6; CromoGAt5; ReATf4; CromoAt2_1109	CromoGAt2_1109: check Anthonomus sp.	Absence of *Anthonomus* spp., Presence of *Orius majusculus, Nysius* sp., *Cicadella viridis, Ophraella communa, Hylaeus* sp.
14_09_2015	Sampling and replacement spore caps. Replacement 5 apple baits for Phytophthora, check and replacemen 2 traps for Anthonomus.	Cap1_1409; Mela1, Mela2, Mela3, Mela4, Mela5; CromoGAt4_1409, CromoGAt7_1409	Cap1_1409: Microscope observations	Presence of *Alternaria* sp. spores, *Epicoccum* sp., *Drechslera* sp., *Cladosporium* sp.
			CromoGAt4_1409:visual analysis and stereo-microscopic observation	Absence of Anthonomus spp., Presence of Ophraella communa, Halyomorpha halys
			CromoGAt7_1409:visual analysis and stereo-microscopic observation	Absence of *Anthonomus* spp., Presence of *Cicadella viridis, Nysius* sp., *Dictya* sp.
15_09_2015	check and replacemen 5 yellow chromotropic traps, check 1 Planococcus trap and 1 for Thaumathotibia. New installation of 1 apple bait for Phytophthora	CromoG1_1509, CromoG2_1509, CromoG3_1509, CromoG4_1509, CromoG5_1509; PlPci1; PgTh1; Mela6.	CromoG1_1509: visual analysis and stereo-microscopic observation	Presence of *Scymnus* sp., *Stethorus* sp., *Orius majusculus*
			CromoG2_1509: visual analysis and stereo-microscopic observation	Presence of *Dictya* sp.
			CromoG3_1509: visual analysis and stereo-microscopic observation	Presence of *Dictya* sp.
			CromoG4_1509: visual analysis and stereo-microscopic observation	Presence of *Plutella xylostella* and *Halyomorpha halys*
			CromoG5_1509: visual analysis and stereo-microscopic observation	Presence of *Ophraella communa* and *Dictya* sp.
17_09_2015	Replacement 4 blue and 3 yellow chromotropic traps, new installation of 1 yellow chromotropic trap for Thrips, check 6 apple baits for Phytophthora, 2 traps for Popillia, disinstallation of 3 pagodas and replacemen of 1 for Thaumatotibia, Sampling of Arbutus branches for Phytophthora, and Citrus leaves from spy plant for Citrus tristezza virus.	CromoB5_1709, CromoB6_1709, CromoB7_1709, CromoB16_1709; CromoG12_1709, CromoG13_1709, CromoG14_1709; CromoGTh7, Mela1_1709, Mela2_1709, Mela3_1709, Mela4_1709, Mela5_1709, Mela6_1709; Tfly2, Tfly3; PgTh1_1709, PgTh2_1709, PgTh4_1709; PgTh3_1709; 4373642; PsCi2_1709	CromoB5_1709: visual analysis and stereo-microscopic observation	NEGATIVE (no insect)
			CromoB6_1709: visual analysis and stereo-microscopic observation	Presence of *Cricotopus* sp., *Haplothrips* sp., and *Dictya* sp.
			CromoB7_1709: visual analysis and stereo-microscopic observation	*Laodelphax striatellus, Nysius sp.*
			CromoB16_1709: visual analysis and stereo-microscopic observation	*Dictya* sp.
			CromoG12_1709: visual analysis and stereo-microscopic observation	Presence of *Daraeocoris nebulosus, Hydroptila vectis, Cicadella viridis, Ophraella communa, Coenosia attenuata, Thaumatomyia notata, Laodelphax striatellus*
			CromoG13_1709: visual analysis and stereo-microscopic observation	Presence of Aphelinus sp.?, Dictya sp., Coenosia attenuata, Thaumatomyia notata
			CromoG14_1709: visual analysis and stereo-microscopic observation	Presence of *Spodoptera exigua, Stegobium paniceum, Frankliniella occidentalis, Thrips hawaiiensis, Dictya* sp.
			PgTh1_1709: check *Thaumatotibia* sp.	Presence of Grapholita funebrana, Grapholita janthinana
			PgTh2_1709: check *Thaumatotibia* sp.	Presence of *Pammene albuginana*
			PgTh3_1709: check *Thaumatotibia* sp.	*Monopis imella*
			PgTh4_1709: check *Thaumatotibia* sp.	Presence of *Tuta absoluta, Cadra cautella*
			PsCi2_1709: check Citrus tristezza virus	*Dialeurodes citri*
			Sample for *Phytophthora ramorum* :4373642	Presence of *Pestalotiopsis* sp.
			mela1_1709: Search *Phytophthora* sp.	Absent
			mela2_1709: Search *Phytophthora* sp.	Presence of *Pythium dissotocum.*
			mela3_1709: Search *Phytophthora* sp.	Presence of *Colletotrichum acutatum*
			mela4_1709: Search *Phytophthora* sp.	Absent
			mela5_1709: Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela6_1709: Search *Phytophthora* sp.	Absent

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
18_09_2015	Meeting with press.			
21_09_2015	Meeting with press, sampling and replacemen spore caps	Cap1_2109	Cap1_2109: Microscope observations	Presence of *Alternaria* sp. spores, *Cladosporium* sp., *Drechslera* sp., *Epicoccum* sp., *Pestalotiopsis* sp.
22_09_2015	Sampling of rosemary, myrtle, cherry and broom branches for Xylella fastidiosa, and 1 sample for Erwinia.	4094718, 4094719, 4094720, 4094721, 4094722, 4094723, 4094724, 4094725; 3020475	Sample for Xylella: 4094718	Absent
			Sample for Xylella: 4094719	Absent
			Sample for Xylella: 4094720	Absent
			Sample for Xylella: 4094721	Absent
			Sample for Xylella: 4094722	Absent
			Sample for Xylella: 4094723	Absent
			Sample for Xylella: 4094724	Absent
			Sample for Xylella: 4094725	Absent
			Sample for Erwinia: 3020475	Absent
24_09_2015	Replacement 6 yellow and 5 blue chromotropic traps for Thrips and their pheromone lures, change only pheromone lure for 2 blue and 2 yellow chromotropic traps for Thrips, new installation of 1yellow chromotropic trap, check 1 planotrap for Planococcus ficus, 1 trap for popilia and replacemen 1 trap for Thaumatotibia.	CromoGTh1_2409, CromoGTh2_2409, CromoGTh3_2409, CromoGTh5_2409, CromoGTh7_2409, CromoGTh10_2409; CromoBTh1_2409, CromoBTh3_2409, CromoBTh7_2409, CromoBTh8_2409, CromoBTh10_2409; CromoBTh2, CromoBTh4; CromoGTh4, CromoGTh6; CRomoG6; PlPf2; TFly5; PgTh5_2409	CromoGTh1_2409: visual analysis and stereo-microscopic observation	Presence about twenty Thrips of *Thrips tabaci* species
			CromoGTh2_2409: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Thrips tabaci* species
			CromoGTh3_2409: visual analysis and stereo-microscopic observation	NEGATIVE (no insect)
			CromoGTh5_2409: visual analysis and stereo-microscopic observation	Presence about twenty Thrips of *Frankliniella intonsa* and *Microcephalothrips abdominalis* species
			CromoGTh10_2409: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* and *Haplothrips aculeatus* species
24_09_2015	Replacement 6 yellow and 5 blue chromotropic traps for Thrips and their pheromone lures, change only pheromone lure for 2 blue and 2 yellow chromotropic traps for Thrips, new installation of 1yellow chromotropic trap, check 1 planotrap for Planococcus ficus, 1 trap for popilia and replacemen 1 trap for Thaumatotibia.	CromoGTh1_2409, CromoGTh2_2409, CromoGTh3_2409, CromoGTh5_2409, CromoGTh7_2409, CromoGTh10_2409; CromoBTh1_2409, CromoBTh3_2409, CromoBTh7_2409, CromoBTh8_2409, CromoBTh10_2409; CromoBTh2, CromoBTh4; CRomoGTh4, CromoGTh6; CRomoG6; PlPf2; TFly5; PgTh5_2409	CromoBTh1_2409: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* species
			CromoBTh3_2409: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella intonsa* and *Thrips tabaci* species
			CromoBTh7_2409: visual analysis and stereo-microscopic observation	Presence of una ventina of Thrips della specie Frankliniella occidentalis
			CromoBTh8_2409: visual analysis and stereo-microscopic observation	Presence of dozen of Thrips of *Frankliniella occidentalis* and *Thrips tabaci* species
			CromoBTh10_2409: visual analysis and stereo-microscopic observation	Presence of decine of Thrips della specie Frankliniella occidentalis
			PgTh5_2409: check *Thaumatotibia*	Absence of *Thaumatotibia leucotreta.* Presence of P*ammene albuginana*
25_09_2015	Meeting with Bosnian Workgroup			
28_09_2015	Sampling and replacement spore caps. Replacemen 4 blue and 2 yellow chromotropic traps for Thrips and their pheromone lures. Check 1 blue and 2 yellow chromotropic traps for Thrips, replacemen 1 yellow chromotropic trap. Sampling olive branches for Xylella fastidiosa and uninstall 1 trap for Anthonomus with sampling.	Cap1_2809; CromoBTh2_2809, CromoBTh5_2809, CromoBTh6_2809, CromoBTh9_2809; CromoGTh8_2809, CromoGTh9_2809; CromoGTh4, CromoGTh6; CromoBTh4; CromoG7_2809; 3020476, 3020477; CromoGAt2_2809	Cap1_2809: Microscope observations	Presence of *Alternaria* sp. spores, *Epicoccum* sp.,*Drechslera* sp.,*Diplodia* sp.,*Cladosporium* sp.
			CromoBTh2_2809: analisi visiva and osservazione stereo-microscopica	Presence of una decina of Thrips della specie Thrips physapus
			CromoBTh5_2809: analisi visiva and osservazione stereo-microscopica	Presence of hundreds of Thrips of *Thrips tabaci* species
			CromoBTh6_2809: analisi visiva and osservazione stereo-microscopica	Presence of hundreds of Thrips of *Aeolothrips intermedius, Fieberiella floriie* and *Japananus hyalinus*
28_09_2015	Sampling and replacement spore caps. Replacemen 4 blue and 2 yellow chromotropic traps for Thrips and their pheromone lures. Check 1 blue and 2 yellow chromotropic traps for Thrips, replacemen 1 yellow chromotropic trap. Sampling olive branches for Xylella fastidiosa and uninstall 1 trap for Anthonomus with sampling.	Cap1_2809; CromoBTh2_2809, CromoBTh5_2809, CromoBTh6_2809, CromoBTh9_2809; CromoGTh8_2809, CromoGTh9_2809; CromoGTh4, CromoGTh6; CromoBTh4; CromoG7_2809; 3020476, 3020477; CromoGAt2_2809	CromoBTh9_2809: analisi visiva and osservazione stereo-microscopica	Presence of dozen of Thrips of *Frankliniella occidentalis* species.
			CromoGTh8_2809: analisi visiva and osservazione stereo-microscopica	Presence of dozen of Thrips of *Frankliniella occidentalis* species. Presence of *Dictya* sp.
			CromoGTh9_2809: analisi visiva and osservazione stereo-microscopica	Absence of Thrips, Presence of esemplari of Psychodidae
			CromoG7_2809: analisi visiva and osservazione stereo-microscopica	Presence of *Dictya* sp., Trichoptera, Psychodidae, C*adra cautella*
			CromoGAt2_2809: analisi visiva and osservazione stereo-microscopica	Absence of *Anthonomus* spp., Presence of *Lygus* sp.
			Sample for Xylella: 3020476	Absent
			Sample for Xylella: 3020477	Absent

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
29_09_2015	Uninstall 3 multi-funnel traps for Anoplophora and sampling material of them, uninstall 1 multi-funnel traps for Monochamus, pagodas traps for Keiferia, with sampling of each of them, check 1 Popillia trap, uninstall 1 trap for Anthonomus with sampling, check and replacemen 4 blue and 6 yellow chromotropic traps, only check for 2 yellow and 1 blue chromotropic trap, check and replacemen 1 amarillo rebel trap for Tephritidae, new installation of 1 yellow and 1 blue chromotropic trap and 1 yellow chromotropic trap for Thrips.	MfAn1, MfAn2_2909, MfAn4; MfMo2; PgKe1_2909, PgKe2_29009, PgKe3_2909; TFly4; CromoGAt1_2909; CromoB2_2909, CromoB10_2909, CromoB11_2909, CromoB12_2909; CromoG6_2909, CromoG8_2909, CromoG9_2909, CromoG16_2909, CromoG17_2909, CromoG21_2909; CromoG22, CromoG23; CromoB13; ReATf6_2909; CromoG20; CromoB1; CromoGTh3	MfAn2_2909:breeding and identification	Absence of *Anoplophora* spp., Presence of *Halyomorpha halis* , *Vespa crabro* and *Chlorophorus varius varius*
			CromoB2_2909:visual analysis and stereo-microscopic observation	Presence of *Scymnus* sp., and *Stethorus* sp.
			CromoB10_2909:visual analysis and stereo-microscopic observation	Presence of *Lygus* sp. and Psychodidae
			CromoB11_2909: visual analysis and stereo-microscopic observation	Presence of *Cicadella viridis*
			CromoB12_2909: visual analysis and stereo-microscopic observation	Presence of *Ophraella communa*
			CromoG6_2909: visual analysis and stereo-microscopic observation	Absence of Diptera Tephritidae
			CromoG8_2909: visual analysis and stereo-microscopic observation	*Scymnus* sp., *Stethorus* sp., *Cicadella viridis, Thaumatomyia notata*
			CromoG9_2909: visual analysis and stereo-microscopic observation	Presence of *Scymnus* sp., *Stethorus* sp., and *Japananus hyalinus*
29_09_2015	Uninstall 3 multi-funnel traps for Anoplophora and sampling material of them, uninstall 1 multi-funnel traps for Monochamus, pagodas traps for Keiferia, with sampling of each of them, check 1 Popillia trap, uninstall 1 trap for Anthonomus with sampling, check and replacemen 4 blue and 6 yellow chromotropic traps, only check for 2 yellow and 1 blue chromotropic trap, check and replacemen 1 amarillo rebel trap for Tephritidae, new installation of 1 yellow and 1 blue chromotropic trap and 1 yellow chromotropic trap for Thrips.	MfAn1, MfAn2_2909, MfAn4; MfMo2; PgKe1_2909, PgKe2_29009, PgKe3_2909; TFly4; CromoGAt1_2909; CromoB2_2909, CromoB10_2909, CromoB11_2909, CromoB12_2909; CromoG6_2909, CromoG8_2909, CromoG9_2909, CromoG16_2909, CromoG17_2909, CromoG21_2909; CromoG22, CromoG23; CromoB13; ReATf6_2909; CromoG20; CromoB1; CromoGTh3	CromoG16_2909: visual analysis and stereo-microscopic observation	Presence of *Zygina rhamni, Dictya* sp., *Scymnus* sp., and *Stethorus* sp.
			CromoG17_2909: visual analysis and stereo-microscopic observation	Presence of *Laodelphax striatellus* , *Dictya* sp., and Psychodidae
			CromoG21_2909: visual analysis and stereo-microscopic observation	Presence of *Kalotermes flavicollis* and Psychodidae
			PgKe1_2909: visual analysis and stereo-microscopic observation	Absence of *Keiferia* spp., Presence of *Tuta absoluta*
			PgKe2_2909: visual analysis and stereo-microscopic observation	Absence of *Keiferia* spp., Presence of Psychodidae
			PgKe3_2909: visual analysis and stereo-microscopic observation	Absence of *Keiferia* spp., Presence of *Ophraella communa* and *Propylaea 14-punctata*
			CromoGAt1_2909: visual analysis and stereo-microscopic observation	Absence of esemplari of Anthonomus spp., Presence of Galerucella luteola, of Cicadella viridis, of Corticarina sp., of Alloxysta sp., and Cryptophagus pilosus
			ReATf6_2909:visual analysis and stereo-microscopic observation	Absence of Diptera Tephritidae, Presence of *Ophraella communa, Athalia rosae rosae, Scymnu* s sp. and *Stethorus* sp.
02_10_2015	Replacement 6 apple baits for Phytophthora	Mela1; Mela2; Mela3; Mela4; Mela5; Mela6		
05_10_2015	Sampling and replacement spore caps, check apple baits mela1, mela2, mela3, mela4, mela5 and mela6, uninstall 4 multi-funnel traps for Anoplophora, sampling of 4 olive trees samples for Xylella fastidiosa.	Cap1_0510; Mela1_0510, Mela2_0510, Mela3_0510, Mela4_0510, Mela5_0510, Mela6_0510, MfAn7, MfAn8, MfAn9, MfAn10; 3020405, 3020406, 3020407, 3020408	Cap1_0510: Microscope observations	Presence of *Epicoccum* sp. spores
			mela1_0510: Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela2_0510: Search *Phytophthora* sp.	Absent
			mela3_0510: Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela4_0510: Search *Phytophthora* sp.	Absent
			mela5_0510: Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela6_0510: Search *Phytophthora* sp.	Absent
			Sample for Xylella: 3020405	Absent
			Sample for Xylella: 3020406	Absent
			Sample for Xylella: 3020407	Absent
			Sample for Xylella: 3020408	Absent

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
06_10_2015	Uninstalling 5 CromoG traps for Anthonomus and 3 pagodas traps for Spodoptera	CromoGAt3_0610, CromoGAt4_0610, CromoGAt5_0610, CromoGAt6_0610, CromoGAt8_0610; PgSp1_0610, PgSp2_0610, PgSp3_0610	CromoGAt3_0610: visual analysis and stereo-microscopic observation	Absence of *Anthonomus* spp., Presence of *Ophraella communa, Lyctus africanus*
			CromoGAt4_0610: visual analysis and stereo-microscopic observation	Absence of *Anthonomus* spp., Presence of *Ophraella communa, Cicadella viridis* and *Luperomorpha xanthodera*
			CromoGAt5_0610: visual analysis and stereo-microscopic observation	Absence of *Anthonomus* spp.
			CromoGAt6_0610: visual analysis and stereo-microscopic observation	Absence of *Anthonomus* spp.
			CromoGAt8_0610: visual analysis and stereo-microscopic observation	Absence of *Anthonomus* spp., Presence of *Halyomorpha halys, Sitotroga cerealella, Monopis imella*
			PgSp1_0610: visual analysis and stereo-microscopic observation	Absence of *Spodoptera* spp., Presence of *Metcalfa pruinosa*
			PgSp2_0610: visual analysis and stereo-microscopic observation	Absence of *Spodoptera* spp., Presence of *Ophraella communa* and *Hylaeus* sp.
			PgSp3_0610: visual analysis and stereo-microscopic observation	Absence of *Spodoptera* spp.
13_10_2015	Sampling and replacement spore caps.	Cap1_1310	Cap1_1310: Microscope observations	Presence of *Cladosporium* sp. spores, *Epicoccum* sp., *Alternaria* sp..
14_10_2015	Sampling 6 coffea trees samples and 2 mulch bark samples	3020409, 3020410, 3020482, 4096584, 4096586, 4096587; 3020483, 4096585	Sample for cochineal: 3020409	Presence of *Saissetia coffeae*
			Sample for cochineal: 3020410	Presence of *Pseudococcus viburni*
			Sample for cochineal: 3020482	Presence of *Pseudococcus longispinus*
			Sample for Xylella: 4096584	Absent
			Sample for Xylella: 4096586	Absent
			Sample for Xylella: 4096587	Absent
16_10_2015	Sampling 2 mulch bark samples.	3020484, 3020485	Samples for nematodes: 3020484	Absent
			Samples for nematodes: 3020485	Absent
19_10_2015	Sampling and replacement spore caps, 3 mulch bark samples and 4 soil samples for banana nematodes. Replacement 6 apple baits for Phytophthora and check 1 trap for Popillia	Cap1_1910; 3020486, 3020487, 3020488, 3020489, 3020490, 3020491, 3020492; Mela1, Mela2, Mela3, Mela4, Mela5, Mela6; Tfly5	Cap1_1910: Microscope observations	Presence of *Epicoccum* sp. spores and *Alternaria* sp.
			Sample for nematodes: 3020486	Absent
			Sample for nematodes: 3020487	Absent
			Sample for nematodes: 3020488	Absent
			Sample for nematodes: 3020489	Absent
			Sample for nematodes: 3020490	Absent
			Sample for nematodes: 3020491	Absent
			Sample for nematodes: 3020492	Absent
22_10_2015	Sampling 6 apple baits for Phytophthora	Mela1_2210, Mela2_2210, Mela3_2210, Mela4_2210, Mela5_2210, Mela6_2210	mela1_2210: Search *Phytophthora* sp.	Presence of *Colletotrichum acutatum*
			mela2_2210: Search *Phytophthora* sp.	Absent
			mela3_2210: Search *Phytophthora* sp.	Presence of *C. acutatum*
			mela4_2210: Search *Phytophthora* sp.	Presence of *C. acutatum*
			mela5_2210: Search *Phytophthora* sp.	Presence of *Pythium dissotocum*
			mela6_2210: Search *Phytophthora* sp.	Absent
23_10_2015	Sampling and uninstall 7 Rebel traps for Tephritidae, 2 traps for Thaumatotibia and 1 trap for Sesia	ReATf1_2310, ReATf2_2310, ReATf3_2310, ReATf4_2310, ReATf5_2310, ReATf6_2310, ReATf7_2310; PgTh3_2310, PgTh5_2310; Sesia_2310	ReATf5_2310: visual analysis and stereo-microscopic observation	*Ophraella communa, Coccinella decempunctata, Dictyophora europaea, Altica oleracea, Euleia sp., Trixagus meybohmi*
			ReATf1_2310: visual analysis and stereo-microscopic observation	*Ophraella communa, Metcalfa pruinosa, Scymnus sp.*
			ReATf2_2310: visual analysis and stereo-microscopic observation	*Typhlocyba sp., Hydroptila angulans, Scymnus sp.*

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
23_10_2015	Sampling and uninstall 7 Rebel traps for Tephritidae, 2 traps for Thaumatotibia and 1 trap for Sesia	ReATf1_2310, ReATf2_2310, ReATf3_2310, ReATf4_2310, ReATf5_2310, ReATf6_2310, ReATf7_2310; PgTh3_2310, PgTh5_2310; Sesia_2310	PgTh3_2310: visual analysis and stereo-microscopic observation	/
			ReATf3_2310: visual analysis and stereo-microscopic observation	*Chaetocnema tibialis, Zyginidia pullula, Scymnus sp., Tingidae*
			ReATf6_2310: visual analysis and stereo-microscopic observation	*Hishimonus hamatus, Typhlocyba sp., Psychodidae, Macrosteles sp., Ephemeroptera, Scymnus sp.*
			ReATf4_2310: visual analysis and stereo-microscopic observation	*Psammotettix sp. (prob. confinis), Sitona hispidulus, Trichoptera, Orientus ishidae, Scymnus sp., Stethorus sp., Anoecia sp., Thaumatomyia notata, Euleia sp.*
			PgTh5_2310: visual analysis and stereo-microscopic observation	*Hydroptila angulata, Corticarina sp.*
			Sesia_2310: visual analysis and stereo-microscopic observation	*Corticarina sp., Ophraella communa, Scymnus sp.*
26_10_2015	Sampling and replacement spore caps, uninstall 2 Popillia traps, sampling and uninstall 2 blue and 4 yellow chromotropic traps, disappeared 1 yellow chromotropic trap (CromoG23), sampling and uninstall 1 Planococcus trap	Cap1_2610; Tfly5, Tfly10; CromoB12_2610, CromoB13_2610; CromoG1_2610, CromoG7_2610, CromoG21_2610, CromoG22_2610; PlPf1_2610	Cap1_2610: Microscope observations	*Presence of spore of Epicoccum sp., and Alternaria sp..*
			CromoG7_2610: visual analysis and stereo-microscopic observation	*Hydroptila angulata, Psychodidae*
			PlPf1_2610: visual analysis and stereo-microscopic observation	*Laodelphax striatellus, Psychodidae, Macrosteles sp.*
			CromoB12_2610: visual analysis and stereo-microscopic observation	*Psammotettix sp. (prob. confinis), Anoecia sp., Psychodidae, Trixagus meybohmi*
			CromoG1_2610: visual analysis and stereo-microscopic observation	*Acrolepiopsis assectella, Zygina rahmni, Ophraella communa, Stethorus sp.*
			CromoG21_2610: visual analysis and stereo-microscopic observation	*Presence of Frankliniella occidentalis, Psychodidae, Zyginidia pullula, and Tomosvaryella kuthyi*
			CromoG22_2610: visual analysis and stereo-microscopic observation	*Zyginidia pullula, Psychodidae*
			CromoB13_2610: visual analysis and stereo-microscopic observation	*Hydroptila angulata, Calathus sp., Stephanitis pyri*
28_10_2015	Uninstall 5 Popillia traps	TFfly1, Tfly6, Tfly7, Tfly8, Tfly9		
29_10_2015	Uninstall 4 buckets traps for rhynchophorus and 2 traps for Popillia, sampling and uninstall 1 blue and 6 yellow chromotropic traps for Thrips, 5 blue and 8 yellow chromotropic traps, 1 Planococcus trap, and disappeared 1 yellow chromotropic trap (CromoG5)	SRh1, SRh2, SRh3, SRh4; Tfly3, Tfly2; CromoBTh1_2910; CromoGTh1_2910, CromoGTh2_2910, CromoGTh3_2910, CromoGTh8_2910, CromoGTh9_2910, CromoGTh10_2910; CromoB5_2910, CromoB6_2910, CromoB7_2910, CromoB10_2910, CromoB16_2910, CromoG2_2910, CromoG3_2910, CromoG4_2910, CromoG6_2910, CromoG12_2910, CromoG13_2910, CromoG14_2910, CromoG16_2910; PlPc1_2910	CromoBTh1_2910: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoGTh1_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoGTh2_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoGTh3_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoGTh8_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis, Psammotettix confinis, Hydroptila vectis, Adalia bipunctata*
			CromoGTh9_2910: visual analysis and stereo-microscopic observation	no Thrips, Psychodida
			CromoGTh10_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis, Cicadella viridis, Psychodidae*
			CromoB5_2910: visual analysis and stereo-microscopic observation	Presence of *Frankliniella intonsa, Thaumatomyia notata, Psychodidae,* and *Ischnopterapion virens*
			CromoB6_2910: visual analysis and stereo-microscopic observation	*Frankliniella intonsa*
			CromoB7_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*
			CromoB10_2910: visual analysis and stereo-microscopic observation	*Thrips tabaci, Ophraella communa*
			CromoB16_2910: visual analysis and stereo-microscopic observation	*Thrips tabaci, Ophraella communa*
			CromoG2_2910: visual analysis and stereo-microscopic observation	Presence of *Cicadella viridis, Lycaenidae, Zygina sp.* and *Empoasca pteridis*
			CromoG3_2910: visual analysis and stereo-microscopic observation	Presence of *Zyginidia pullula, Thaumatomyia notata*
			CromoG4_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis, Cicadella viridis, Zygina rhamni*
			CromoG6_2910: visual analysis and stereo-microscopic observation	*No Thrips, Rhopalosiphum padi, Zygina rhamni*
			CromoG12_2910: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis, Cicadella viridis, Dictya sp.,* Psychodidae
			CromoG13_2910: visual analysis and stereo-microscopic observation	*Frankliniella occidentalis*

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
29_10_2015	Uninstall 4 buckets traps for rhynchophorus and 2 traps for Popillia, sampling and uninstall 1 blue and 6 yellow chromotropic traps for Thrips, 5 blue and 8 yellow chromotropic traps, 1 Planococcus trap, and disappeared 1 yellow chromotropic trap (CromoG5)	SRh1, SRh2, SRh3, SRh4; Tfly3, Tfly2; CromoBTh1_2910; CromoGTh1_2910, CromoGTh2_2910, CromoGTh3_2910, CromoGTh8_2910, CromoGTh9_2910, CromoGTh10_2910; CromoB5_2910, CromoB6_2910, CromoB7_2910, CromoB10_2910, CromoB16_2910, CromoG2_2910, CromoG3_2910, CromoG4_2910, CromoG6_2910, CromoG12_2910, CromoG13_2910, CromoG14_2910, CromoG16_2910; PlPc1_2910	CromoG14_2910: visual analysis and stereo-microscopic observation	unidentifiable, degraded
			CromoG16_2910: visual analysis and stereo-microscopic observation	No Thrips, *Rhopalosiphum padi, Euleia* sp., Psychodidae
			PlPc1_2910: visual analysis and stereo-microscopic observation	Negative for *Planococcus* sp.
30_10_2015	Uninstall and sampling 10 chromotropic traps for fruit flies, 6 for Thrips, 1 multi-funnel trap for Anoplophora and 1 planotrap for planococcus; Uninstall 1 multi-funnel trap for pityophthorus and 1 multi-funnel trap for Monochamus. 2 yellow (CromoG18, CromoG20) and 1 blue (CromoB2) chromotropic traps was disappeared; Uninstall 3 apple baits; Uninstall and sampling spore caps.	CromoB1_3010, CromoG9_3010, CromoB11_3010, CromoGTh4_3010, CromoG8_3010, CromoBTh3_3010, CromoGTh5_3010, CromoG17_3010, MfAn3_3010, MfMo1, MfMPi1, CromoGTh6_3010, CromoBTh4_3010, CromoB14_3010, CromoB15_3010, CromoG15_3010, CromoB8_3010, CromoG9_3010, CromoBTh2_3010, PlPf2_3010; Mela1, Mela2, Mela3, Cap1_3010	Cap1_3010: Microscope observations	Presence of *Epicoccum* sp spores. and *Alternaria* sp.
			Cromo B1_3010: visual analysis and stereo-microscopic observation	*Drosophila suzukii,* Psychodidae
			CromoB8_3010: visual analysis and stereo-microscopic observation	Psychodidae
			CromoB11_3010: visual analysis and stereo-microscopic observation	*Drosophila suzukii, Drosophila* sp. (prob. *simulans*)
			CromoB14_3010: visual analysis and stereo-microscopic observation	*Hydroptila vectis, Psychodidae, Anoecia* sp.
			CromoB15_3010: visual analysis and stereo-microscopic observation	*Anoecia* sp., Trichoptera
			CromoBTh2_3010: visual analysis and stereo-microscopic observation	No Thrips, *Anoecia* sp., *Hydroptila vectis*
			CromoBTh3_3010: visual analysis and stereo-microscopic observation	No Thrips, *Drosophila* sp., *Eucallipterus tiliae, Omalium caesum, Rhopalosiphum rufiabdominalis*
			CromoBTh4_3010: visual analysis and stereo-microscopic observation	Presence of *Thrips tabaci, Frankliniella occidentalis* and *Trechus quadristriatus*
			CromoG8_3010: visual analysis and stereo-microscopic observation	Presence of Psychodidae, *Zyginidia pullula, Zygina rhamni*
			CromoG9_3010: visual analysis and stereo-microscopic observation	Presence of *Euleia* sp.
			CromoG15_3010: visual analysis and stereo-microscopic observation	Presence of *Zygina lunaris, Siphoninus phillyreae, Hydroptila vectis,* Aleyrodidae
			CromoG17_3010: visual analysis and stereo-microscopic observation	Presence of *Hydroptila angulata, Psychodidae,* Aleyrodidae
			CromoG19_3010: visual analysis and stereo-microscopic observation	Presence of Psychodidae
			CromoGTh4_3010: visual analysis and stereo-microscopic observation	No Thrips, *Thaumatomyia notata, Zygina* sp., *Zyginidia pullula, Ophraella communa*
			CromoGTh5_3010: visual analysis and stereo-microscopic observation	*Presence of Thrips tabaci, Frankliniella occidentalis*
			CromoGTh6_3010: visual analysis and stereo-microscopic observation	Presence of *Zyginidia pullula,* Aleyrodidae and *Trechus quadristriatus*
			PlPf2_3010: visual analysis and stereo-microscopic observation	NEGATIVE (no insects)
			MfAn3_3010: visual analysis and stereo-microscopic observation	NEGATIVE (no insects)
02_11_2015	Check and uninstall 1 blue and 2 yellow chromotropic traps and 1 blue chromotropic trap for thrips. Check and uninstall 1 Popillia trap, 1 blue chromotropic trap (CromoB3) was disappeared	CromoB4_0211; CromoG10_0211, CromoG11_0211; CromoBTh6_0211; Tfly4	CromoB4_0211: visual analysis and stereo-microscopic observation	Presence of *Coccotrypes dactyliperda, Hydroptila vectis, Dictya* sp., and *Stephanitis pyri*
			CromoBTh6_0211: visual analysis and stereo-microscopic observation	*Presence of Tenothrips sp. (prob. frici), Japananus hyalinus, Anoecia sp.*
			CromoG10_0211: visual analysis and stereo-microscopic observation	Presence of *Zygina nivea, Ophraella communa, Dictya* sp.
			CromoG11_0211: visual analysis and stereo-microscopic observation	Presence of *Hydroptila vectis, Ophraella communa, Zygina* sp.
05_11_2015	Check and uninstall 2 multi-funnel traps for anoplophora, check and replacement 4 Amsa chromotropic traps, installation of 2 chromotropic traps for thrips, 1 yellow and 2 blue chromotropic traps was disappeared (CromoB1_amsa, CromoB2_amsa, CromoG2_amsa)	CromoGTh1_amsa, CromoBTh1_Amsa, CromoG1amsa_0511, MfAn5, MfAn6	CromoG1Amsa_0511: visual analysis and stereo-microscopic observation	*Zyginidia pullula, Psammotettix* sp.
10_11_2015	uninstall and sampling 8 prism traps for Agrilus	PrAg1_1011, PrAg2_1011, PrAg3_1011, PrAg4_1011, PrAg5_1011, PrAg6_1011, PrAg9_1011, PrAg10_1011		

DATE	REASON	SAMPLE CODES	ANALYSIS CARRIED OUT	ANALYSIS RESULTS
16_11_2015	uninstall and sampling 1 prism trap for Agrilus	PrAg8_1611		
27_11_2015	uninstall and sampling 1 prism trap for Agrilus, 1 yellow and 4 blue chromotropic traps for Thrips. Check for CromoBTh7 (disappeared)	PrAg7_2711, CromoGTh7_2711, CromoBTh5, CromoBTh8_2711, CromoBTh9_2711, CromoBTh10_2711	CromoGTh7_2711: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis* and *Fieberiella florii*
			CromoBTh5_2711: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis*
			CromoBTh8_2711: visual analysis and stereo-microscopic observation	NEGATIVE (no insects)
			CromoBTh9_2711: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis*
			CromoBTh10_2711: visual analysis and stereo-microscopic observation	Presence of *Frankliniella occidentalis*
			PrAg7_2711: visual analysis and stereo-microscopic observation	NEGATIVE (no insects)
18_01_2016	Installation 3 apple baits for Phytophthora.	Mela1, Mela3, Mela5		
25_01_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_2501, Mela3_2501, Mela5_2501		
01_02_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_0102, Mela3_0102, Mela5_0102		
08_02_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_0802, Mela3_0802, Mela5_0802		
15_02_2016	Sampling and replacement 2 apple baits for Phytophthora, and new installation of 1apple bait.	Mela3_1502, Mela5_1502, reinstallata Mela1		
22_02_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_2202, Mela3_2202, Mela5_2202		
01_03_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_0103, Mela3_0103, Mela5_0103		
07_03_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_0703, Mela3_0703, Mela5_0703		
14_03_2016	Sampling and replacement 3 apple baits for Phytophthora, check 3 spy plants for Erwinia amylovora.	Mela1_1403, Mela3_1403, Mela5_1403; PsEr1, PsEr2, PsEr3		
21_03_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_2103, Mela3_2103, Mela5_2103		
29_03_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_2903, Mela3_2903, Mela5_2903		
04_04_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_0404, Mela3_0404, Mela5_0404		
11_04_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_1104, Mela3_1104, Mela5_1104		
18_04_2016	Sampling and replacement 3 apple baits for Phytophthora.	Mela1_1804, Mela3_1804, Mela5_1804		

ANNEX 8_EXPO CONTROLS

			Control Date											
N.° traps	**Trap code**	**Trapping Position Date**	**February**			**March**					**April**			
10	MfAn1	21-May												
	MfAn2	21-May												
	MfAn3	27-May												
	MfAn4	28-May												
	MfAn5	28-May												
	MfAn6	28-May												
	MfAn7	24-Jun												
	MfAn8	24-Jun												
	MfAn9	29-Jun												
	MfAn10	29-Jun												
1	MfPi1	28-May												
8	MfMo1	27-May												
	MfMo2	29-Jun												
	MfMo3	29-Jul												
10	PrAg1	1-Jun												
	PrAg2	1-Jun												
	PrAg3	25-Jun												
	PrAg4	25-Jun												
	PrAg5	25-Jun												
	PrAg6	25-Jun												
	PrAg7	25-Jun												
	PrAg8	25-Jun												
	PrAg9	25-Jun												
	PrAg10	25-Jun												
4	SRh1	5-May												
	SRh2	5-May												
	SRh3	5-May												
	SRh4	5-May												
10	Tfly1	11-May												
	Tfly2	11-May												
	Tfly3	10-Jun												
	Tfly4	10-Jun												
	Tfly5	10-Jun												
	Tfly6	15-Jun												
	Tfly7	15-Jun												
	Tfly8	15-Jun												
	Tfly9	24-Jun												
	Tfly10	24-Jun												
5	PgTh1	15-Jun												
	PgTh2	15-Jun												
	PgTh3	15-Jun												
	PgTh4	24-Jun												
	PgTh5	24-Jun												
1	PgSe	14-May												
3	PgTe1	15-Jun												
	PgTe2	24-Jun												
	PgTe3	13-Jul												
3	PgSp1	27-May												
	PgSp2	27-May												
	PgSp3	27-May												
3	PgKe1	24-Jun												
	PgKe2	15-Jul												
	PgKe3	15-Jul												
1	PlPc1	21-May												
2	Plpf1	26-May												
	Plpf2	27-May												
7	ReATf1	28-May												
	ReATf2	28-May												
	ReATf3	28-May												
	ReATf4	28-May												
	ReATf5	13-Jul												
	ReATf6	13-Jul												
	ReATf7	13-Jul												
1	ReARh1	13-Jul												
8	ReB1	21-May												
	ReB2	10-Jul												
	ReB3	10-Jul												
	ReB4	10-Jul												
	ReB5	15-Jul												
	ReB6	10-Jul												
	ReB7	20-Jul												
	ReB8	20-Jul												
6	mela1	2-Feb	9-Feb	16-Feb	23-Feb	2-Mar	9-Mar	16-Mar		30-Mar	7-Apr	13-Apr	20-Apr	27-Apr
	mela2	2-Feb	9-Feb	16-Feb	23-Feb	2-Mar	9-Mar	16-Mar	23-Mar	30-Mar	7-Apr	13-Apr	20-Apr	27-Apr
	mela3	9-Feb		16-Feb	23-Feb	2-Mar	9-Mar	16-Mar	23-Mar	30-Mar	7-Apr	13-Apr	20-Apr	27-Apr
	mela4	6-May												
	mela5	6-May												
	mela6	7-May												
1	Cap1	2-Feb	9-Feb	16-Feb	23-Feb	2-Mar	9-Mar	16-Mar	23-Mar	30-Mar	7-Apr	13-Apr	20-Apr	27-Apr
5	CromoBTh1	26-May												
	CromoBTh2	26-May												
	CromoBTh3	1-Jun												
	CromoBTh4	1-Jun												
	CromoBTh5	15-Jul												

			Control Date												
N.° traps	Trap code	Trapping Position Date	February			March					April				
5	CromoBTh6	15-Jul													
	CromoBTh7	15-Jul													
	CromoBTh8	15-Jul													
	CromoBTh9	16-Jul													
	CromoBTh10	16-Jul													
15	CromoB1	14-May													
	CromoB2	14-May													
	CromoB3	14-May													
	CromoB4	14-May													
	CromoB5	19-May													
	CromoB6	19-May													
	CromoB7	19-May													
	CromoB8	19-May													
	CromoB9	19-May													
	CromoB10	19-May													
	CromoB11	19-May													
	CromoB12	26-May													
	CromoB13	26-May													
	CromoB14	26-May													
	CromoB15	26-May													
	CromoB16	19-May													
2	CromoB1amsa	29-Jul													
	CromoB2amsa	29-Jul													
10	CromoGTh1	26-May													
	CromoGTh2	26-May													
	CromoGTh3	28-May													
	CromoGTh4	28-May													
	CromoGTh5	1-Jun													
	CromoGTh6	1-Jun													
	CromoGTh7	15-Jul													
	CromoGTh8	15-Jul													
	CromoGTh9	15-Jul													
	CromoGTh10	16-Jul													
8	CromoGAt1	29-Jun													
	CromoGAt2	29-Jun													
	CromoGAt3	29-Jun													
	CromoGAt4	29-Jun													
	CromoGAt5	14-Jul													
	CromoGAt6	14-Jul													
	CromoGAt7	16-Jul													
	CromoGAt8	16-Jul													
23	CromoG1	11-May													
	CromoG2	11-May													
	CromoG3	11-May													
	CromoG4	11-May													
	CromoG5	11-May													
	CromoG6	11-May													
	CromoG7	14-May													
	CromoG8	14-May													
	CromoG9	14-May													
	CromoG10	14-May													
	CromoG11	13-Jul													
	CromoG12	19-May													
	CromoG13	19-May													
	CromoG14	19-May													
	CromoG15	19-May													
	CromoG16	19-May													
	CromoG17	19-May													
	CromoG18	13-Jul													
	CromoG19	14-Jul													
	CromoG20	13-Jul													
	CromoG21	5-Aug													
	CromoG22	5-Aug													
	CromoG23	5-Aug													
2	CromoG1amsa	29-Jul													
	CromoG2amsa	29-Jul													
3	PsAn1	26-Aug													
	PsAn2	26-Aug													
	PsAn3	26-Aug													
3	PsAg1	28-Jul													
	PsAg2	26-Aug													
	PsAg3	26-Aug													
3	PsEr1	26-Aug													
	PsEr2	4-Sep													
	PsEr3	4-Sep													
3	PsPh1	3-Sep													
	PsPh2	3-Sep													
	PsPh3	3-Sep													
1	PsBG1	3-Sep													
1	PsFl1	4-Sep													
1	PsSh1	4-Sep													
2	PsCi1	22-Jun													
	PsCi2	27-Aug													

N.° traps	Trap code	Trapping Position Date	May							June						
10	MfAn1	21-May														
	MfAn2	21-May														
	MfAn3	27-May														
	MfAn4	28-May														
	MfAn5	28-May														
	MfAn6	28-May														
	MfAn7	24-Jun														
	MfAn8	24-Jun														
	MfAn9	29-Jun														
	MfAn10	29-Jun														
1	MfPi1	28-May														
8	MfMo1	27-May														
	MfMo2	29-Jun														
	MfMo3	29-Jul														
10	PrAg1	1-Jun														
	PrAg2	1-Jun														
	PrAg3	25-Jun														
	PrAg4	25-Jun														
	PrAg5	25-Jun														
	PrAg6	25-Jun														
	PrAg7	25-Jun														
	PrAg8	25-Jun														
	PrAg9	25-Jun														
	PrAg10	25-Jun														
4	SRh1	5-May					19-May									
	SRh2	5-May														
	SRh3	5-May					19-May									
	SRh4	5-May					19-May							15-Jun		
10	Tfly1	11-May													22-Jun	
	Tfly2	11-May												15-Jun		
	Tfly3	10-Jun														
	Tfly4	10-Jun														
	Tfly5	10-Jun														
	Tfly6	15-Jun														
	Tfly7	15-Jun														
	Tfly8	15-Jun														
	Tfly9	24-Jun														
	Tfly10	24-Jun														
5	PgTh1	15-Jun														
	PgTh2	15-Jun														
	PgTh3	15-Jun														
	PgTh4	24-Jun														
	PgTh5	24-Jun														
1	PgSe	14-May														
3	PgTe1	15-Jun														
	PgTe2	24-Jun														
	PgTe3	13-Jul														
3	PgSp1	27-May													22-Jun	24-Jun
	PgSp2	27-May													22-Jun	24-Jun
	PgSp3	27-May													22-Jun	24-Jun
3	PgKe1	24-Jun														
	PgKe2	15-Jul														
	PgKe3	15-Jul														
1	PlPc1	21-May														
2	Plpf1	26-May													22-Jun	
	Plpf2	27-May													22-Jun	
7	ReATf1	28-May													22-Jun	
	ReATf2	28-May													22-Jun	
	ReATf3	28-May													22-Jun	
	ReATf4	28-May													22-Jun	
	ReATf5	13-Jul														
	ReATf6	13-Jul														
	ReATf7	13-Jul														
1	ReARh1	13-Jul														
8	ReB1	21-May													22-Jun	
	ReB2	10-Jul														
	ReB3	10-Jul														
	ReB4	10-Jul														
	ReB5	15-Jul														
	ReB6	10-Jul														
	ReB7	20-Jul														
	ReB8	20-Jul														
6	mela1	2-Feb	4-May	11-May	14-May	18-May		25-May		1-Jun		8-Jun		22-Jun		
	mela2	2-Feb	4-May	11-May	14-May	18-May		25-May		1-Jun		8-Jun		22-Jun		
	mela3	9-Feb	4-May	11-May	14-May	18-May		25-May		1-Jun		8-Jun		22-Jun		
	mela4	6-May		11-May	14-May	18-May		25-May		1-Jun		8-Jun		22-Jun		
	mela5	6-May		11-May	14-May	18-May		25-May		1-Jun		8-Jun		22-Jun		
	mela6	7-May		11-May				25-May		1-Jun		8-Jun		22-Jun		
1	Cap1	2-Feb	4-May	11-May		18-May		25-May		1-Jun		8-Jun		22-Jun		29-Jun
5	CromoBTh1	26-May												22-Jun		
	CromoBTh2	26-May												22-Jun		
	CromoBTh3	1-Jun									4-Jun		15-Jun			
	CromoBTh4	1-Jun									4-Jun		15-Jun			
	CromoBTh5	15-Jul														

N.° traps	Trap code	Trapping Position Date	May							June						
5	CromoBTh6	15-Jul														
	CromoBTh7	15-Jul														
	CromoBTh8	15-Jul														
	CromoBTh9	16-Jul														
	CromoBTh10	16-Jul														
15	CromoB1	14-May							27-May							
	CromoB2	14-May														
	CromoB3	14-May												22-Jun		
	CromoB4	14-May												22-Jun		
	CromoB5	19-May							27-May							
	CromoB6	19-May							27-May							
	CromoB7	19-May							27-May							
	CromoB8	19-May												22-Jun		
	CromoB9	19-May												22-Jun		
	CromoB10	19-May							27-May					22-Jun		
	CromoB11	19-May														
	CromoB12	26-May														
	CromoB13	26-May												22-Jun		
	CromoB14	26-May												22-Jun		
	CromoB15	26-May												22-Jun		
	CromoB16	19-May							27-May							
2	CromoB1amsa	29-Jul														
	CromoB2amsa	29-Jul														
10	CromoGTh1	26-May														
	CromoGTh2	26-May												22-Jun		
	CromoGTh3	28-May														
	CromoGTh4	28-May														
	CromoGTh5	1-Jun									4-Jun		15-Jun			
	CromoGTh6	1-Jun											15-Jun			
	CromoGTh7	15-Jul														
	CromoGTh8	15-Jul														
	CromoGTh9	15-Jul														
	CromoGTh10	16-Jul														
8	CromoGAt1	29-Jun														
	CromoGAt2	29-Jun														
	CromoGAt3	29-Jun														
	CromoGAt4	29-Jun														
	CromoGAt5	14-Jul														
	CromoGAt6	14-Jul														
	CromoGAt7	16-Jul														
	CromoGAt8	16-Jul														
23	CromoG1	11-May							27-May							
	CromoG2	11-May							27-May							
	CromoG3	11-May							27-May							
	CromoG4	11-May							27-May							
	CromoG5	11-May							27-May							
	CromoG6	11-May							27-May							
	CromoG7	14-May												22-Jun		
	CromoG8	14-May							27-May							
	CromoG9	14-May							27-May							
	CromoG10	14-May													24-Jun	
	CromoG11	13-Jul														
	CromoG12	19-May							27-May							
	CromoG13	19-May							27-May							
	CromoG14	19-May							27-May							
	CromoG15	19-May												22-Jun		
	CromoG16	19-May												22-Jun		
	CromoG17	19-May												22-Jun		
	CromoG18	13-Jul														
	CromoG19	14-Jul														
	CromoG20	13-Jul														
	CromoG21	5-Aug														
	CromoG22	5-Aug														
	CromoG23	5-Aug														
2	CromoG1amsa	29-Jul														
	CromoG2amsa	29-Jul														
3	PsAn1	26-Aug														
	PsAn2	26-Aug														
	PsAn3	26-Aug														
3	PsAg1	28-Jul														
	PsAg2	26-Aug														
	PsAg3	26-Aug														
3	PsEr1	26-Aug														
	PsEr2	4-Sep														
	PsEr3	4-Sep														
3	PsPh1	3-Sep														
	PsPh2	3-Sep														
	PsPh3	3-Sep														
1	PsBG1	3-Sep														
1	PsFl1	4-Sep														
1	PsSh1	4-Sep														
2	PsCi1	22-Jun														
	PsCi2	27-Aug														

N.° traps	Trap code	Trapping Position Date	July													
10	MfAn1	21-May														
	MfAn2	21-May														
	MfAn3	27-May														
	MfAn4	28-May														
	MfAn5	28-May														
	MfAn6	28-May														
	MfAn7	24-Jun														
	MfAn8	24-Jun														
	MfAn9	29-Jun														
	MfAn10	29-Jun														
1	MfPi1	28-May														
8	MfMo1	27-May														
	MfMo2	29-Jun														
	MfMo3	29-Jul														29-Jul
10	PrAg1	1-Jun														
	PrAg2	1-Jun														
	PrAg3	25-Jun														
	PrAg4	25-Jun							16-Jul							
	PrAg5	25-Jun														
	PrAg6	25-Jun														
	PrAg7	25-Jun														
	PrAg8	25-Jun														
	PrAg9	25-Jun							16-Jul							
	PrAg10	25-Jun														
4	SRh1	5-May				13-Jul					20-Jul					
	SRh2	5-May													28-Jul	
	SRh3	5-May										22-Jul				
	SRh4	5-May				13-Jul					20-Jul					
10	Tfly1	11-May		9-Jul		13-Jul					20-Jul		23-Jul	27-Jul		
	Tfly2	11-May		9-Jul		13-Jul		15-Jul						27-Jul		
	Tfly3	10-Jun		9-Jul	10-Jul									27-Jul		
	Tfly4	10-Jun		9-Jul							20-Jul			27-Jul		
	Tfly5	10-Jun		9-Jul				15-Jul					23-Jul			
	Tfly6	15-Jun		9-Jul									23-Jul			
	Tfly7	15-Jun		9-Jul									23-Jul			
	Tfly8	15-Jun		9-Jul							20-Jul		23-Jul			
	Tfly9	24-Jun		9-Jul									23-Jul			
	Tfly10	24-Jun		9-Jul										27-Jul		
5	PgTh1	15-Jun										22-Jul				
	PgTh2	15-Jun										22-Jul				
	PgTh3	15-Jun										22-Jul				
	PgTh4	24-Jun											23-Jul			
	PgTh5	24-Jun														
1	PgSe	14-May														
3	PgTe1	15-Jun										22-Jul				
	PgTe2	24-Jun										22-Jul				
	PgTe3	13-Jul														
3	PgSp1	27-May														29-Jul
	PgSp2	27-May											23-Jul			29-Jul
	PgSp3	27-May										22-Jul				29-Jul
3	PgKe1	24-Jun														
	PgKe2	15-Jul														
	PgKe3	15-Jul														
1	PlPc1	21-May														
2	Plpf1	26-May														
	Plpf2	27-May														29-Jul
7	ReATf1	28-May													28-Jul	
	ReATf2	28-May														
	ReATf3	28-May										22-Jul			28-Jul	
	ReATf4	28-May														
	ReATf5	13-Jul														
	ReATf6	13-Jul										22-Jul				
	ReATf7	13-Jul														
1	ReARh1	13-Jul														
8	ReB1	21-May														29-Jul
	ReB2	10-Jul														
	ReB3	10-Jul														
	ReB4	10-Jul														
	ReB5	15-Jul														
	ReB6	10-Jul													28-Jul	
	ReB7	20-Jul														
	ReB8	20-Jul														
6	mela1	2-Feb			10-Jul									27-Jul		
	mela2	2-Feb			10-Jul									27-Jul		
	mela3	9-Feb			10-Jul									27-Jul		
	mela4	6-May			sparita									27-Jul		
	mela5	6-May			sparita									27-Jul		
	mela6	7-May			10-Jul									27-Jul		
1	Cap1	2-Feb	7-Jul				14-Jul				20-Jul			27-Jul		
5	CromoBTh1	26-May											23-Jul			
	CromoBTh2	26-May											23-Jul			
	CromoBTh3	1-Jun			10-Jul							22-Jul				
	CromoBTh4	1-Jun			10-Jul								23-Jul			
	CromoBTh5	15-Jul														

N.° traps	Trap code	Trapping Position Date	July														
5	CromoBTh6	15-Jul															
	CromoBTh7	15-Jul															
	CromoBTh8	15-Jul															
	CromoBTh9	16-Jul															
	CromoBTh10	16-Jul															
15	CromoB1	14-May									20-Jul						
	CromoB2	14-May									20-Jul						
	CromoB3	14-May										22-Jul					
	CromoB4	14-May										22-Jul					
	CromoB5	19-May										22-Jul					
	CromoB6	19-May										22-Jul					
	CromoB7	19-May										22-Jul					
	CromoB8	19-May											23-Jul				
	CromoB9	19-May										22-Jul					
	CromoB10	19-May									20-Jul						
	CromoB11	19-May									20-Jul						
	CromoB12	26-May									20-Jul						
	CromoB13	26-May											23-Jul				
	CromoB14	26-May											23-Jul				
	CromoB15	26-May											23-Jul				
	CromoB16	19-May										22-Jul					
2	CromoB1amsa	29-Jul															
	CromoB2amsa	29-Jul															
10	CromoGTh1	26-May										22-Jul					
	CromoGTh2	26-May											23-Jul				
	CromoGTh3	28-May										22-Jul					
	CromoGTh4	28-May									20-Jul						
	CromoGTh5	1-Jun			10-Jul							22-Jul					
	CromoGTh6	1-Jun			10-Jul								23-Jul				
	CromoGTh7	15-Jul															
	CromoGTh8	15-Jul															
	CromoGTh9	15-Jul															
	CromoGTh10	16-Jul															
8	CromoGAt1	29-Jun															
	CromoGAt2	29-Jun															
	CromoGAt3	29-Jun															
	CromoGAt4	29-Jun															
	CromoGAt5	14-Jul															
	CromoGAt6	14-Jul															
	CromoGAt7	16-Jul															
	CromoGAt8	16-Jul															
23	CromoG1	11-May								17-Jul							
	CromoG2	11-May									20-Jul						
	CromoG3	11-May								17-Jul							
	CromoG4	11-May								17-Jul							
	CromoG5	11-May								17-Jul							29-Jul
	CromoG6	11-May										22-Jul					
	CromoG7	14-May									20-Jul						
	CromoG8	14-May									20-Jul						
	CromoG9	14-May								17-Jul							
	CromoG10	14-May									20-Jul						
	CromoG11	13-Jul															
	CromoG12	19-May										22-Jul					
	CromoG13	19-May										22-Jul					
	CromoG14	19-May										22-Jul					
	CromoG15	19-May											23-Jul				
	CromoG16	19-May									20-Jul						
	CromoG17	19-May										22-Jul					
	CromoG18	13-Jul															
	CromoG19	14-Jul															
	CromoG20	13-Jul															
	CromoG21	5-Aug															
	CromoG22	5-Aug															
	CromoG23	5-Aug															
2	CromoG1amsa	29-Jul															
	CromoG2amsa	29-Jul															
3	PsAn1	26-Aug															
	PsAn2	26-Aug															
	PsAn3	26-Aug															
3	PsAg1	28-Jul															
	PsAg2	26-Aug															
	PsAg3	26-Aug															
3	PsEr1	26-Aug															
	PsEr2	4-Sep															
	PsEr3	4-Sep															
3	PsPh1	3-Sep															
	PsPh2	3-Sep															
	PsPh3	3-Sep															
1	PsBG1	3-Sep															
1	PsFl1	4-Sep															
1	PsSh1	4-Sep															
2	PsCi1	22-Jun															
	PsCi2	27-Aug															

N.° traps	Trap code	Trapping Position Date	August												
10	MfAn1	21-May										26-Aug			
	MfAn2	21-May										26-Aug			
	MfAn3	27-May													
	MfAn4	28-May										26-Aug			
	MfAn5	28-May										26-Aug			
	MfAn6	28-May										26-Aug			
	MfAn7	24-Jun										26-Aug			
	MfAn8	24-Jun										26-Aug			
	MfAn9	29-Jun						12-Aug				26-Aug			
	MfAn10	29-Jun										26-Aug			
1	MfPi1	28-May									25-Aug				
8	MfMo1	27-May													
	MfMo2	29-Jun										26-Aug			
	MfMo3	29-Jul													
10	PrAg1	1-Jun													
	PrAg2	1-Jun													
	PrAg3	25-Jun					11-Aug								
	PrAg4	25-Jun													
	PrAg5	25-Jun													
	PrAg6	25-Jun													
	PrAg7	25-Jun													
	PrAg8	25-Jun					11-Aug								
	PrAg9	25-Jun													
	PrAg10	25-Jun													
4	SRh1	5-May							17-Aug						
	SRh2	5-May							17-Aug						
	SRh3	5-May							17-Aug						
	SRh4	5-May							17-Aug						
10	Tfly1	11-May			5-Aug							26-Aug			
	Tfly2	11-May			5-Aug						25-Aug				
	Tfly3	10-Jun			5-Aug										
	Tfly4	10-Jun			5-Aug							26-Aug			
	Tfly5	10-Jun			5-Aug							26-Aug			
	Tfly6	15-Jun			5-Aug										
	Tfly7	15-Jun			5-Aug							26-Aug			
	Tfly8	15-Jun			5-Aug										
	Tfly9	24-Jun			5-Aug							26-Aug			
	Tfly10	24-Jun			5-Aug							26-Aug			
5	PgTh1	15-Jun		4-Aug											
	PgTh2	15-Jun		4-Aug											
	PgTh3	15-Jun		4-Aug											
	PgTh4	24-Jun													
	PgTh5	24-Jun													
1	PgSe	14-May		4-Aug											
3	PgTe1	15-Jun		4-Aug											
	PgTe2	24-Jun		4-Aug											
	PgTe3	13-Jul						12-Aug							
3	PgSp1	27-May													
	PgSp2	27-May													
	PgSp3	27-May													
3	PgKe1	24-Jun						12-Aug	17-Aug						
	PgKe2	15-Jul							17-Aug						
	PgKe3	15-Jul										26-Aug			
1	PlPc1	21-May													
2	Plpf1	26-May													
	Plpf2	27-May													
7	ReATf1	28-May													
	ReATf2	28-May													
	ReATf3	28-May													
	ReATf4	28-May			5-Aug										
	ReATf5	13-Jul													
	ReATf6	13-Jul													
	ReATf7	13-Jul			5-Aug										
1	ReARh1	13-Jul			5-Aug										31-Aug
8	ReB1	21-May													
	ReB2	10-Jul													31-Aug
	ReB3	10-Jul													
	ReB4	10-Jul													31-Aug
	ReB5	15-Jul						12-Aug							31-Aug
	ReB6	10-Jul													31-Aug
	ReB7	20-Jul													31-Aug
	ReB8	20-Jul													31-Aug
6	mela1	2-Feb												28-Aug	
	mela2	2-Feb												28-Aug	
	mela3	9-Feb												28-Aug	
	mela4	6-May												28-Aug	
	mela5	6-May												28-Aug	
	mela6	7-May												28-Aug	
1	Cap1	2-Feb				10-Aug			17-Aug		25-Aug				31-Aug
5	CromoBTh1	26-May	3-Aug												
	CromoBTh2	26-May	3-Aug												
	CromoBTh3	1-Jun	3-Aug												
	CromoBTh4	1-Jun	3-Aug					12-Aug							
	CromoBTh5	15-Jul							17-Aug						

N.° traps	Trap code	Trapping Position Date	August													
5	CromoBTh6	15-Jul			5-Aug				17-Aug							
	CromoBTh7	15-Jul														
	CromoBTh8	15-Jul						12-Aug	17-Aug							
	CromoBTh9	16-Jul							17-Aug							
	CromoBTh10	16-Jul														
15	CromoB1	14-May									25-Aug					
	CromoB2	14-May									25-Aug					
	CromoB3	14-May									25-Aug					
	CromoB4	14-May									25-Aug					
	CromoB5	19-May								18-Aug						
	CromoB6	19-May								18-Aug						
	CromoB7	19-May								18-Aug						
	CromoB8	19-May											27-Aug			
	CromoB9	19-May									25-Aug					
	CromoB10	19-May									25-Aug					
	CromoB11	19-May									25-Aug					
	CromoB12	26-May									25-Aug					
	CromoB13	26-May									25-Aug					
	CromoB14	26-May											27-Aug			
	CromoB15	26-May											27-Aug			
	CromoB16	19-May								18-Aug						
2	CromoB1amsa	29-Jul														
	CromoB2amsa	29-Jul														
10	CromoGTh1	26-May	3-Aug													
	CromoGTh2	26-May	3-Aug													
	CromoGTh3	28-May	3-Aug													
	CromoGTh4	28-May	3-Aug													
	CromoGTh5	1-Jun	3-Aug													
	CromoGTh6	1-Jun	3-Aug					12-Aug								
	CromoGTh7	15-Jul							17-Aug							
	CromoGTh8	15-Jul							17-Aug							
	CromoGTh9	15-Jul							17-Aug							
	CromoGTh10	16-Jul							17-Aug							
8	CromoGAt1	29-Jun	3-Aug					12-Aug								
	CromoGAt2	29-Jun	3-Aug													
	CromoGAt3	29-Jun	3-Aug													
	CromoGAt4	29-Jun		4-Aug												
	CromoGAt5	14-Jul						12-Aug		18-Aug						
	CromoGAt6	14-Jul			5-Aug					18-Aug						
	CromoGAt7	16-Jul						12-Aug		18-Aug						
	CromoGAt8	16-Jul						12-Aug		18-Aug						
23	CromoG1	11-May								18-Aug						
	CromoG2	11-May								18-Aug						
	CromoG3	11-May								18-Aug						
	CromoG4	11-May								18-Aug						
	CromoG5	11-May								18-Aug						
	CromoG6	11-May									25-Aug					
	CromoG7	14-May											27-Aug			
	CromoG8	14-May									25-Aug					
	CromoG9	14-May									25-Aug					
	CromoG10	14-May											27-Aug			
	CromoG11	13-Jul		4-Aug							25-Aug					
	CromoG12	19-May								18-Aug						
	CromoG13	19-May								18-Aug						
	CromoG14	19-May								18-Aug						
	CromoG15	19-May											27-Aug			
	CromoG16	19-May									25-Aug					
	CromoG17	19-May									25-Aug					
	CromoG18	13-Jul											27-Aug			
	CromoG19	14-Jul											27-Aug			
	CromoG20	13-Jul						12-Aug			25-Aug					
	CromoG21	5-Aug														
	CromoG22	5-Aug														
	CromoG23	5-Aug														
2	CromoG1amsa	29-Jul														
	CromoG2amsa	29-Jul														
3	PsAn1	26-Aug														
	PsAn2	26-Aug														
	PsAn3	26-Aug														
3	PsAg1	28-Jul														
	PsAg2	26-Aug														
	PsAg3	26-Aug														
3	PsEr1	26-Aug														
	PsEr2	4-Sep														
	PsEr3	4-Sep														
3	PsPh1	3-Sep														
	PsPh2	3-Sep														
	PsPh3	3-Sep														
1	PsBG1	3-Sep														
1	PsFl1	4-Sep														
1	PsSh1	4-Sep														
2	PsCi1	22-Jun														
	PsCi2	27-Aug														

N.° traps	Trap code	Trapping Position Date	September														
10	MfAn1	21-May															29-Sep
	MfAn2	21-May															29-Sep
	MfAn3	27-May			3-Sep												
	MfAn4	28-May															29-Sep
	MfAn5	28-May			3-Sep												
	MfAn6	28-May			3-Sep												
	MfAn7	24-Jun															
	MfAn8	24-Jun															
	MfAn9	29-Jun															
	MfAn10	29-Jun															
1	MfPi1	28-May		2-Sep													
8	MfMo1	27-May			3-Sep												
	MfMo2	29-Jun						8-Sep									29-Sep
	MfMo3	29-Jul															
10	PrAg1	1-Jun		2-Sep													
	PrAg2	1-Jun			3-Sep												
	PrAg3	25-Jun		2-Sep													
	PrAg4	25-Jun		2-Sep													
	PrAg5	25-Jun		2-Sep													
	PrAg6	25-Jun		2-Sep													
	PrAg7	25-Jun		2-Sep													
	PrAg8	25-Jun			3-Sep												
	PrAg9	25-Jun			3-Sep												
	PrAg10	25-Jun			3-Sep												
4	SRh1	5-May					7-Sep										
	SRh2	5-May						8-Sep									
	SRh3	5-May						8-Sep									
	SRh4	5-May					7-Sep										
10	Tfly1	11-May						8-Sep									
	Tfly2	11-May	1-Sep				7-Sep						17-Sep				
	Tfly3	10-Jun															
	Tfly4	10-Jun						8-Sep									29-Sep
	Tfly5	10-Jun											17-Sep		24-Sep		
	Tfly6	15-Jun															
	Tfly7	15-Jun															
	Tfly8	15-Jun						8-Sep									
	Tfly9	24-Jun															
	Tfly10	24-Jun															
5	PgTh1	15-Jun										15-Sep	17-Sep				
	PgTh2	15-Jun											17-Sep				
	PgTh3	15-Jun					7-Sep						17-Sep				
	PgTh4	24-Jun											17-Sep				
	PgTh5	24-Jun													24-Sep		
1	PgSe	14-May															
3	PgTe1	15-Jun		2-Sep													
	PgTe2	24-Jun			3-Sep												
	PgTe3	13-Jul			3-Sep												
3	PgSp1	27-May															
	PgSp2	27-May															
	PgSp3	27-May															
3	PgKe1	24-Jun															29-Sep
	PgKe2	15-Jul															29-Sep
	PgKe3	15-Jul															29-Sep
1	PlPc1	21-May					7-Sep					15-Sep					
2	Plpf1	26-May															
	Plpf2	27-May						8-Sep							24-Sep		
7	ReATf1	28-May							9-Sep								
	ReATf2	28-May							9-Sep								
	ReATf3	28-May							9-Sep								
	ReATf4	28-May								11-Sep							
	ReATf5	13-Jul							9-Sep								
	ReATf6	13-Jul							9-Sep								29-Sep
	ReATf7	13-Jul							9-Sep								
1	ReARh1	13-Jul															
8	ReB1	21-May		2-Sep													
	ReB2	10-Jul															
	ReB3	10-Jul		2-Sep													
	ReB4	10-Jul															
	ReB5	15-Jul															
	ReB6	10-Jul															
	ReB7	20-Jul															
	ReB8	20-Jul															
6	mela1	2-Feb											17-Sep				
	mela2	2-Feb											17-Sep				
	mela3	9-Feb											17-Sep				
	mela4	6-May											17-Sep				
	mela5	6-May											17-Sep				
	mela6	7-May											17-Sep				
1	Cap1	2-Feb					7-Sep				14-Sep			21-Sep		28-Sep	
5	CromoBTh1	26-May					7-Sep								24-Sep		
	CromoBTh2	26-May						8-Sep							24-Sep	28-Sep	
	CromoBTh3	1-Jun						8-Sep							24-Sep		
	CromoBTh4	1-Jun						8-Sep							24-Sep		
	CromoBTh5	15-Jul					7-Sep									28-Sep	

N.° traps	Trap code	Trapping Position Date	September														
5	CromoBTh6	15-Jul					7-Sep									28-Sep	
	CromoBTh7	15-Jul							9-Sep						24-Sep		
	CromoBTh8	15-Jul					7-Sep								24-Sep		
	CromoBTh9	16-Jul						8-Sep								28-Sep	
	CromoBTh10	16-Jul							9-Sep						24-Sep		
15	CromoB1	14-May															29-Sep
	CromoB2	14-May															29-Sep
	CromoB3	14-May															
	CromoB4	14-May															
	CromoB5	19-May											17-Sep				
	CromoB6	19-May											17-Sep				
	CromoB7	19-May											17-Sep				
	CromoB8	19-May															
	CromoB9	19-May															
	CromoB10	19-May															29-Sep
	CromoB11	19-May															29-Sep
	CromoB12	26-May															29-Sep
	CromoB13	26-May															29-Sep
	CromoB14	26-May															
	CromoB15	26-May															
	CromoB16	19-May											17-Sep				
2	CromoB1amsa	29-Jul				4-Sep											
	CromoB2amsa	29-Jul				4-Sep											
10	CromoGTh1	26-May					7-Sep								24-Sep		
	CromoGTh2	26-May						8-Sep							24-Sep		
	CromoGTh3	28-May					7-Sep								24-Sep		29-Sep
	CromoGTh4	28-May					7-Sep								24-Sep	28-Sep	
	CromoGTh5	1-Jun					7-Sep								24-Sep		
	CromoGTh6	1-Jun						8-Sep							24-Sep	28-Sep	
	CromoGTh7	15-Jul					7-Sep								24-Sep		
	CromoGTh8	15-Jul						8-Sep								28-Sep	
	CromoGTh9	15-Jul						8-Sep								28-Sep	
	CromoGTh10	16-Jul					7-Sep								24-Sep		
8	CromoGAt1	29-Jun							9-Sep								29-Sep
	CromoGAt2	29-Jun							9-Sep	11-Sep						28-Sep	
	CromoGAt3	29-Jun							9-Sep								
	CromoGAt4	29-Jun									14-Sep						
	CromoGAt5	14-Jul								11-Sep							
	CromoGAt6	14-Jul							9-Sep	11-Sep							
	CromoGAt7	16-Jul									14-Sep						
	CromoGAt8	16-Jul							9-Sep								
23	CromoG1	11-May										15-Sep					
	CromoG2	11-May										15-Sep					
	CromoG3	11-May										15-Sep					
	CromoG4	11-May										15-Sep					
	CromoG5	11-May										15-Sep					
	CromoG6	11-May															29-Sep
	CromoG7	14-May														28-Sep	
	CromoG8	14-May															29-Sep
	CromoG9	14-May															29-Sep
	CromoG10	14-May															
	CromoG11	13-Jul															
	CromoG12	19-May											17-Sep				
	CromoG13	19-May											17-Sep				
	CromoG14	19-May											17-Sep				
	CromoG15	19-May															
	CromoG16	19-May															29-Sep
	CromoG17	19-May															29-Sep
	CromoG18	13-Jul															
	CromoG19	14-Jul															
	CromoG20	13-Jul															29-Sep
	CromoG21	5-Aug				4-Sep											29-Sep
	CromoG22	5-Aug				4-Sep											29-Sep
	CromoG23	5-Aug				4-Sep											29-Sep
2	CromoG1amsa	29-Jul				4-Sep											
	CromoG2amsa	29-Jul				4-Sep											
3	PsAn1	26-Aug															
	PsAn2	26-Aug															
	PsAn3	26-Aug															
3	PsAg1	28-Jul															
	PsAg2	26-Aug															
	PsAg3	26-Aug															
3	PsEr1	26-Aug															
	PsEr2	4-Sep															
	PsEr3	4-Sep															
3	PsPh1	3-Sep															
	PsPh2	3-Sep															
	PsPh3	3-Sep															
1	PsBG1	3-Sep															
1	PsFl1	4-Sep															
1	PsSh1	4-Sep															
2	PsCi1	22-Jun															
	PsCi2	27-Aug											17-Sep				

N.° traps	Trap code	Trapping Position Date	October										
10	MfAn1	21-May											
	MfAn2	21-May											
	MfAn3	27-May											30-Oct
	MfAn4	28-May											
	MfAn5	28-May											
	MfAn6	28-May											
	MfAn7	24-Jun		5-Oct									
	MfAn8	24-Jun		5-Oct									
	MfAn9	29-Jun		5-Oct									
	MfAn10	29-Jun		5-Oct									
1	MfPi1	28-May											30-Oct
8	MfMo1	27-May											30-Oct
	MfMo2	29-Jun											
	MfMo3	29-Jul											
10	PrAg1	1-Jun											
	PrAg2	1-Jun											
	PrAg3	25-Jun											
	PrAg4	25-Jun											
	PrAg5	25-Jun											
	PrAg6	25-Jun											
	PrAg7	25-Jun											
	PrAg8	25-Jun											
	PrAg9	25-Jun											
	PrAg10	25-Jun											
4	SRh1	5-May										29-Oct	
	SRh2	5-May										29-Oct	
	SRh3	5-May										29-Oct	
	SRh4	5-May										29-Oct	
10	Tfly1	11-May									28-Oct		
	Tfly2	11-May										29-Oct	
	Tfly3	10-Jun										29-Oct	
	Tfly4	10-Jun											
	Tfly5	10-Jun						22-Oct		26-Oct			
	Tfly6	15-Jun									28-Oct		
	Tfly7	15-Jun									28-Oct		
	Tfly8	15-Jun									28-Oct		
	Tfly9	24-Jun									28-Oct		
	Tfly10	24-Jun								26-Oct			
5	PgTh1	15-Jun											
	PgTh2	15-Jun											
	PgTh3	15-Jun							23-Oct				
	PgTh4	24-Jun											
	PgTh5	24-Jun							23-Oct				
1	PgSe	14-May							23-Oct				
3	PgTe1	15-Jun											
	PgTe2	24-Jun											
	PgTe3	13-Jul											
3	PgSp1	27-May			6-Oct								
	PgSp2	27-May			6-Oct								
	PgSp3	27-May			6-Oct								
3	PgKe1	24-Jun											
	PgKe2	15-Jul											
	PgKe3	15-Jul											
1	PlPc1	21-May										29-Oct	
2	Plpf1	26-May								26-Oct			
	Plpf2	27-May											30-Oct
7	ReATf1	28-May							23-Oct				
	ReATf2	28-May							23-Oct				
	ReATf3	28-May							23-Oct				
	ReATf4	28-May							23-Oct				
	ReATf5	13-Jul							23-Oct				
	ReATf6	13-Jul							23-Oct				
	ReATf7	13-Jul							23-Oct				
1	ReARh1	13-Jul											
8	ReB1	21-May											
	ReB2	10-Jul											
	ReB3	10-Jul											
	ReB4	10-Jul											
	ReB5	15-Jul											
	ReB6	10-Jul											
	ReB7	20-Jul											
	ReB8	20-Jul											
6	mela1	2-Feb	2-Oct	5-Oct				22-Oct					
	mela2	2-Feb	2-Oct	5-Oct				22-Oct					
	mela3	9-Feb	2-Oct	5-Oct				22-Oct					
	mela4	6-May	2-Oct	5-Oct				22-Oct					
	mela5	6-May	2-Oct	5-Oct				22-Oct					
	mela6	7-May	2-Oct	5-Oct				22-Oct					
1	Cap1	2-Feb		5-Oct		13-Oct	19-Oct			26-Oct			30-Oct
5	CromoBTh1	26-May										29-Oct	
	CromoBTh2	26-May											30-Oct
	CromoBTh3	1-Jun											30-Oct
	CromoBTh4	1-Jun											30-Oct
	CromoBTh5	15-Jul											

N.° traps	Trap code	Trapping Position Date	October										
5	CromoBTh6	15-Jul											
	CromoBTh7	15-Jul											
	CromoBTh8	15-Jul											
	CromoBTh9	16-Jul											
	CromoBTh10	16-Jul											
15	CromoB1	14-May											30-Oct
	CromoB2	14-May											30-Oct
	CromoB3	14-May											
	CromoB4	14-May											
	CromoB5	19-May										29-Oct	
	CromoB6	19-May										29-Oct	
	CromoB7	19-May										29-Oct	
	CromoB8	19-May											30-Oct
	CromoB9	19-May								26-Oct			
	CromoB10	19-May										29-Oct	
	CromoB11	19-May											30-Oct
	CromoB12	26-May								26-Oct			
	CromoB13	26-May								26-Oct			
	CromoB14	26-May											30-Oct
	CromoB15	26-May											30-Oct
	CromoB16	19-May										29-Oct	
2	CromoB1amsa	29-Jul											
	CromoB2amsa	29-Jul											
10	CromoGTh1	26-May										29-Oct	
	CromoGTh2	26-May										29-Oct	
	CromoGTh3	28-May										29-Oct	
	CromoGTh4	28-May											30-Oct
	CromoGTh5	1-Jun											30-Oct
	CromoGTh6	1-Jun											30-Oct
	CromoGTh7	15-Jul											
	CromoGTh8	15-Jul										29-Oct	
	CromoGTh9	15-Jul										29-Oct	
	CromoGTh10	16-Jul										29-Oct	
8	CromoGAt1	29-Jun											
	CromoGAt2	29-Jun											
	CromoGAt3	29-Jun			6-Oct								
	CromoGAt4	29-Jun			6-Oct								
	CromoGAt5	14-Jul			6-Oct								
	CromoGAt6	14-Jul			6-Oct								
	CromoGAt7	16-Jul			6-Oct								
	CromoGAt8	16-Jul			6-Oct								
23	CromoG1	11-May								26-Oct			
	CromoG2	11-May										29-Oct	
	CromoG3	11-May										29-Oct	
	CromoG4	11-May										29-Oct	
	CromoG5	11-May										29-Oct	
	CromoG6	11-May										29-Oct	
	CromoG7	14-May								26-Oct			
	CromoG8	14-May											30-Oct
	CromoG9	14-May											30-Oct
	CromoG10	14-May											
	CromoG11	13-Jul											
	CromoG12	19-May										29-Oct	
	CromoG13	19-May										29-Oct	
	CromoG14	19-May										29-Oct	
	CromoG15	19-May											30-Oct
	CromoG16	19-May										29-Oct	
	CromoG17	19-May											30-Oct
	CromoG18	13-Jul											30-Oct
	CromoG19	14-Jul											30-Oct
	CromoG20	13-Jul											30-Oct
	CromoG21	5-Aug								26-Oct			
	CromoG22	5-Aug								26-Oct			
	CromoG23	5-Aug								26-Oct			
2	CromoG1amsa	29-Jul											
	CromoG2amsa	29-Jul											
3	PsAn1	26-Aug											
	PsAn2	26-Aug											
	PsAn3	26-Aug											
3	PsAg1	28-Jul											
	PsAg2	26-Aug											
	PsAg3	26-Aug											
3	PsEr1	26-Aug											
	PsEr2	4-Sep											
	PsEr3	4-Sep											
3	PsPh1	3-Sep											
	PsPh2	3-Sep											
	PsPh3	3-Sep											
1	PsBG1	3-Sep											
1	PsFl1	4-Sep											
1	PsSh1	4-Sep											
2	PsCi1	22-Jun											
	PsCi2	27-Aug											

N.° traps	Trap code	Trapping Position Date	November					December	January		February			
10	MfAn1	21-May												
	MfAn2	21-May												
	MfAn3	27-May												
	MfAn4	28-May												
	MfAn5	28-May		5-Nov										
	MfAn6	28-May		5-Nov										
	MfAn7	24-Jun												
	MfAn8	24-Jun												
	MfAn9	29-Jun												
	MfAn10	29-Jun												
1	MfPi1	28-May												
8	MfMo1	27-May												
	MfMo2	29-Jun												
	MfMo3	29-Jul		5-Nov										
10	PrAg1	1-Jun			10-Nov									
	PrAg2	1-Jun			10-Nov									
	PrAg3	25-Jun			10-Nov									
	PrAg4	25-Jun			10-Nov									
	PrAg5	25-Jun			10-Nov									
	PrAg6	25-Jun			10-Nov									
	PrAg7	25-Jun												
	PrAg8	25-Jun				16-Nov	27-Nov							
	PrAg9	25-Jun			10-Nov									
	PrAg10	25-Jun			10-Nov									
4	SRh1	5-May												
	SRh2	5-May												
	SRh3	5-May												
	SRh4	5-May												
10	Tfly1	11-May												
	Tfly2	11-May												
	Tfly3	10-Jun												
	Tfly4	10-Jun	2-Nov											
	Tfly5	10-Jun												
	Tfly6	15-Jun												
	Tfly7	15-Jun												
	Tfly8	15-Jun												
	Tfly9	24-Jun												
	Tfly10	24-Jun												
5	PgTh1	15-Jun												
	PgTh2	15-Jun												
	PgTh3	15-Jun												
	PgTh4	24-Jun												
	PgTh5	24-Jun												
1	PgSe	14-May												
3	PgTe1	15-Jun												
	PgTe2	24-Jun												
	PgTe3	13-Jul												
3	PgSp1	27-May												
	PgSp2	27-May												
	PgSp3	27-May												
3	PgKe1	24-Jun												
	PgKe2	15-Jul												
	PgKe3	15-Jul												
1	PlPc1	21-May												
2	Plpf1	26-May												
	Plpf2	27-May												
7	ReATf1	28-May												
	ReATf2	28-May												
	ReATf3	28-May												
	ReATf4	28-May												
	ReATf5	13-Jul												
	ReATf6	13-Jul												
	ReATf7	13-Jul												
1	ReARh1	13-Jul												
8	ReB1	21-May												
	ReB2	10-Jul												
	ReB3	10-Jul												
	ReB4	10-Jul												
	ReB5	15-Jul												
	ReB6	10-Jul												
	ReB7	20-Jul												
	ReB8	20-Jul												
6	mela1	2-Feb							18-Jan	25-Jan	1-Feb	8-Feb		22-Feb
	mela2	2-Feb												
	mela3	9-Feb							18-Jan	25-Jan	1-Feb	8-Feb	15-Feb	22-Feb
	mela4	6-May												
	mela5	6-May							18-Jan	25-Jan	1-Feb	8-Feb	15-Feb	22-Feb
	mela6	7-May												
1	Cap1	2-Feb												
5	CromoBTh1	26-May												
	CromoBTh2	26-May												
	CromoBTh3	1-Jun												
	CromoBTh4	1-Jun												
	CromoBTh5	15-Jul					27-Nov							

N.° traps	Trap code	Trapping Position Date	November					December	January		February			
5	CromoBTh6	15-Jul	2-Nov											
	CromoBTh7	15-Jul					27-Nov							
	CromoBTh8	15-Jul					27-Nov							
	CromoBTh9	16-Jul					27-Nov							
	CromoBTh10	16-Jul					27-Nov							
15	CromoB1	14-May												
	CromoB2	14-May												
	CromoB3	14-May	2-Nov											
	CromoB4	14-May	2-Nov											
	CromoB5	19-May												
	CromoB6	19-May												
	CromoB7	19-May												
	CromoB8	19-May												
	CromoB9	19-May												
	CromoB10	19-May												
	CromoB11	19-May												
	CromoB12	26-May												
	CromoB13	26-May												
	CromoB14	26-May												
	CromoB15	26-May												
	CromoB16	19-May												
2	CromoB1amsa	29-Jul		5-Nov										
	CromoB2amsa	29-Jul		5-Nov										
10	CromoGTh1	26-May												
	CromoGTh2	26-May												
	CromoGTh3	28-May												
	CromoGTh4	28-May												
	CromoGTh5	1-Jun												
	CromoGTh6	1-Jun												
	CromoGTh7	15-Jul					27-Nov							
	CromoGTh8	15-Jul												
	CromoGTh9	15-Jul												
	CromoGTh10	16-Jul												
8	CromoGAt1	29-Jun												
	CromoGAt2	29-Jun												
	CromoGAt3	29-Jun												
	CromoGAt4	29-Jun												
	CromoGAt5	14-Jul												
	CromoGAt6	14-Jul												
	CromoGAt7	16-Jul												
	CromoGAt8	16-Jul												
23	CromoG1	11-May												
	CromoG2	11-May												
	CromoG3	11-May												
	CromoG4	11-May												
	CromoG5	11-May												
	CromoG6	11-May												
	CromoG7	14-May												
	CromoG8	14-May												
	CromoG9	14-May												
	CromoG10	14-May	2-Nov											
	CromoG11	13-Jul	2-Nov											
	CromoG12	19-May												
	CromoG13	19-May												
	CromoG14	19-May												
	CromoG15	19-May												
	CromoG16	19-May												
	CromoG17	19-May												
	CromoG18	13-Jul												
	CromoG19	14-Jul												
	CromoG20	13-Jul												
	CromoG21	5-Aug												
	CromoG22	5-Aug												
	CromoG23	5-Aug												
2	CromoG1amsa	29-Jul		5-Nov										
	CromoG2amsa	29-Jul		5-Nov										
3	PsAn1	26-Aug												
	PsAn2	26-Aug												
	PsAn3	26-Aug												
3	PsAg1	28-Jul												
	PsAg2	26-Aug												
	PsAg3	26-Aug												
3	PsEr1	26-Aug												
	PsEr2	4-Sep												
	PsEr3	4-Sep												
3	PsPh1	3-Sep												
	PsPh2	3-Sep												
	PsPh3	3-Sep												
1	PsBG1	3-Sep												
1	PsFl1	4-Sep												
1	PsSh1	4-Sep												
2	PsCi1	22-Jun												
	PsCi2	27-Aug												

N.° traps	Trap code	Trapping Position Date	March					April		
10	MfAn1	21-May								
	MfAn2	21-May								
	MfAn3	27-May								
	MfAn4	28-May								
	MfAn5	28-May								
	MfAn6	28-May								
	MfAn7	24-Jun								
	MfAn8	24-Jun								
	MfAn9	29-Jun								
	MfAn10	29-Jun								
1	MfPi1	28-May								
8	MfMo1	27-May								
	MfMo2	29-Jun								
	MfMo3	29-Jul								
10	PrAg1	1-Jun								
	PrAg2	1-Jun								
	PrAg3	25-Jun								
	PrAg4	25-Jun								
	PrAg5	25-Jun								
	PrAg6	25-Jun								
	PrAg7	25-Jun								
	PrAg8	25-Jun								
	PrAg9	25-Jun								
	PrAg10	25-Jun								
4	SRh1	5-May								
	SRh2	5-May								
	SRh3	5-May								
	SRh4	5-May								
10	Tfly1	11-May								
	Tfly2	11-May								
	Tfly3	10-Jun								
	Tfly4	10-Jun								
	Tfly5	10-Jun								
	Tfly6	15-Jun								
	Tfly7	15-Jun								
	Tfly8	15-Jun								
	Tfly9	24-Jun								
	Tfly10	24-Jun								
5	PgTh1	15-Jun								
	PgTh2	15-Jun								
	PgTh3	15-Jun								
	PgTh4	24-Jun								
	PgTh5	24-Jun								
1	PgSe	14-May								
3	PgTe1	15-Jun								
	PgTe2	24-Jun								
	PgTe3	13-Jul								
3	PgSp1	27-May								
	PgSp2	27-May								
	PgSp3	27-May								
3	PgKe1	24-Jun								
	PgKe2	15-Jul								
	PgKe3	15-Jul								
1	PlPc1	21-May								
2	Plpf1	26-May								
	Plpf2	27-May								
7	ReATf1	28-May								
	ReATf2	28-May								
	ReATf3	28-May								
	ReATf4	28-May								
	ReATf5	13-Jul								
	ReATf6	13-Jul								
	ReATf7	13-Jul								
1	ReARh1	13-Jul								
8	ReB1	21-May								
	ReB2	10-Jul								
	ReB3	10-Jul								
	ReB4	10-Jul								
	ReB5	15-Jul								
	ReB6	10-Jul								
	ReB7	20-Jul								
	ReB8	20-Jul								
6	mela1	2-Feb	1-Mar	7-Mar	14-Mar	21-Mar	29-Mar	08_apr	11_apr	18_apr
	mela2	2-Feb								
	mela3	9-Feb	1-Mar	7-Mar	14-Mar	21-Mar	29-Mar	08_apr	11_apr	18_apr
	mela4	6-May								
	mela5	6-May	1-Mar	7-Mar	14-Mar	21-Mar	29-Mar	08_apr	11_apr	18_apr
	mela6	7-May								
1	Cap1	2-Feb								
5	CromoBTh1	26-May								
	CromoBTh2	26-May								
	CromoBTh3	1-Jun								
	CromoBTh4	1-Jun								
	CromoBTh5	15-Jul								

N.° traps	Trap code	Trapping Position Date	March					April		
5	CromoBTh6	15-Jul								
	CromoBTh7	15-Jul								
	CromoBTh8	15-Jul								
	CromoBTh9	16-Jul								
	CromoBTh10	16-Jul								
15	CromoB1	14-May								
	CromoB2	14-May								
	CromoB3	14-May								
	CromoB4	14-May								
	CromoB5	19-May								
	CromoB6	19-May								
	CromoB7	19-May								
	CromoB8	19-May								
	CromoB9	19-May								
	CromoB10	19-May								
	CromoB11	19-May								
	CromoB12	26-May								
	CromoB13	26-May								
	CromoB14	26-May								
	CromoB15	26-May								
	CromoB16	19-May								
2	CromoB1amsa	29-Jul								
	CromoB2amsa	29-Jul								
10	CromoGTh1	26-May								
	CromoGTh2	26-May								
	CromoGTh3	28-May								
	CromoGTh4	28-May								
	CromoGTh5	1-Jun								
	CromoGTh6	1-Jun								
	CromoGTh7	15-Jul								
	CromoGTh8	15-Jul								
	CromoGTh9	15-Jul								
	CromoGTh10	16-Jul								
8	CromoGAt1	29-Jun								
	CromoGAt2	29-Jun								
	CromoGAt3	29-Jun								
	CromoGAt4	29-Jun								
	CromoGAt5	14-Jul								
	CromoGAt6	14-Jul								
	CromoGAt7	16-Jul								
	CromoGAt8	16-Jul								
23	CromoG1	11-May								
	CromoG2	11-May								
	CromoG3	11-May								
	CromoG4	11-May								
	CromoG5	11-May								
	CromoG6	11-May								
	CromoG7	14-May								
	CromoG8	14-May								
	CromoG9	14-May								
	CromoG10	14-May								
	CromoG11	13-Jul								
	CromoG12	19-May								
	CromoG13	19-May								
	CromoG14	19-May								
	CromoG15	19-May								
	CromoG16	19-May								
	CromoG17	19-May								
	CromoG18	13-Jul								
	CromoG19	14-Jul								
	CromoG20	13-Jul								
	CromoG21	5-Aug								
	CromoG22	5-Aug								
	CromoG23	5-Aug								
2	CromoG1amsa	29-Jul								
	CromoG2amsa	29-Jul								
3	PsAn1	26-Aug								
	PsAn2	26-Aug								
	PsAn3	26-Aug								
3	PsAg1	28-Jul								
	PsAg2	26-Aug								
	PsAg3	26-Aug								
3	PsEr1	26-Aug								
	PsEr2	4-Sep								
	PsEr3	4-Sep								
3	PsPh1	3-Sep								
	PsPh2	3-Sep								
	PsPh3	3-Sep								
1	PsBG1	3-Sep								
1	PsFl1	4-Sep								
1	PsSh1	4-Sep								
2	PsCi1	22-Jun								
	PsCi2	27-Aug								

ANNEX 9_GENERIC SAMPLES

Sample Name	Sample Date	Material Type	Sample Location	Notes / Suspicions / Sample reason
Var1_0406	4-Jun	Quinoa	Cereals and tubers Cluster	Foliar miners control
Var2_0106	1-Jun	Pieris's chrysalis on Brassica	Brazil Pavilion	Foliar miners control
Va3_0106	1-Jun	Cocoons on Brassica	Brazil Pavilion	Generic check
Var4_0406	4-Jun	Lupinus sp.	Biomediterranean Cluster	Foliar miners control
Var6_1506	15-Jun	Quinoa leaves with parasitized caterpillar	Cereals and tubers Cluster	Controllo generico su insetto
Var7_1506	15-Jun	Juglans	Pad. Iran	Check on Juglans leaves
Var8_1506	15-Jun	Juglans	Hortus next Mexican Pavilion	Check on rotted fruits
Var9_2206	22-Jun	Wisteria	Hortus Eataly	Aphid check
Var10_2206	22-Jun	Moth	Hortus Eataly	Controllo generico su insetto
Var11_1007	10-Jul	Thrips material	Brazil Pavilion	Foliar miners control
Var12_2707	27-Jul	Thrips material	Brazil Pavilion	Foliar miners control
Var13_0408	4-Aug	PgTe1 trap material	Cereals and tubers Cluster	Tecia check
Var14_0508	5-Aug	TFly7 trap material	Green perimeter behind coffee cluster	Popillia check
Var15_0508	5-Aug	ReATf7 trap material	UN Garden	Tephritidae chack
Var16_0508	5-Aug	TFly9 trap material	Green perimeter behind San paolo Pavillion	Popillia check
Var17_0508	5-Aug	ReATf4 trap material	Children park	Tephritidae chack
Var18_0809	8-Sep	Leaves and Cochineal on Limon trees	Children park	Planococcus citri check

ANNEX 10_INSECTS TABLE

Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Coleoptera	Apionidae	*Protapion fulvipes*		(Geoffroy, 1785)	Present in most of Europe and North America	Parasitic Larva of Trifolium spp.(White Clover Seed Weevil)	Infesting Insect
			Ischnopterapion virens		(Herbst 1797)	Palearctic distribution bug, new for North America	Infesting insect of herbaceous plants, various Fabaceae. Favorite Plant Clover	Infesting Insect
		Anobidae	*Stegobium paniceum*		(Linnaeus, 1758)	Widespread insect in all parts of the world	Anobio of bread, substances of vegetable origin rich in starch	Foodstuffs
		Bostrichidae	*Lyctus africanus*		(Lesne, 1907)	The beetle is widespread in Africa especially in the western and Central, the presence in Italy can be found occasionally	Wood, cellulose, papaya seeds, ginger, licorice	Foodstuffs
		Carabidae	*Calathus* sp.		(Bonelli, 1810)		Predators	Predator
			Trechus quadristriatus		(Schrank 1781)	Present throughout Europe	Predator Insect	Predator
			Harpalus rufipes		(De Geer, 1774)	From Arizona, across Europe, to North Africa and eastern China.	Polyphagous insect, prefers open, dry habitats and light soils. Once it was parasite of strawberries	Polyphagous
		Cerambycidae	*Chlorophorus varius varius*		(Muller, 1766)	Present in central and southern Europe, Western Siberia and Asia minor.	Xylophagous insect, of various hardwoods and dry, including vine branches	Xylophagous
		Coccinellidae	*Adalia bipunctata*		(Linnaeus, 1758)	Common throughout Europe	The Ladybird is aphidophagous, especially living aphids on cereals and vegetable	Predator
			Adalia decempunctata		(Linnaeus, 1758)	Common throughout Europe	The Ladybird preys on the living aphids on different plants	Predator
				Harmonia axyridis	(Pallas, 1773)	Species originating in eastern Asia, introduced in Europe in 1982	Predator, a species of Asian origin	Exotic Predator
			Propylaea 14-punctata		(Linnaeus, 1758)	Common insect in almost all of Europe	Predator of living aphids on maize	Predator
			Scymnus auritus		(Thunberg, 1795)	Ladybug present throughout Europe	Predator	Predator
			Scymnus sp.		Kugelann, 1794		Predator	Predator
			Stethorus sp.		Weise, 1885		Predator	Predator
			Stethorus punctillum		(Weise, 1891)	Common throughout Europe	Predator of mites, used for biological fight	Predator of mites
		Chrisomelidae	*Altica oleracea*		(Linnaeus, 1758)	Present throughout Europe	Larvae and adult phytophagous, different species, especially Onagraceae and Rosaceae	Weed
			Bruchus brachialis		(Fahraeus, 1839)	Present in southern Europe	Fabaceae parasites	Weed
			Chaetocnema tibialis		(Illiger, 1807)	Spread throughout the Euro-Asian regions of south, central Asia and the Far East region. In Italy It is common everywhere	Swiss chard, adult	Weed beet
				Diabrotica virgifera virgifera	(LeConte, 1858)	Originating in America, spread out of control and destined to grow	Maize, larvae attack roots leading to stunted plant development up to entrapment, North American origin	Exotic Corn Insect
			Galerucella luteola		(Muller)	Spread across Europe, accidentally introduced to North America. In Italy it is present in all regions.	Elm Specific Leafer	Elm Weed
				Luperomorpha xanthodera	(Fairmaire, 1888)	Originating in Asia and widely spread in China, it has come to Europe accidentally. In Italy the presence was detected in Tuscany in 2007, in the greenhouse, in nurseries of ornamental plants.	Parasite of Rosa spp. and various ornamental, Asian origin (China), found in Tuscany in 2007. Phytophagous in the larval and adult stage, feeding on roots and stems (larvae), leaves, flowers and pollen (larvae and adults).	Exotic Polyphagous Insect
				Ophraella communa	(LeSage, 1986)	Present in Italy	It feeds at the expense of the leaves and flowers of Ambrosia Artemisiiflora, Origin North America	Exotic Polyphagous Insect
		Curculionidae	*Coccotrypes dactyliperda*		(Fabricius, 1801)	Widespread in tropical, subtropical and temperate-warm regions. In Italy It was observed on batches of scrap dates destined to the distillation industries and on ornamental palm fruits.	Date stone beetle(Phoenix dactylifera), it attacks the maturing dates.	Haunting dates Insect

Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Coleoptera	Curculionidae	*Ips sexdentatus*		(Börner, 1776)	Originating in Asia and Europe. Widespread throughout Europe and Turkey, also present in Siberia. In Italy widespread especially in the northern regions, Tuscany Calabria and Sardinia	Black Pine and Scots pine	Infesting Pines Insect
			Sitona hispidulus		(Fabricius, 1776)	Beetle widespread in Europe, Siberia, Caucasus and Syria and for accidental introduction to the United States	Clover weevil, on clover, alfalfa and other forage legumes	Infesting Insect
			Xyleborinus saxeseni		(Ratzeburg, 1837)	Present throughout Europe	Hardwood polyphagous, preference for Castanea, Fagus, Juglans, Prunus, Quercus,occasionally on conifers	Infesting Insect
		Elateridae	*Athous haemorrhoidalis*		(Fabricius, 1801)	Widespread throughout Europe, the Middle East and Siberia.	Polyphagous, damage to radical apparatus, herbaceous and forestry crops in nursery	Infesting Insect
		Throscidae	*Trixagus meybohmi*		(Leseigneur, 2005)	Species described recently, is distributed in different countries Central and Southern Europe (Leseigneur, 2005).	There's no data on his biology. Leseigneur (2005) assumes that habits are similar to those of Trixagus Dermestoides (Linnaeus, 1767), whose larvae live in Soil around dead trees and stumps just below Humus layer, nourishing of associated mushrooms The Roots (Burakowski, 1975).	Decomposer Insect
		Cryptophagidae	*Cryptophagus pilosus*		(Gyllenhal, 1827)	Present in Europe	Wheat, flour, bread and dried fruit infesting insect	Foodstuffs insect
		Lathridiidae	*Corticarina sp.*		(Reitter, 1880)		Family found on fungi, rotting substances, stored products	Foodstuffs insect
		Mycetophagidae	*Typhaea stercorea*		(Linnaeus, 1758)	Cosmopolitan insect and is common everywhere	It feeds on molds and pollens, moldy foodstuffs	Foodstuffs insect
		Scarabaeidae	*Cetonia aurata*		(Linnaeus, 1761)	Spreading Area comprising Europe, part of Asia and North Africa.	Damage to flowers of some ornamental plants (rosae, peony, Viburnum,..)	Adult infesting Ornamental plants. Detritophagous larvae
			Oryctes nasicornis		(Linnaeus, 1758)	Beetle is widespread in most of Europe, including Italy.	Adult polyphagous. Larvae feeds at the expense of wood rotting tree plants. In some circumstances, attacks on roots were also observed.	Larvae decomposers of wood and roots
		Staphylinidae	*Omalium caesum*		(Gravenhorst, 1806)	Present throughout Europe		Predator Insect
	Lepidoptera	Acrolepiidae	*Acrolepiopsis assectella*		(Zeller, 1839)	Distribution Palearctic, present in the northern areas in Italy .	Leek Moth, preferential leek parasite, less frequently garlic onion	Parasitic leek Insect
		Choreutidae	*Choreutis nemorana*		(Hübner, 1799)	Widespread in all the Italian regions and in the remaining Mediterranean area, up to central Asia and the Canary Islands	Fig Moth	Exclusive Fig Pest Insect (Ficus carica)
		Crambidae	*Duponchelia fovealis*		(Zeller, 1847)	Endemic species of the Mediterranean region and the Canary Islands, widespread in Africa, Asia minor, in the greenhouse crops of Liguria and northern Europe	Pest insect of plants bred in greenhouse) Lisianthus, Begonia, Gerbera, Kalanchoe, Cyclamen, Poissetia)	Infesting Insect
			Nomophila noctuella		(Denis & Schiffermüller, 1775)	Cosmopolitan Moth	Larvae feed of various herbaceous and clover	Parasitic insect polyphagous of alfalfa, cereals, several grasses grassland and spontaneous plants
			Paracorsia repandalis		(Denis & Schiffermüller, 1775)	Cosmopolitan Moth	Larvae feed Teucrium scorodonia, Solidago virgaurea, Origanum vulgare etc.	infesting insect of different species of Verbascum
		Gelechiidae	*Aproaerema anthyllidella*		(Hübner, 1813)	Present throughout Europe	Larvae feed on different herbaceous species	Infesting Insect
			Scrobipalpa ocellatella		(Boyd, 1858)	Distribution throughout Europe, North Africa, Asia minor	Deleafer Chenopodiaceae, lives at the expense of coltivatee beets on Beta maritima	Infesting Insect
			Sitotroga cerealella		(Olivier, 1789)	Native to tropical America, cosmopolitan spread	Foodstuffs insect Real Moth of wheat	Foodstuffs
			Tuta absoluta		(Meyrick, 1917)	Originating South America, introduced in Italy from 2008	Invasive of cultivated and spontaneous Solanaceae plants	Exotic invasive Species
		Lycaelidae	n.i.					

Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Lepidoptera	Noctuidae	*Autographa gamma*		(Linnaeus, 1758)	Entire European continent, Asia, North Africa and North America distribution	Silvery leafer, polyphagous species of vegetable plants	Vegetable infestation Species
			Helicoverpa armigera		(Hübner, 1813)	Very large distribution includes Europe, Africa, Asia, Oceania, Australia and New Zealand	Tomato silvery, polyphagous	Weed species
			Heliothis peltigera		([Denis & Schiffermüller], 1775)	Widespread in Europe, near East, South Asia, northern Africa and North America	Polyphagous species, nourishes of cultivated and spontaneous herbaceous plants	Weed species of herbaceous plants
			Spodoptera exigua		(Hübner, 1808)	Subtropical, cosmopolitan origin	Polyphagous species of numerous cultivated plants and spontaneous herbs	Weed species of herbaceous plants
		Plutellidae	*Plutella xylostella*		(Linnaeus, 1758)	Euro-Asian distribution also present in America, Africa Australia	Larva cruciferous Miner	Cruciferous infestation species
		Pieridae	*Pieris rapae*		(Linnaeus, 1758)	Widespread throughout the IS region	The butterfly lives on turnip cabbages and numerous cruciferous.	Cruciferous infestation species
		Pyralidi	*Cadra cautella*		(Walker, 1863)	Cosmopolitan Moth	The larva live on dried fruit, cocoa, walnuts and cereal seeds. Dried fruit Moth	Foodstuffs
		Tineidae	*Monopis imella*		(Hübner, 1813)		Larvae feed on material of animal origin and dried vegetable	Decomposer Dry Organic Substance
			Nemapogon clematella		(Fabricius, 1781)	Cosmopolitan	The larva feeds on kernels, peanuts, dried fruits and fungi and flours.	Foodstuffs
		Tortricidae	*Cacoecimorpha pronubana*		(Hübner, 1799)	Mediterranean basin origin, widespread in most of the European region excluding colder areas	Carnations are guests of choice but very polyphagous found also on various ornamental plants	Insect infesting Ornamental Plants
			Clepsis peritana		(Clemens, 1860)	Present in Spain, Cuba, North America.	Strawberries and Gardens moth	Haunting insect gardens and strawberries
			Cydia (Grapholita) funebrana		(Treitschke, 1835)	Origin Central Europe, distribution Palearctic, in Italy everywhere	Larva lives at the expense of the plum, with also attacks on peach, apricot and cherry	Insect infestation of the Pomacee
			Grapholita janthinana		(Godart, 1835)	Present in most of Europe	Larva feeds on Craetegus, Prunus and Sorbo	Infesting insect of different plants
			Gypsonoma aceriana		(Duponchel, 1843)	Widespread in southern Europe, in Italy, common especially in the northern regions.	Moth lives on Poplar	Poplar infesting Insect
			Pammene albuginana		(Guenée, 1845)	Asia Minor origin, located in Italy in 1992	Host plants genus Quercus	Exotic invasive Species
	Tisanotteri	Aeolothripidae	*Aeolothrips intermedius*		(Bagnall, 1934)	Cosmopolitan	Thrip Predator of other insects	Predator species
		Phlaeothripidae	*Haplothrips aculeatus*		(Fabricius, 1803)	Cosmopolitan	Host of different species belonging to the family of the Poaceae and Malvaceae	Weed species
			Haplothrips leucanthemi		(Schrank, 1781)	Originating in northern Europe	Various Asteraceae, also Trifolium sp. (Fabaceae) and Plantago sp. (Plantaginaceae)	Weed species
			Haplothrips sp.		(Amoyet & Serville, 1853)			
		Thripidae	*Anaphothrips obscurus*		(Muller, 1776)	Possible Origin Western Asia, cosmopolitan distribution in all temperate regions	Grasses and various cereal crops, including Avena, Hordeum, Secale, Triticum and Zea (Poaceae)	Weed species
			Frankliniella intonsa		(Trybom, 1895)	Thrip common throughout Italy and is also present in other European countries	Species remarkably polyphagous, lives on considerable number of cultivated and spontaneous plants.	Weed species
			Frankliniella occidentalis		(Pergande, 1895)	Originating from the Nearctic region, it is present in the western half of the American continent as well as South America New Zealand and Hawaii, present in Europe. In Italy it is mainly located in Liguria and in the southern regions	Polyphagous species, is reported on more than 250 hosts	Weed species
			Frankliniella tenuicornis		(Uzel, 1895)	Cosmopolitan species. In Italy It is common throughout the territory	Species that habitually lives on spontaneous grasses	Weed species

Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Tisanotteri	Thripidae	*Microcephalothrips abdominalis*		(D. L. Crawford, 1910)	Originario dell'America centrale, diffuso in america e nelle regioni tropicali e subtropicali	Various Asteraceae, sometimes abundant in crops of sunflowers (Helianthus annuus) (Asteraceae).	Weed species
			Tenothrips frici		(Uzel, 1895)	Originating in the Mediterranean Area spread throughout Europe, introduced in California, Oregon, Washington, Georgia, Colombia, Argentina, Uruguay, Hawaii, New Zealand, southern Australia.	Associated with various Asteraceae, particularly weedy species	Weed species
			Thrips flavus		(Schrank, 1776)	Eurasian origin diffused everywhere	Highly polyphagous species	Weed species
				Thrips hawaiiensis	(Morgan, 1913)	Originating in South Asia or the Pacific, abundantly present in Asia and the Pacific Islands, also in Jamaica and some Southern states in the USA, including California. Introduced in Europe in 2008, there is no presence in Italy even if presumed	Highly polyphagous, but often associated with the genus Gardenia (Rubiaceae)	Weed species
			Thrips major		(Uzel, 1895)	Thrip present in Europe, Siberia and North America. In Italy common in they of the regions	Harmful to nectarines and roses, also found on various herbaceous and arboreal plants	Weed species
			Thrips physapus		(Linnaeus, 1758)	Cosmopolitan distribution	Host species of flowers of numerous Asteraceae	Weed species
			Thrips tabaci		(Lindeman, 1889)	Submediterranean distribution. In Italy is present everywhere	Considerably polyphagous. Host of choice is the onion but it is also common on other vegetable and on floral plants as well as on numerous herbaceous plants belonging to several families and even hardwood and bearing	Weed species
			Thrips trehernei		(Priesner, 1927)	Abundant species in Europe and North America	Host species on different plants belonging to the family Asteraceae	Weed species
	Rincota	Aphididae	*Anoecia* sp.		(Koch, 1857)			
			Aphis craccivora		(Koch, 1854)		Black aphid of legumes	Infestant Legumes
			Eucallipterus tiliae		(Linnaeus, 1758)	Originating in Eurasia	Linden aphid	Linden Weed
			Rhopalosiphum padi		(Linnaeus, 1758)	Palearctic origin, currently widespread in the new and old continent	Aphid of Prunus pado	Prunus pado Weed
			Rhopalosiphum rufiabdominalis		(Sasaki, 1899)	Cosmopolitan species. Present in Italy	Infesting Rice Insect (Oryza sativa), Banana, several Cyperaceae, Poaceae (Graminae), Solanaceae and various ornamental	Infesting Insect
		Aphrophoridae	*Philaenus spumarius*		(Linnaeus, 1758)	Widespread throughout the Palearctic region	Polyphagous develops on a large number of spontaneous and cultivated herbaceous plants	Weed infestation, Xylella vector
		Aleyrodidae	*Dialeurodes citri*		(Ashmead, 1885)	Originating in eastern Asia, widespread in East Asia, in the countries of the Mediterranean basin, in Russia, Turkey and the American continent.	He usually lives on Citrus spp. but also present on Kaki and Privet	Tree Weed Infestation
			Siphoninus phillyreae		(Haliday, 1835)	Widespread in most of Europe, in North Africa, in the Middle East, India and Pakistan	The insect lives on the apple tree and a few other plants (pear, hawthorn, ash, olive and olive)	Tree Weed Infestation
		Anthocoridae	*Orius majusculus*		(Reuter, 1879)	Insect in Italy common in the northern regions	Insect that constitutes a better instrument for the control of thrips in those crops where these colonize and damage not only the flowers but also or more the leaves, as in the case of cucumber, eggplant or others.	Predatory thrips Insect
		Cicadellidae	*Cicadella viridis*		(Linnaeus, 1758)	Widespread in most of Europe, central Asia and Japan	Buzzer lives on Arundo, Brassica, Cyperus, Juncus, Ranunculus and Panicum	Infestation in different species, in America transmits bacterial of the Pierce screw

Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Rincota	Cicadellidae	*Euscelidius variegatus*		(Kirschbaum, 1858)	Widespread in Europe, Asia and the Mediterranean countries, in Italy in all regions	The insect lives on different grassland plants but is frequently found also in the vineyards	Weeding plants and vines, vector of phytoplasmas, (phyllody of Trifolium spp.) (Grapevine of the Vine)
				Eutettix variabilis	(Hepner, 1942)	Insect described for North America (Canada) present in South Ontario		Infesting Oaks Insect
			Fieberiella florii		(Stal, 1864)	In Italy widespread mainly northern regions	The insect lives above all on dicotyledonous shrubs but also found in apples and other orchards of Rosaceae	Infestation of various shrubs and arboreal species. Phytoplasmas vector (Aster jellow) (vein of Stone ESFYP)
				Hishimonus hamatus	(Kuoh 1976)	Originating in Asia, the first report for Europe in Slovenia in 2012 (EPPO). For Italy, there are various unofficial reports, in the Internet forums (Www.entomologiitaliani.net and www.naturamediterraneo.com), of finds in Lombardy already in 2008, on the island of Elba in July 2010, in Bovolone (VR) in November 2011 and in Castel Maggiore (BO) at the end of 2012	The insect feeds on several ornamental plants, Ligustrum lucidum, L. japonicum, Lagerstroemia sp. indica, Euonymus japonicus, Chamaecyparis lawsoniana and Cupressus sempervirens.	Weeding ornamental plants, potential phytoplasmas vector
			Japananus hyalinus		(Osborn, 1900)	East-Asian origin, widespread in the United States and in various European countries. In Italy it is widespread in the north, Tuscany and Umbria	The insect develops on Acer plants, also present in the vineyards with maples nearby.	Insect infesting of Maples
				Jikradia sp.	(Nielson, 1979)			
			Macrosteles sp.		(Fieber, 1866)	Cosmopolitan		Insect infesting polyphagous on horticultural species
			Orientus ishidae		(Matsumura, 1902)	Originating East Asia, Palearctic, Nearctic and eastern distribution. Present in Italy	The species can be found on Salix, Acer, Betula, Carpinus, Crataegus and Malus and vine	Insect infesting tree species, potential vector FD (Flavescence dorée)
			Psammotettix sp.		(Haupt, 1929)			
			Psammotettix confinis		(Dahlbom, 1850)	Species of Palearctic origin. Throughout Europe and in all of our regions.	Insect linked to herbaceous plants, they frequent humid environments.	Weeding herbaceous species
			Scaphoideus titanus		(Ball, 1932)	Originating in the United States and Canada. Widespread in Europe in the southern departments from the Atlantic to the Mediterranean, Present in Italy	Ampelophagous species	Haunting vine insect
			Empoasca pteridis		(Dahlbom 1850)	Insect present in Europe, in the Palearctica ecozone, in North Africa and in the ' Afro-tropical ' ecozone	Nymphs and adults are pests of different crops. They have a large number of guest plants Vicia Faba, Phaseolus vulgaris, Pisum sativum, Solanum tuberum, Solanum lycopersicum, Solanum Melongena and Cucumis sativus.	Insect infesting horticultural Plants
			Typhlocyba sp.		(Germar, 1833)			
			Zygina lunaris		(Mulsant & Rey, 1855)	Species present in Europe and Italy	The insect lives on different tree species: Acer sp., Populus nigra, Rosa sp., Salix Alba, Salix babylonic, Salix fragilis, Salix purpurea, Salix sp.	Insect infesting Tree Plants
			Zygina nivea		(Mulsant & Rey, 1855)	Species present in Europe and Italy	Weed species usually associated with white poplar	Infesting Insect
			Zygina rhamni		(Ferrari, 1882)	Widespread in southern Europe, present in all regions in Italy	Insect lies on Vine	Haunting vine insect
			Zyginidia pullula		(Boheman, 1845)	Widespread widely in the Palearctic region, present in Europe and northern Italy	The insect lives on maize, sorghum and other cereals (wheat, barley, rye), as well as to several spontaneous and horticultural herbaceous plants	infesting Insect polyphagous

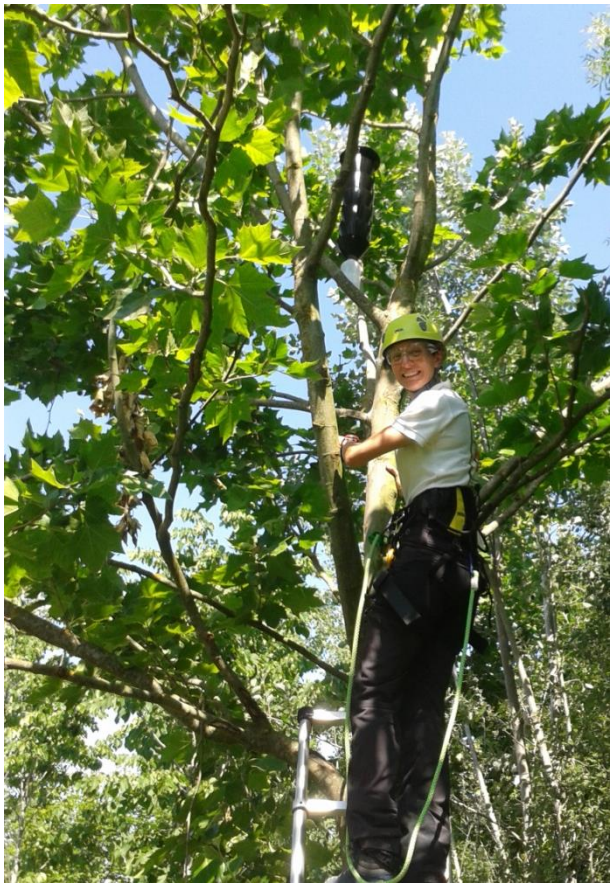
Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Rincota	Coccidae	*Saissetia coffeae*		(Walker, 1852)	Present throughout Italy	Present on ornamental plants in greenhouse, it can be found on ferns, Ficus, Cymbidium, Juniper, Cycas, Cordyline, Croton, Anthurium, Asperagus, Begonia etc.	Insect infesting Greenhouse Plants
		Corixidae	*Sigara striata*		(Linnaeus, 1758)		Aquatic insect	Aquatic insect
		Delphacidae	*Laodelphax striatellus*		(Fallen, 1826)	Palearctic distribution, present throughout Italy	This insect has as regular guests, wheat, barley, rice and several spontaneous grasses	infesting insect grasses
		Dictyopharidae	*Dictyophora europaea*		(Linnaeus, 1758)	Cosmopolitan	Insect polyphagous of numerous herbaceous pinae, although its favorite guest plants seem to be. Amaranth (Amaranthus retroflexus) and nettle (Urtica dioica)	Insect infesting herbaceous plants, it has been shown that it can be vector of flavescence doree from C. Vitalba to vine variety but not from grape to grape variety
		Flatidae	*Metcalfa pruinosa*		(Say, 1830)	Originating in North America. Present in Italy	Polyphagous species of about forty hosts including bushes and weeds.	Invasive insect
		Lygaeidae	*Nysius senecionis*		(Schilling, 1829)	Widespread Europe West Asia and Africa	This bug has as host different species of cultivated and spontaneous composites	Insect infesting herbaceous plants
			Nysius graminicola		(Kolenati, 1845)	The distribution includes the entire southern Central Europe and the entire Circummediterranea region. In Italy It is common everywhere	Polyphagous species of several spontaneous herbaceous plants and different horticultural crops	Insect infesting herbaceous plants
			Nysius sp.		(Dallas, 1852)			
			Megalonotus sp.		(Fieber, 1860)			
			Megalonotus sabulicola		(Thomson, 1870)	Species present in Europe	This European insect feeds on seeds of Centaurea spp. (Asteraceae).	Asteraceae Seeds infesting Insect
		Membracidae	*Stictocephala bisonia*		(Kopp & Yonke, 1977)	Originating in North America, spread throughout southern Europe. In Italy present only in the northern regions.	Several bearing host plants, forest essences, cultivated herbaceous plants and spontaneous herbs.	Insect infesting Plants
		Miridae		*Deraeocoris nebulosus*	(Wheeler et al. 1975)	Insect found in Canada, it has spread to the United States. Common insect in eastern states	This insect is a generalist predator of several insects that feed on plants and mites	Predator Insect
			Deraeocoris lutescens		(Schilling, 1837)	Present in most of Europe	Common predator of caterpillars	Predator Insect
			Deraeocoris sp.		(Kirschbaum, 1856)			
			Lygus sp.		(Hahn, 1833)			
			Lygus pratensis		(Linnaeus, 1758)	Diffusion Olopaleartica. In Italy present on most of the Territory	Insect polyphagous of many herbaceous plants, vegetable and fruit trees	Infesting Insect
			Taylorilygus apicalis		(Fieber, 1861)	Cosmopolitan spread	polyphagous; Asteraceae etc.	Infesting polyphagous Insect
		Pentatomidae	*Aelia acuminata*		(Linnaeus, 1758)	Present throughout Europe	The larvae feed on the seeds to ripen of a series of herbs of the Poaceae	Poaceae Infesting Insect
			Eurydema ventralis		(Kolenati, 1846)	Distribution Palearctic and present in all Italian regions	Insect lives on cultivated and spontaneous cruciferous but also reported on cereals and potato	cruciferous infesting Insect
				Halyomorpha halys	(Stål, 1855)	East Asia origin	H. Halys is a highly polyphagous infesting insect that can cause extensive damage to fruit-growing (especially to Rosaceae) and horticulture (especially Fabaceae).	Exotic invasive Insect
			Rhaphigaster sp.		(Laporte de Castelnau, 1833)			
			Rhaphigaster nebulosa		(Poda, 1761)	In Italy it is common in all regions	Adults and young polyphagous forms, sting the gems and flowers of bearing, hazelnuts and various aerial organs of garden plants	Infesting insect but also predator (Halta lythri and Galerucella luteola)
		Pseudococcidae	*Planococcus citri*		(Risso, 1813)	Diffused in all the temperate and warm areas, common in Italy in Citrus cultivation areas	The cochineal infests mainly citrus	Insect Citrus infesting

Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Rincota	Pseudococcidae	*Pseudococcus viburni*		(Signoret, 1875)	First found in California, it has been reported in South Africa, England, southern France and southern central Italy	It lives on different ornamental plants, on kaki, on Vines (occasionally), citrus and Dianthus sp.. Occasional attacks also on Malus and Pyrus	Tree species weeding
			Pseudococcus longispinus		(Targioni Tozzetti, 1867	Cosmopolitan, Italy is common in the greenhouses	It infests preferably ornamental plants but can also attack citrus, vine, fig and Kako	Weed species
		Rhopalidae	*Chorosoma schillingi*		(Schilling, 1829)	Present in almost all of Europe	Insect nourishing of various herbs	Weed species
		Tingidae	*Corythucha ciliata*		(Say, 1832)	American origin, reported in Italy in the middle years 60 and now common in all regions	He lives mainly on plane trees	Species weeding plane
			Stephanitis pyri		(Fabricius 1775)	Widespread throughout temperate and southern Europe. In Italy present Everywhere	It lives on apple tree, pear, cherry, plum, Piracanta, Rose, rhododendron, Azalea, Sorbus torminalis and Hawthorn.	Species infesting various tree species
	Diptera	Anthomyiidae	*Pegomya sp.*		(Robineau-Desvoidy, 1830)		Some are considered harmful because of their foliar miner larvae	Miner species
		Calliphoridae	*Lucilia sericata*		(Meigen, 1826)	Cosmopolitan	Larva Saprophage	Saprophage species
		Chironomidae	*Cricotopus sp.*		(van der Wulp, 1874)		Larvae live in water or rotting substances	Decomposing species
			Cryptochironomus supplicans		(Meigen, 1830)		Larvae live in water or rotting substances	Decomposing species
			n.i.					
		Chloropidae	*Thaumatomyia notata*		(Meigen, 1830)	Widespread in most of Europe and also in Italy	Adults feed on nectar and sugary substances, larvae predatory terrestrial	Species Predator Terrestrial
		Dolichopodidae	*Chrysotus* n. sp. choricus grp.		(Meigen, 1824)		Family of Predators	Predators
		Drosophilidae	*Drosophila* sp. (prob. *simulans*)		(Fallen, 1823)		Carpophagous	Species infesting Fruits
			Drosophila suzukii		(Matsumura, 1931)	A company originating in the east, found in Italy in Trentino in 2009 now present also in other regions	Carpophagous, pest of small fruits	Species infesting Fruits
		Muscidae	*Coenosia attenuata*		Stein, 1903		Predator	Predator species
		Psychodidae	*Clogmia albipunctata*		(Williston, 1893)		The larvae live in aquatic environments, feeding on organic substances, weeds of domestic discharges	Species feeds on organic substance
			Psychoda alternata		(Say, 1824)	Cosmopolitan	The larvae live in aquatic environments, feeding on organic substances, weeds of domestic discharges	Species feeds on organic substance
			n.i.					
		Pipunctulidae	*Tomosvaryella kuthyi*		Aczel, 1944		The larvae live, as endophagous of species of Cercopidi, leafhoppers and Fulgorii, at maturity escapes from the body of the victim to land in the soil	Endoparasites species
		Sciomyzidae	*Dictya* sp.		(Meigen, 1803)			
			Sepedon sphegea		(Fabricius, 1775)	Europe, North Africa, Asia	marsh flies or snail-killing flies	Species feeds on organic substance or predator of other Sciomyzidae
		Syrphidae	*Eristalinus taeniops*		(Wiedemann, 1818)	Africa, Asia, some European countries (France, Italy, Spain, Portugal, Albany,..	Predatory larvae, adults feed with nectar	Predatory larvae
			Eristalinus sp.		(Rondani, 1845)		Predatory larvae, adults feed with nectar	Predatory larvae
		Stratiomyidae	*Oplodontha viridula*		(Fabricius, 1775)	Europe, North Africa, Asia	Aquatic Larva	Aquatic Larva
		Tephritidae	*Acanthiophilus helianthi*		(Rossi, 1794)	Europe, Africa, Asia	The larva feeds on the flowers of Carthamus tinctorius (safflower) and several other Asteraceae	Miner or saprophagous larvae
			Dioxyna sp. (prob. bidentis)		Frey, 1945	Sweden, Norway	About Asteraceae	
			Euleia sp.		(Walker, 1835)			
			Rhagoletis cerasi		(Linnaeus, 1758)	Fruit fly is present in southern Europe. In Italy is present everywhere.	Cherry Fly	Carpophagous larva

Groups of organisms		Family	Autochthonous/ naturalized	Allochthonous	Author Year	Distribution	Note	Insect type
Insecta	Diptera	Tephritidae	*Tephritis formosa*		(Loew, 1844)	Europe and Near East	Nematodes, on several Asteraceae	Galligena
	Hymenoptera	Aphelinidae	*Aphelinus* sp.		Dalman, 1820	Cosmopolitan genre	Parasitoids family, generally of aphids	Parasitoids
		Apidae	*Apis mellifera*		Linnaeus, 1758			
		Colletidae	*Hylaeus* sp.		Fabricius, 1793	Cosmopolitan genre	They feed on pollen and nectar, solitary, build small nests in cavities	
			Hylaeus leptocephalus		(Morawitz, 1870)	Central Europe	They feed on pollen and nectar, solitary, build small nests in cavities	
		Eulophidae	*Euplectrus* sp.		Westwood, 1832	Cosmopolitan genre	Ectoparasitoids of moth larvae	
		Figitidae	*Alloxysta* sp.		Forster, 1869	Cosmopolitan genre	Hyperparasitoid of aphids (Braconidi and Calcidoidei)	
		Formicidae	*Formica* sp.		Linnaeus, 1758	Europe, North America, Asia		
		Vespidae	*Polistes* sp.		Latreille, 1802	Cosmopolitan genre		
			Vespa crabro		Linnaeus, 1758	Europe, North America, Asia		
			Vespula sp.		Thomson, 1869	Europe, North America, Asia		
		Tenthredinidae	*Athalia rosae rosae*		(Linnaeus, 1758)	The insect is widespread throughout Europe, North America, Asia minor, Africa	Cruciferous parasite	Cruciferous infestation Larva
	Isoptera	Kalotermitidae	*Kalotermes flavicollis*		(Fabricius, 1793)	The insect is widespread in southern Italy and goes back along the coasts to Liguria and Istria.	One of the two species of termites present in Italy, wood of trees and wood in opera	
	Trichoptera	Hydropsychidae	*Hydropsyche modesta*					
			Hydropsyche sp					
		Hydroptilidae	*Agraylea sexmaculata*					
			Hydroptila angulata					
			Hydroptila angulans					
			Hydroptila sp.					
			Hydroptila vectis					
		Leptoceridae	*Mystacides azureus*					
			n.i.					
	Neuroptera	Coniopterygidae	*Semidalis sp. (prob. aleyrodiformis)*					
	Ephemeroptera		*n.i.*					
Aracnidi	Acaridi	Eriophyidae	*Aceria erineus*					

ANNEX 11_ PHOTOGRAPHIC REPORT
TRAPPING

1. Multi Funnel Trap - *Monochamus* sp. trap



2. Multi Funnel Trap - *Anoplophora* sp. trap

3.Sticky prism trap - *Agrilus* sp. prismatic adhesive chromotrap

4. Biogard Rebel Amarillo – Tefritides chromo adhesive trap

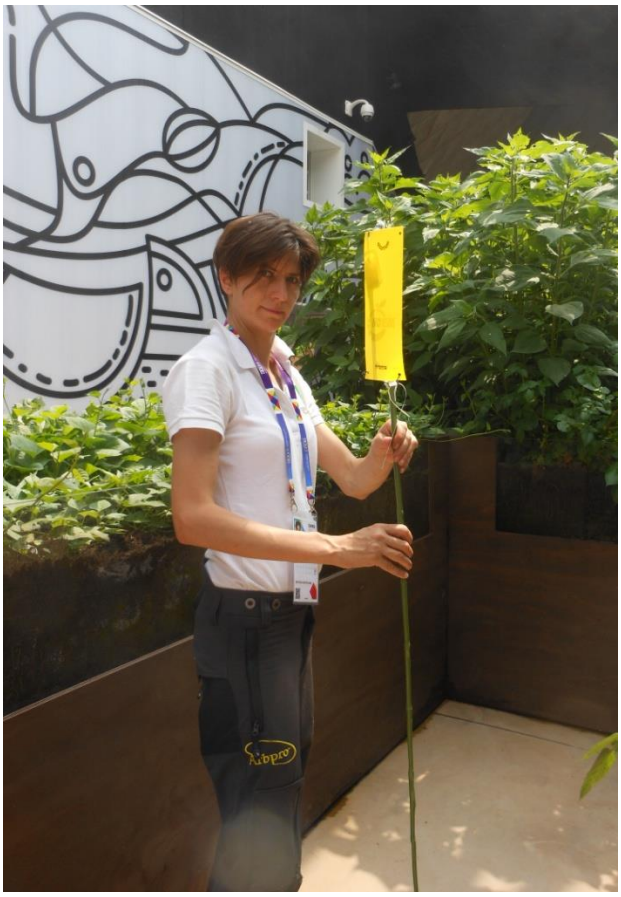
5. Catch Can Trap - *Popillia japonica* double attractive trap

6. Delta Trap – pagoda trap with specific pheromone

7. Positioning *Phytophthora ramorum* apple bait

8. *Phytophthora* apple bait in the Expo ditch

9. Biogard Planotrap - *Planococcus* sp. glue hut



10. Biogard Glutor yellow – Thrips yellow adhesive chromotrap

11. Captaspore control in the Green Expo Perimeter

a. *Hishimonus hamatus* (photo Taddei)

b. *Deraeocoris nebulosus*

c. *Drosophila suzukii*

d. *Acrolepiopsis assectella*

e. *Frankliniella occidentalis*

f. *Eucallipterus tiliae*

g. *Pieris rapae*, crisalide

h. *Plutella xylostella*

i. *Diabrotica virgifera virgifera*

l. *Sigara striata*

m. *Hydroptila vectis*

n. *Omalium caesum*

o. *Psammotettix confinis*

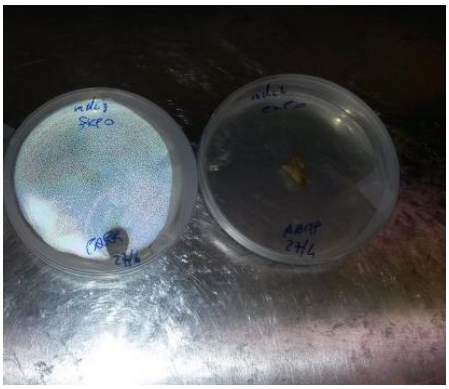
p. *Sitona hispidulus*

q. *Euleia* sp.

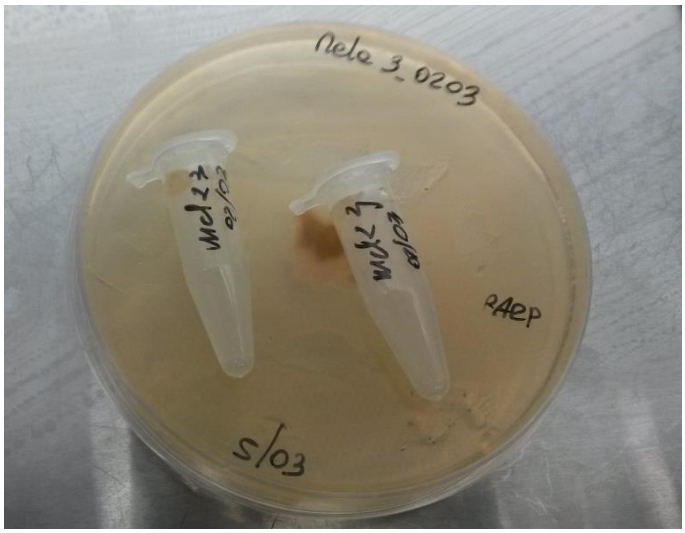
r. *Euleia* sp.

s. *Thaumatomyia notata*

t. *Protapion fulvipes*



1.-2. Photo of oomicities isolating method from apple bait: The apple fragments were incubated on selective substrate Parp agar for the research of oomicities in surface water.



3. Isolation on PARP purification of strain and sequencing led to the identification of *Pythium dissotocum* on apple 3 of 02.03 15_0087.3.

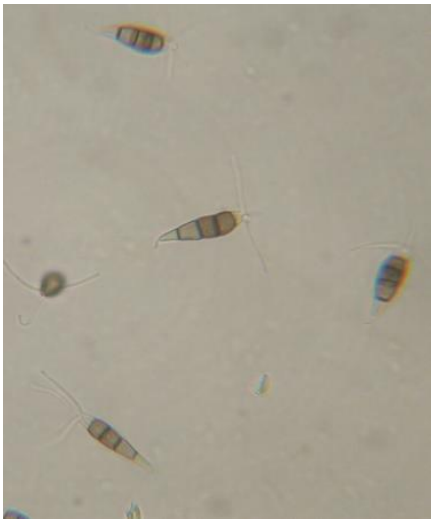
4.*Phythopthora* sp. colony on Parp Agar

5. Preparation of slides: cutting of ribbons coming and collected weekly from the captaspore, fixing on a gel slide and subsequent observation to the optical microscope at 400 X

TYPOLOGIES OF SPORTS OBSERVED ON THE CAPTASPORE RIBBON (OPTICAL MICROSCOPE 400 X)

7. *Epicoccum* sp.- Photo A. Tantardini

8. *Cladosporium* sp. - Photo A. Tantardini



9. *Pestalotiopsis* sp.- Photo A. Tantardini

10. *Alternaria* sp. - Photo A. Tantardini

11. *Curvularia* sp. - Photo A. Tantardini

12. *Drechslera* sp. - Photo A. Tantardini

ANNEX 12_ORGANISMS CAPTURED IN TRAPS

ORGANISMS CAPTURED BY TYPE OF TRAP USED									
Trap type	**Trade names**	**Research Organisms**	**N.° trap.**	**Trap Code**	**Trap Placement**			**Organisms found**	
					Coord. Map	**Zone**	**Plants**	**Orders**	**Species**
Multi-funnel traps with attractiveness	WitaTrap Multi Funnel Trap + Pheromone αβ-Kairomone 1,2,3 - formulation of ChemTica Internacional	*Anoplophora spp.*	10	MfAn1	26H	green area with *Acer campestre*	*Acer campestre*	Coleoptera	*Chlorophorus varius varius*
				MfAn2	28G	green perimeter behind slow food	*Platanus sp.*		
				MfAn3	22D	biodiversity Park	*Carpinus sp.*		
				MfAn4	20B	green perim. behind Tree of Life	*Platanus sp.*	Hymenoptera	*Vespa cabro*
				MfAn5	16F	green perim. behind spices cluster	*Platanus sp.*		
				MfAn6	19J	green perim. behind future food	*Platanus sp.*		
				MfAn7	9J	green perim. Zero pavilion	*Platanus sp.*		
				MfAn8	8E	green perim. behind Angola	*Platanus sp.*	Rhynchota	*Halyomorpha halis*
				MfAn9	14I	green perim. behind dining area 96	*Platanus sp.*		
				MfAn10	23C	green perim. behind arid areas cluster	*Acer platanoides*		
	WitaTrap Multi Funnel Trap with bubble cap formulation of WTB lure from Contech Enterprises Inc.	*Pityophthorus juglandis*	1	MfPi1	19I	Hortus 3 near Mexico pav.	*Juglans regia.*	No organism found	
	WitaTrap Multi Funnel Trap with Galloprotect Pack (1 Pheromone, 3 Kairomone) - SEDQ	*Monochamus spp.*	3	MfMo1	23D	biodiversity Park	*Pinus sp.*	Coleoptera	*Ips sexdentatus*
				MfMo2	14K	Roundabout in the outer perim., behind Eataly	*Pinus sp.*	Hymenoptera	*Apis mellifera*
				MfMo3	E29	AMSA Area			*Polistes sp.* *Vespula sp.*
Purple prism Trap, adhesive on the three sides with attractive	Purple Prism Trap (EAB Trap) with double lure (Manuka oil and Leaf alcool)	*Agrilus spp. (Agrilus planipennis, Agrilus anxius, Agrilus auroguttatus)*	10	PrAg1	H27	green perim. behind biodiversity square/mediterranean hill	*Quercus sp.*	Coleoptera	*Ophraella communa*
				PrAg2	16F	green perim. behind end of Children Park	*Fraxinus sp.*		*Adalia bipunctata*
				PrAg3	25I	green perim. behind McDonald	*Quercus sp.*		*Bruchus brachialis*
				PrAg4	18J	green perim. behind Auditorium	*Fraxinus sp.*		*Harmonia axyridis*
				PrAg5	11I	green perim. behind rice cluster	*Quercus sp.*		*Scymnus sp.*
				PrAg6	6K	green perim. behind zero pav.	*Fraxinus sp.*		*Stethorus punctillum*
				PrAg7	7E	green perim. behind Czech Rep. pav.	*Quercus sp.*	Hymenoptera	*Hylaeus sp.*
				PrAg8	18D	green perim. behind Italy pav.	*Quercus sp.*	Rhynchota	*Deraocoris sp.* *Metcalfa pruinosa*
				PrAg9	22C	green perim. behind Islands cluster	*Quercus sp.*		*Orientus ishidae* *Deraococoris sp.*
				PrAg10	27C	green perimeter near office	*Fraxinus sp.*	Trichoptera	*Hydroptila angulata*
Bucket trap with attractiveness	Biogard Rhynchotrap with RHYFER 220 – BIOGARD/ CBC	*Rhyncophorus spp.*	4	SRh1	23I	Morocco pav.	Palm *Phoenix spp.*	Coleoptera	*Oryctes nasicornis*
				SRh2	14G	Colombia pav.			
				SRh3	8G	Bahrain pav.			
				SRh4	26F	Oman pav.			
Double attractive Traps	Trécé Catch Can Trap - (TBC Japanese Beetle Trap) with Double lure baitpack system from GREAT LAKES IPM - USA	*Popilia japonica*	10	Tfly1	26D	Corner NORD-EST-of fence	Fence	Coleoptera	*Adalia decempunctata*
				Tfly2	26I	Corner SUD-EST-of fence	Fence		*Cetonia aurata*
				Tfly3	(20J) - 21H	green perim., behind Oper Air Theatre - 10/07 moved to Austria pav.	*Fagus sylvatica*		*Diabrotica v. virgifera* *Harpalus rufipes*
				Tfly4	15K	green perim. behind Eataly	*Carpinus sp.*	Hymenoptera	*Formica sp.*
				Tfly5	16F	green perim. behind spices cluster	*Fraxinus sp.*	Rhynchota	*Corythucha ciliata*
				Tfly6	7J	green perim. behind zero pav.	*Quercus sp.*		*Halyomorpha halys*
				Tfly7	12I	green perim. behind cocoa cluster	*Acer campestre*		*Rhaphigaster nebulosa*
				Tfly8	9E	green perim. behind Brazil pav.	*Platanus sp.*		*Rhaphigaster sp.*
				Tfly9	6G	green perim. behind San Paolo	*Carpinus sp.*	Diptera	*Lucilia sericata*
				Tfly10	19B	green perim. behind Tree of Life	*Carpinus sp.*	Lepidoptera	*Paracorsia repandalis*
Delta Trap Pagoda with adhesive bottom with attractiveness	*Koppert Delta trap + Erogatore in gomma con feromone specie-speciFicus sp. PHERODIS - KOPPERT*	*Thaumatotibia leucotreta*	5	PgTh1	26G	Hortus near Oman	*Citrus ps.*	Coleoptera	*Corticarina sp.*
				PgTh2	23F	Hortus near Turkey	*Prunus sp.*	Lepidoptera	*Cadra cautella*
				PgTh3	22H	cereals and tubers cluster	*Amaranthus sp.*		*Grapholita funebrana,* *Grapholita janthinana*
				PgTh4	15G	Fruits and vegetables cluster	*Citrus ps.*		*Monopis imella* *Pammene albuginana*
				PgTh5	11E	green area behind Cascina Triulza	*Punica granatum*		*Tuta absoluta* *Hydroptila angulata*

ORGANISMS CAPTURED BY TYPE OF TRAP USED

Trap type	Trade names	Research Organisms	N.° trap.	Trap Code	Trap Placement			Organisms found	
					Coord. Map	Zone	Plants	Orders	Species
Delta Trap Pagoda with adhesive bottom with attractiveness	*Koppert Delta trap + Erogatore in gomma con feromone specie-speciFicus sp. PHERODIS - KOPPERT*	*Tecia solanivora*	3	PgTe1	22I	cereals and tubers cluster	*Solanum tuberosum*	Lepidoptera	*Clepsis peritana*
				PgTe2	26H	Slow food pav.	*Solanum tuberosum*		*Scrobipalpa ocellatella*
				PgTe3	18H	Hungary pav.	*Solanum tuberosum*		*Heliothis peltigera* *Monopis imella*
Pagoda trap with side openings and adhesive bottom with attractiveness	*Biogard BDT Delta Trap + Erogatore in gomma con feromone speciFicus sp. BIOGARD/ CBC*	*Spodoptera littoralis*	3	PgSp1	22D	biodiversity Park	*Zea mais*	Coleoptera	*Ophraella communa*
				PgSp2	16G	Fruits and vegetables cluster	*Solanum lycopersicum.*	Hymenoptera	*Hylaeus sp.*
				PgSp3	15I	Hortus Eataly (near Liguria)	*Wisteria sp.*	Rhynchota	*Metcalfa pruinosa*
		Keiferia lycopersicella	3	PgKe1	15G	Fruits and vegetables cluster	*Solanum lycopersicum.*	Coleoptera	*Ophraella communa*
				PgKe2	20G	Hortus near Cardo	*Solanum lycopersicum.*		*Propylaea 14-punctata*
				PgKe3	6H	Hortus near building services near Expo centre	*Solanum lycopersicum.*	Diptera	*Psychodidae*
								Lepidoptera	*Tuta Absoluta*
Adhesive trap with roof and adhesive bottom with attractiveness	Traptest Isagro + Isagro pheromone dispenser	*Sesia spp.*	1	Sesia	26E	biodiversity Park	*Malus sp.*	Coleoptera	*Corticarina sp.* *Ophraella communa* *Scymnus sp.*
Red Hut trap with attractiveness	*Biogard Planotrap + Erogatore in gomma con feromone speciFicus sp. BIOGARD/ CBC*	*Planococcus ficus*	1	PlPf1	22D	biodiversity Park	*Ficus sp.*	Diptera	*Psychodidae*
		Planococcus ficus	1	PlPf2	14D	Children park	*Ficus sp.*	Rhynchota	*Laodelphax striatellus* *Macrosteles sp.* *Sigara striata*
		Planococcus citri	1	PlPc1	26F	Hortus front Hill	*Citrus sp.*	No organism found	
Chrome Yellow Cross Glue Traps	Biogard Rebel Amarillo	Tephritidae *(Bactrocera spp.; Ceratitis spp.)*	7	ReATf1	26E	Hortus behind ass. mondiale agronomi	*Prunus avium*	Coleoptera	*Altica oleracea* *Adalia bipuntata* *Chaetocnema tibialis* *Coccinella decempunctata* *Ophraella communa* *Scymnus sp.* *Sitona hispidulus* *Stethorus sp.* *Trixagus meybohmi*
				ReATf2	23F	Hortus near Turkey	*Prunus avium*		
				ReATf3	19H	Hortus near Mexico	*Juglans regia.*	Diptera	*Eristalinus taeniops* *Euleia sp.* *Psychodidae* *Ephemeroptera* *Rhagoletis cerasi*
				ReATf4	13D	Children park	*Prunus avium*	Hymenoptera	*Hylaeus sp.* *Athalia rosae rosae*
								Lepidoptera	*Thaumatomyia notata* *Spodoptera exigua*
				ReATf5	27G	mediterranean hill (High part)	*Olea europaea*	Rhynchota	*Hishimonus hamatus* *Anoecia sp.* *Cicadella viridis* *Dyctiophora europaea* *Euscelidius variegatus* *Macrosteles sp.* *Metcalfa pruinosa* *Orientus ishidae* *Philenus spumarius* *Psammotettix sp. (prob. confinis),* *Scaphoideus titanus* *Tingidae* *Typhlocyba sp.* *Zyginidia pullula*
				ReATf6	19G	Hortus behind Italy pav., near Israel pav. Restaurant	*Prunus avium*		
				ReATf7	7H	UN Garden	*Prunus avium*		
								Trichoptera	*Hydroptila angulans*
Chrome yellow Cross glue traps with attractiveness	Biogard Rebel Amarillo + Erogatore a cilindro con attrattivo ammoniacale BIOGARD/CBC	*Rhagoletis spp.*	1	ReARh1	17J	behind stand Kinder+sport	*Prunus avium*	Coleoptera	*Adalia bipunctata* *Ophraella communa*
								Diptera	*Hydropsyche modesta*
								Rhynchota	*Fieberiella florii*
								Trichoptera	*Hydroptila vectis*
Chrome White Cross Glue Traps	Biogard Rebel Bianco	Beetles *(Byturus spp.);* Hymenoptera *(Hoplocampa spp.)*	8	ReB1	26 G	pic nic area near Oman	*Prunus avium*	Coleoptera	*Ophraella communa*
				ReB2	21 I	green area behind Austria pav.	*Rubus idaeus*	Hymenoptera	*Athalia rosae rosae*
				ReB3	19 F	Hortus behind Italy pav., near pav. Israele	*Prunus sp.*	Lepidoptera	*Aproaerema anthyllidella* *Nomophila noctuella*
				ReB4	18I	Future food district, behind Spain pav.	*Malus sp.*	Rhynchota	*Cicadella viridis*
				ReB5	14I	green perim., behind coffee cluster	*Tilia sp.*		*Cryptochironomus supplicans* *Eristalinus taeniops* *Halyomorpha halys* *Hylaeus leptocephalus* *Laodelphax striatellus* *Lygus pratensis* *Oplodontha viridula*
				ReB6	18J	bath perim., near Food future district	*Malus sp.*		
				ReB7	24I	Flowerbed with mulberry trees, more behind Morocco pav..	*Rubus sp.*		
				ReB8	10F	Flowerbed with Poplars and blackberries near Korea pav./Cascina Triulza restaurant	*Rubus sp.*	Tricoptera	*Mystacides azureus*

ORGANISMS CAPTURED BY TYPE OF TRAP USED

Trap type	Trade names	Research Organisms	N.° trap.	Trap Code	Trap Placement			Organisms found	
					Coord. Map	Zone	Plants	Orders	Species
Bait (apples)	/	*Phythophtora acerina et al.*	6	mela1	21B	green perimeter behind biomediterranean cluster	Apples	Dothideomycetes	*Alternaria alternata* *Colletotrichum acutatum* *Colletotrichum florinae*
				mela2	22C	green perimeter behind biomediterranean cluster		Oomycetes	*Phytophthora lacustris* *Pythium dissotocum* *Pythium litorale*
				mela3	26H	green perimeter behind slow food		Saccharomycetales	*Candida sp.* *Galactomyces sp.* *Geotrichum candidum*
				mela4	16F	green perimeter behind spices cluster, end of children park		Sordariomycetes	*Clonostachys rosea* *Fusarium sp.* *Fusarium tricinctum* *Galactomyces geotrichum* *Mucor sp.* *Trichoderma asperellum* *Trichoderma koningopsis*
				mela5	7K	green perimeter behind zero pav.			
				mela6	18J	phyto bath near Open air theatre			
Captaspore	/	Fungal spores	1	Cap1	21 C	green perimeter, near arid areas cluster.	/	Dothideomycetes	*Alternaria alternata* *Alternaria sp.* *Cercospora sp.* *Cladosporium sp.* *Diplodia sp.* *Drechslera sp.* *Epicoccum sp.*
								Exobasidiomycetes	*Tilletia sp.*
								Lecanoromycetes	*Sphaeropsis sp.*
								Oomycetes	*Plasmopara sp.*
								Sordariomycetes	*Leptosphaeria sp.* *Pestalotiopsis sp.* *Trichothecium roseum*
Chrome Blue glue traps with attractiveness	Biogard Glutor Blu with - LUREM-TR. - KOPPERT	*Thrips spp.*	10	Cromo BTh1	19G	French pav.	*Solanum lycopersicum.*	Coleoptera	*Omalium caesum* *Ophraella communa* *Trechus quadristriatus* *Xyleborinus saxeseni*
				Cromo BTh2	14D	Children park	*Citrus sp.*	Diptera	*Drosophila sp.* *Thaumatomyia notata*
				Cromo BTh3	23H	cereals and tubers cluster	*Chenopodium quinoa.*	Lepidoptera	*Diospyros kuklaemorpha* *Helicoverpa armigera* *Nomophila noctuella* *Pieris sp.* *Sitotroga cerealella* *Tuta absoluta* *Choreutis neRubus sp.na*
				Cromo BTh4	9G	Brazil pav.	Soy/Solanum tuberosum		
				Cromo BTh5	21J	Parking on green perimeter behind Open air theatre	flowerbed of rosa sp. and Acer platanoides near dumpster	Neuroptera	*Semidalis sp. (prob. aleyrodiformis)*
				Cromo BTh6	16K	Parking on green perimeter behind Eataly	flowerbed of *Rosa* sp. and *Acer platanoides* near dumpster	Rhynchota	*Nysius graminicola* *Anoecia sp.* *Athalia rosae rosae* *Cicadella viridis* *Eucallipterus tiliae* *Fieberiella fiorii* *Japananus hyalinus,* *Lygus sp.* *Megalonotus sabulicola* *Rhopalosiphum* *Sepedon sphegea*
				Cromo BTh7	8J	behind zero pav.	grass with dumpsters		
				Cromo BTh8	26I	near Indonesian pav., behind services structure.	grass with dumpsters	Thysanoptera	*Thrips flavus* *Thrips tabaci* *Thrips trehernei* *Aeolothrips intermedius* *Frankliniella intonsa* *Frankliniella occidentalis* *Frankliniella tenuicornis* *Thrips physapus* *Haplothrips leucanthemi* *Tenothrips sp. (prob. frici)*
				Cromo BTh9	14E	flowerbed of UN, behind China pav.	flowerbed of *Populus* sp. near dumpsters		
				Cromo BTh10	24C	flowerbed of smoking pitch behind arid areas cluster	flowerbed of *Punica granatum* near dumpsters	Trichoptera	*Hydropsyche sp.* *Hydroptila vectis*
Chrome Blue Glue Traps	Biogard Glutor Blu	*Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; Agrilus auroguttatus; Thrips palmi.*	15	CromoB1	20F	Hortus	*Malus sp.*	Coleoptera	*Calathus sp.* *Coccotrypes dactyliperda* *Ophraella communa* *Scymnus sp.* *Stethorus sp.* *Trixagus meybohmi* *Ischnopterapion virens*
				CromoB2	20E	Hortus	*Prunus persica*		
				CromoB3	15H	Hortus	*Prunus avium*		
				CromoB4	15I	Hortus	*Malus sp.*	Diptera	*Drosophila sp. (prob. simulans)* *Drosophila suzukii* *Psychodidae*
				CromoB5	27H	mediterranean hill (behind slow food)	*Olea europaea*		
				CromoB6	27H	mediterranean hill (behind slow food)	*Olea europaea*	Lepidoptera	*Autographa gamma* *Spodoptera exigua* *Thaumatomyia notata*
				CromoB7	27G	mediterranean hill	*Olea europaea*	Rhynchota	*Aelia acuminata* *Anoecia sp.* *Chorosoma schillingi*

ORGANISMS CAPTURED BY TYPE OF TRAP USED

Trap type	Trade names	Research Organisms	N.° trap.	Trap Code	Trap Placement			Organisms found	
					Coord. Map	Zone	Plants	Orders	Species
Chrome Blue Glue Traps	Biogard Glutor Blu	*Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; Agrilus auroguttatus; Thrips palmi.*	15	CromoB8	6J	Hortus	*Malus sp.*	Rhynchota	*Cicadella viridis* *Coenosia attenuata* *Cricotopus sp.* *Deraecoris nebulosus* *Dictya sp.*
				CromoB9	8I	Hortus	*Pyrus sp.*		
				CromoB10	19G	Hortus	*Prunus persica*		*Laodelphax striatellus* *Lygus sp.* *Megalonotus sp. (sabulicola o chiragra)* *Metcalfa pruinosa* *Nysius sp.* *Psammotettix sp. (prob. confinis)* *Stephanitis pyri* *Taylorilygus apicalis* *Zyginidia pullula*
				CromoB11	20H	Hortus	*Prunus sp.*		
				CromoB12	23D	biodiversity Park	*Citrus sp.*		
				CromoB13	15F	Fruits and vegetables cluster	*Malus sp.*	Thysanoptera	*Aeolothrips intermedius* *Anaphothrips obscurus* *Frankliniella intonsa* *Frankliniella occidentalis* *Haplothrips aculeatus* *Tenothrips frici* *Thrips hawaiiensis* *Thrips major* *Thrips tabaci* *Thrips trehernei*
				CromoB14	6G	green area	*Malus sp.*		
				CromoB15	6H	green area	*Prunus armeniaca*		
				CromoB16	G27	mediterranean hill (cima)	*Olea europaea*		
				CromoB1 amsa	E29	Amsa area	gate		
				CromoB2 amsa	E29	Amsa area	gate	Trichoptera	*Trichoptera* *Hydroptila angulata* *Hydroptila vectis*
Chrome yellow casting traps with attractiveness	Biogard Glutor Giallo with - LUREM-TR. - KOPPERT	*Thrips spp.*	10	Cromo GTh1	26H	Slow food pav.	*Solanum lycopersicum.*	Coleoptera	*Adalia bipunctata* *Diabrotica v. virgifera* *Ophraella communa* *Trechus quadristriatus*
				Cromo GTh2	15G	Fruits and vegetables cluster	*Solanum lycopersicum*	Diptera	*Psychoda alternata* *Psychodidae* *Thaumatomyia notata*
				Cromo GTh3	23G	Hortus near Turkey	*Prunus persica*	Lepidoptera	*Gypsonoma aceriana* *Nomophila noctuella* *Tuta absoluta*
				Cromo GTh4	20H	Hortus near Martini	*Vitis vinifera*	Rhynchota	*Anoecia sp.* *Aphis craccivora* *Chrysotus n. sp. Choricus grp.* *Cicadella viridis* *Dictya sp.* *Hylaeus sp.* *Lygus pratensis* *Psammotettix sp. (prob. confinis)* *Zygina sp.* *Zyginidia pullula*
				Cromo GTh5	23H	cereals and tubers cluster	*Solanum tuberosum*		
				Cromo GTh6	9F	Brazil pav.	*Abelmoschus esculentus*		
				Cromo GTh7	24J	Parking on green perim. behind Quatar pav.	flowerbed of *Rosa sp.* and *Acer platanoides*		
				Cromo GTh8	10F	green perim. behind Korea pav.	Row of Populus sp. near dumpsters	Thysanoptera	*Aeolothrips intermedius* *Frankliniella intonsa* *Frankliniella occidentalis* *Frankliniella tenuicornis* *Haplothrips aculeatus* *Microcephalothrips abdominalis* *Tenothrips frici* *Thrips tabaci*
				Cromo GTh9	14F	behind Argentina pav. Kitchen	Bush of *Pittosporum* sp. and palms		
				Cromo GTh10	19G	Hortus near Israel pav.	*Solanum lycopersicum*	Trichoptera	*Agraylea sexmaculata* *Hydroptila vectis* *Mystacides azureus*
Chrome yellow traps with double adhesive sheet and attractive	Pherocon Pew Monitoring Trap-two-sided yellow "sticky card" + Pherocon PEW Controlled Release Systems (two component lure) from GREAT LAKES IPM – USA	*Anthonomus eugenii*	8	Cromo GAt1	18H	Hungary pav.	*Capsicum L.*	Coleoptera	*Altica oleracea* *Corticarina sp.* *Cryptophagus pilosus* *Diabrotica virgifera* *Galerucella luteola* *Lyctus africanus* *Ophraella communa*
				Cromo GAt2	16G	spices cluster	*Capsicum L.*		
				Cromo GAt3	8H	Nepal pav.	*Capsicum L.*	Lepidoptera	*Sitotroga cerealella* *Duponchelia fovealis* *Monopis imella* *Sitotroga cerealella*
				Cromo GAt4	26H	Slow food pav.	*Capsicum L.*	Rhynchota	*Alloxysta sp.* *Cicadella viridis* *Corythucha ciliata* *Dictya sp.* *Eurydema ventralis* *Halyomorpha halys* *Hylaeus sp.* *Lygus pratensis* *Nysius graminicola* *Nysius senecionis* *Nysius sp.* *Orius majusculus* *Stictocephala bisonia*
				Cromo GAt5	9G	Brazil pav.	*Capsicum L.*		
				Cromo GAt6	7H	UN Garden	*Capsicum L.*		
				Cromo GAt7	11F	Cascina Triulza Garden	*Capsicum L.*		
				Cromo GAt8	19G	French pav.	*Capsicum L.*		

ORGANISMS CAPTURED BY TYPE OF TRAP USED									
Trap type	Trade names	Research Organisms	N.° trap.	Trap Code	Trap Placement			Organisms found	
					Coord. Map	Zone	Plants	Orders	Species
Chrome Yellow Casting Traps	Biogard Glutor Giallo	*Bactrocera; Ceratitis; Dacus; Rhagoletis; Strauzia longipennis; Toxoptera citricida; Oulema melanopus; Galerucella spp.; Agrilus anxius; Agrilus auroguttatus; Thrips palmi; Diabrotica virgifera; Anthonomus eugenii; Anastrepha spp.; Megacopta cribraria; Trioza eritreae; Bamisia tabaci.*	20	CromoG1	26E	biodiversity Park	*Vitis vinifera*	Coleoptera	*Calathus sp.* *Coccotrypes dactyliperda* *Galerucella luteola* *Harmonia axyridis* *Ophraella communa* *Protapion fulvipes* *Stethorus sp.* *Stegobium paniceum* *Trixagus meybohmi* *Typhaea stercorea* *Xyleborinus saxeseni*
				CromoG2	26E	Hortus near ass. mondiale agronomi	*Citrus sp.*		
				CromoG3	26F	Hortus near Oman	*Prunus sp.*		
				CromoG4	26G	Hortus near Oman	*Wisteria sp.*		
				CromoG5	26G	Hortus near Oman	*Citrus sp.*	Diptera	*Acantothiophilus helianthi* *Clogmia albipunctata* *Dioxyna sp.(prob. bidentis)* *Drosophila sp. (prob. simulans)* *Drosophila suzukii* *Euleia sp.* *Lucilia sericata* *Psychodidae* *Tephritis formosa* *Thaumatomyia notata* *Tomosvaryella kuthyi*
				CromoG6	23E	Hortus near Partner Expo 2	*Prunus sp.*		
				CromoG7	14F	Pergola behind Colombia pav.	*Vitis vinifera*		
				CromoG8	20I	Hortus	*Pyrus sp.*	Hymenoptera	*Aphelinus sp.*
				CromoG9	20G	Hortus	*Prunus persica*	Lepidoptera	*Acrolepsis assectella* *Autographa gamma* *Cadra cautella* *Heliotihs peltigera* *Lycaenidae* *Plutella xylostella* *Spodoptera exigua*
				CromoG10	12F	behind cascina Triulza	*Diospyros kaki*		
				CromoG11	17J	behind stand Kinder+sport	*Prunus sp.*	Rhynchota	*Aelia acuminata* *Allygus atomarius* *Anoecia sp.* *Aphis craccivora* *Chorosoma schillingi* *Cicadella viridis* *Coenosia attenuata* *Cricotopus sp.* *Deraecoris nebulosus* *Dictya sp.* *Empoasca pteridis* *Eutettix sp.* *Fieberiella florii* *Halyomorpha halys* *Japananus hyalinus* *Laodelphax striatellus* *Lygus pratensis* *Lygus sp.* *Megalonotus sp. (sabulicola o chiragra)* *Metcalfa pruinosa* *Nysius sp.* *Orius majusculus* *Psammotettix sp. (prob. confinis)* *Rhopalosiphum padi* *Scymnus sp.* *Siphoninus phillyreae* *Stephanitis pyri* *Taylorilygus apicalis* *Zygina luneris* *Zygina nivea* *Zygina rahmi* *Zygina sp.* *Zyginidia pullula*
				CromoG12	27H	mediterranean hill (behind slow food)	*Olea europaea*		
				CromoG13	27H	mediterranean hill (behind slow food)	*Olea europaea*		
				CromoG14	27G	mediterranean hill	*Olea europaea*		
				CromoG15	6I	Hortus	*Punica granatum*		
				CromoG16	19F	Hortus	*Prunus sp.*		
				CromoG17	19H	Hortus	*Citrus sp.*		
				CromoG18	9G	Save the Children pav.	*Malus sp.*		
				CromoG19	7I	green area behind Fabbrica del Duomo	*Citrus sp.*		
				CromoG20	18I	behind Hungary pav.	*Prunus avium*		
				CromoG21	21D	biomediterranean cluster	*Olea europaea*		
				CromoG22	22D	biomediterranean cluster	*Olea europaea*	Thysanoptera	*Aeolothrips intermedius* *Anaphothrips obscurus* *Frankliniella intonsa* *Frankliniella occidentalis* *Haplothrips aculeatus* *Haplothrips sp.* *Thenothrips frici* *Thrips hawaiiensis* *Thrips major* *Thrips tabaci* *Thrips trehernei*
				CromoG23	21C	biomediterranean cluster	*Olea europaea*		
				CromoG1 amsa	E29	Amsa area	gate		
				CromoG2 amsa	E29	Amsa area	gate	Trichoptera	*Hydroptila angulata* *Hydroptila vectis*















THAILAND